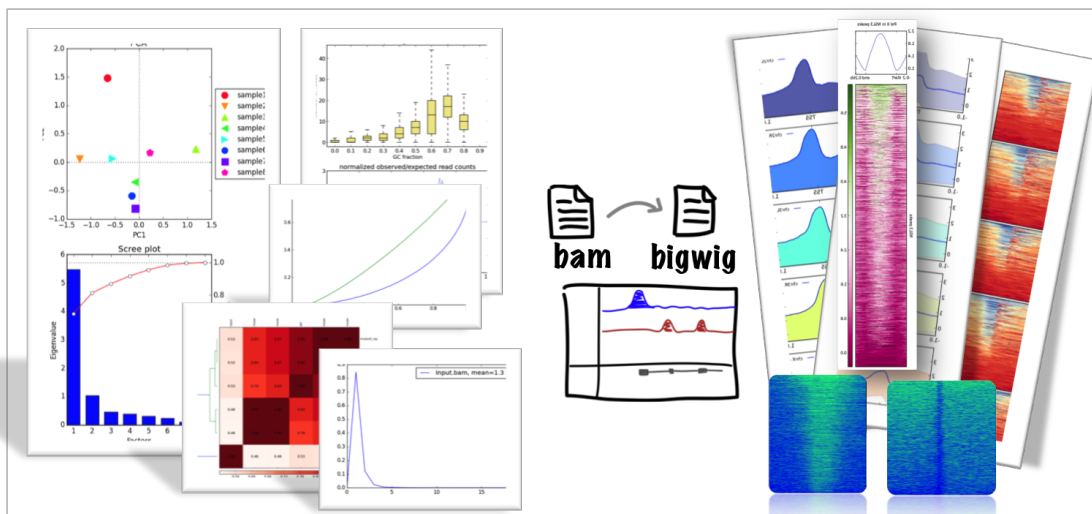

deepTools Documentation

Release 2.1.1

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February 19, 2016

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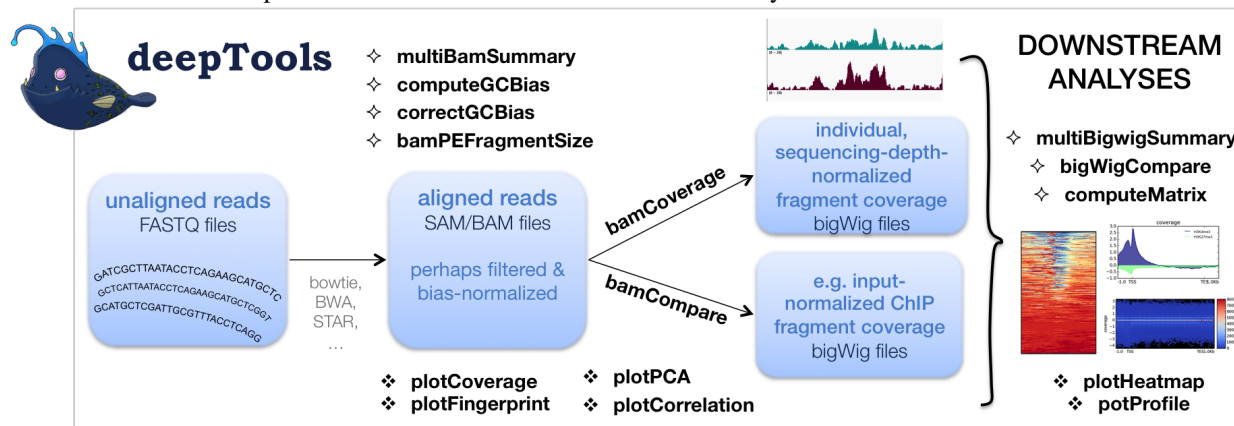
QUALITY CHECKS – FORMAT CONVERSION & NORMALIZATION – PLOTTING

deepTools is a **suite of python tools** particularly developed for the efficient analysis of high-throughput sequencing data, such as ChIP-seq, RNA-seq or MNase-seq.

There are 3 ways for using deepTools:

- **Galaxy usage** – our public [deepTools Galaxy server](#) let's you use the deepTools within the familiar Galaxy framework without the need to master the command line
- **command line usage** – simply download and install the tools (see [Installation](#) and [The tools](#))
- **API** – make use of your favorite deepTools modules in your own python programs (see [deepTools API](#))

The flow chart below depicts the different tool modules that are currently available.



If the file names in the figure mean nothing to you, please make sure to check our [Glossary of NGS terms](#).

Contents:

1.1 Installation

Remember – deepTools are available for **command line usage** as well as for **integration into Galaxy servers!**

- *Requirements*
- *Command line installation using pip*
- *Command line installation without pip*
- *Galaxy installation*
 - *Installation via Galaxy API (recommended)*
 - *Installation via web browser*
 - *Installation with Docker*

1.1.1 Requirements

- Python 2.7
- numpy, scipy, bx-python, and pyBigWig
- pysam >= 0.8

The fastest way to obtain **Python 2.7 together with numpy and scipy** is via the [Anaconda Scientific Python Distribution](#). Just download the version that's suitable for your operating system and follow the directions for its installation. All of the requirements for deepTools can be installed in Anaconda with:

```
$ conda install -c bioconda deeptools
```

1.1.2 Command line installation using pip

Install deepTools using the following command:

```
$ pip install deeptools
```

All python requirements are automatically installed.

If you need to specify a specific path for the installation of the tools, make use of *pip install*'s numerous options:

```
$ pip install --install-option="--prefix=/MyPath/Tools/deepTools2.0" git+https://github.com/fidelram
```

1.1.3 Command line installation without pip

1. Download source code

```
$ git clone https://github.com/fidelram/deepTools.git
```

or if you want a particular release, choose one from <https://github.com/fidelram/deepTools/releases>:

```
$ wget https://github.com/fidelram/deepTools/archive/1.5.12.tar.gz
$ tar -xzf
```

2. The config file will tell you what deepTools expects to be installed properly:

```
$ cat deepTools/deeptools/config/deeptools.cfg

[external_tools]
sort: sort

[general]
# if set to max/2 (no quotes around)
# half the available processors will
# be used
default_proc_number: max/2
test_root: ../deeptools/test/

# temporary dir:
# deepTools bamCoverage, bamCompare and correctGCBias
# write files to a temporary dir before merging them
# and creating a final file. This can be speed up
# by writting to /dev/shm but for this a large
# physical memory of the server is required. If
# this is the case in your system, uncomment
# the following line. Otherwise, setting the
# variable to 'default', deepTools will use the
# temporary file configured in the system.
# Any other path that wants to be used for temporary
# files can by given as well (ie, /tmp)
#tmp_dir: /dev/shm
tmp_dir: default
```

3. install the source code (if you don't have root permission, you can set a specific folder using the --prefix option)

```
$ python setup.py install --prefix /User/Tools/deepTools2.0
```

1.1.4 Galaxy installation

deepTools can be easily integrated into a local [Galaxy](#). All wrappers and dependencies are available in the [Galaxy Tool Shed](#).

Installation via Galaxy API (recommended)

First generate an [API Key](#) for your admin user and run the the installation script:

```
$ python ./scripts/api/install_tool_shed_repositories.py \
    --api YOUR_API_KEY -l http://localhost/ \
    --url http://toolshed.g2.bx.psu.edu/ \
```

```
-o bgruening -r <revision> --name suite_deeptools \
--tool-deps --repository-deps --panel-section-name deepTools
```

The `-r` argument specifies the version of deepTools. You can get the latest revision number from the test tool shed or with the following command:

```
$ hg identify http://toolshed.g2.bx.psu.edu/repos/bgruening/suite_deeptools
```

You can watch the installation status under: Top Panel → Admin → Manage installed tool shed repositories

Installation via web browser

- go to the [admin page](#)
- select *Search and browse tool sheds*
- Galaxy tool shed → Sequence Analysis → deeptools
- install deeptools

Installation with Docker

The deepTools Galaxy instance is also available as a docker container, for those wishing to use the Galaxy framework but who also prefer a virtualized solution. This container is quite simple to install:

```
$ sudo docker pull quay.io/bgruening/galaxy-deeptools
```

To start and otherwise modify this container, please see the instructions on [the docker-galaxy-stable github repository](#). Note that you must use *bgruening/galaxy-deeptools* in place of *bgruening/galaxy-stable* in the examples, as the deepTools Galaxy container is built on top of the galaxy-stable container.

Tip: For support, questions, or feature requests contact: deeptools@googlegroups.com

deepTools Galaxy.	code @ github.
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1.2 The tools

Note: With the release of deepTools 2.0, we renamed a couple of tools:

- **heatmapper** to `tools/plotHeatmap`
- **profiler** to `tools/plotProfile`
- **bamCorrelate** to `tools/multiBamSummary`
- **bigwigCorrelate** to `tools/multiBigwigSummary`
- **bamFingerprint** to `tools/plotFingerprint`.

For more, see [Changes in deepTools2.0](#).

- *General principles*
 - *Parameters to decrease the run time*
 - *Filtering BAMs while processing*
- *Tools for BAM and bigWig file processing*
 - *tools/multiBamSummary*
 - *tools/multiBigwigSummary*
 - *tools/correctGCBias*
 - *tools/bamCoverage*
 - *tools/bamCompare*
 - *tools/bigwigCompare*
 - *tools/computeMatrix*
- *Tools for QC*
 - *tools/plotCorrelation*
 - *tools/plotPCA*
 - *tools/plotFingerprint*
 - *tools/bamPEFragmentSize*
 - *tools/computeGCBias*
 - *tools/plotCoverage*
- *Heatmaps and summary plots*
 - *tools/plotHeatmap*
 - *tools/plotProfile*

tool	type	input files	main output file(s)	application
tools/multiBamSummary	data integration	2 or more BAM	interval-based table of values	perform cross-sample analyses of read counts → plotCorrelation, plotPCA
tools/multiBigwigSummary	data integration	2 or more bigWig	interval-based table of values	perform cross-sample analyses of genome-wide scores → plotCorrelation, plotPCA
tools/plotCorrelation	visualization	bam/multiBigwigSummary output	clustered heatmap	visualize the Pearson/Spearman correlation
tools/plotPCA	visualization	bam/multiBigwigSummary output	2 PCA plots	visualize the principal component analysis
tools/plotFingerPrint	QC	2 BAM	1 diagnostic plot	assess enrichment strength of a ChIP sample
tools/computeGCbias	QC	1 BAM	2 diagnostic plots	calculate the exp. and obs. GC distribution of reads
tools/correctGCbias	QC	1 BAM, output from computeGCbias	1 GC-corrected BAM	obtain a BAM file with reads distributed according to the genome's GC content
tools/bamCoverage	normalization	BAM	bedGraph or bigWig	obtain the normalized read coverage of a single BAM file
tools/bamCompare	normalization	2 BAM	bedGraph or bigWig	normalize 2 files to each other (e.g. log2ratio, difference)
tools/computeMatrix	data integration	1 or more bigWig, 1 or more BED	zipped file for plotHeatmap or plotProfile	compute the values needed for heatmaps and summary plots
tools/plotHeatmap	visualization	computeMatrix output	heatmap of read coverages	visualize the read coverages for genomic regions
tools/plotProfile	visualization	computeMatrix output	summary plot ("meta-profile")	visualize the average read coverages over a group of genomic regions
tools/plotCoverage	visualization	1 or more bam	2 diagnostic plots	visualize the average read coverages over sampled genomic positions
tools/bamPEFragmentSize	information	1 BAM	text with paired-end fragment length	obtain the average fragment length from paired ends

1.2.1 General principles

A typical deepTools command could look like this:

```
$ bamCoverage --bam myAlignedReads.bam \
--outFileName myCoverageFile.bigWig \
--outFileFormat bigwig \
--fragmentLength 200 \
--ignoreDuplicates \
--scaleFactor 0.5
```

You can always see all available command-line options via `--help`:

```
$ bamCoverage --help
```

- Output format of plots should be indicated by the file ending, e.g. `MyPlot.pdf` will return a pdf file,

MyPlot.png a png-file

- All tools that produce plots can also output the underlying data - this can be useful in cases where you don't like the deepTools visualization, as you can then use the data matrices produced by deepTools with your favorite plotting tool, such as R
- The vast majority of command line options are also available in Galaxy (in a few cases with minor changes to their naming).

Parameters to decrease the run time

- **numberOfProcessors** - Number of processors to be used

For example, setting `--numberOfProcessors 10` will split up the workload internally into 10 chunks, which will be processed in parallel.

- **region** - Process only a single genomic region. This is particularly useful when you're still trying to figure out the best parameter setting. You can focus on a certain genomic region by setting, e.g., `--region chr2` or `--region chr2:100000-200000`

These parameters are optional and available throughout almost all deepTools.

Filtering BAMs while processing

Several deepTools modules allow for efficient processing of BAM files, e.g. `bamCoverage` and `bamCompare`. We offer several ways to filter those BAM files on the fly so that you don't need to pre-process them using other tools such as `samtools`

- **ignoreDuplicates** Reads with the same orientation and start position will be considered only once. If reads are paired, the mate is also evaluated
- **minMappingQuality** Only reads with a mapping quality score of at least this are considered
- **samFlagInclude** Include reads based on the SAM flag, e.g. `--samFlagInclude 64` gets reads that are first in a pair. For translating SAM flags into English, go to: <https://broadinstitute.github.io/picard/explain-flags.html>
- **samFlagExclude** Exclude reads based on the SAM flags - see previous explanation.

These parameters are optional and available throughout deepTools.

Warning: If you know that your files will be strongly affected by the filtering of duplicates or reads of low quality then consider removing those reads *before* using `bamCoverage` or `bamCompare`, as the filtering by deepTools is done *after* the scaling factors are calculated!

1.2.2 Tools for BAM and bigWig file processing

`tools/multiBamSummary`

`tools/multiBigwigSummary`

`tools/correctGCBias`

`tools/bamCoverage`

`tools/bamCompare`

`tools/bigwigCompare`

`tools/computeMatrix`

1.2.3 Tools for QC

`tools/plotCorrelation`

`tools/plotPCA`

`tools/plotFingerprint`

`tools/bamPEFragmentSize`

`tools/computeGCBias`

`tools/plotCoverage`

1.2.4 Heatmaps and summary plots

`tools/plotHeatmap`

`tools/plotProfile`

deepTools Galaxy.	code @ github.
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1.3 Example usage

- *How we use deepTools for ChIP-seq analyses*

1.3.1 Step-by-step protocols

This section should give you an overview of how to do many common tasks. We're using **screenshots from Galaxy** here. If you're using the command-line version you can easily follow the given examples since the vast majority of parameters is either indicated in Galaxy, too. Otherwise, just type the program name and the help option (e.g.

`/deepTools/bin/bamCoverage --help`), which will show you all the parameters and options available. Alternatively, you can follow the respective link to the tool documentation here on [readthedocs](#).

Note: Do let us know if you spot things that are missing or should be explained better! Just send an email to deeptools@googlegroups.com.

All protocols assume that you have uploaded your files into a Galaxy instance with a deepTools installation, e.g., [deepTools Galaxy](#). If you need help to get started with Galaxy in general, e.g. to upload your data, see [Using deepTools within Galaxy](#) and [Data import into Galaxy](#).

Tip: If you would like to try out the protocols with **sample data**, go to [deepTools Galaxy](#) → “Shared Data” → “Data Libraries” → “deepTools Test Files”. Simply select BED/BAM/bigWig files and click, “to History”. You can also download the test data sets to your computer by clicking “Download” at the top.

How to do...?

- *QC and data processing*
 - *I have downloaded/received a BAM file - how do I generate a file I can look at in a genome browser?*
 - *How can I assess the reproducibility of my sequencing replicates?*
 - *How do I know whether my sample is GC biased? And if it is, how do I correct for it?*
 - *How do I get an input-normalized ChIP-seq coverage file?*
 - *How can I compare the ChIP strength for different ChIP experiments?*
- *Heatmaps and summary plots*
 - *How do I get a (clustered) heatmap of sequencing-depth-normalized read coverages around the transcription start site of all genes?*
 - *How can I compare the average signal for X-specific and autosomal genes for 2 or more different sequencing experiments?*
 - * *How to obtain a BED file for X chromosomal and autosomal genes each*
 - * *Compute the average values for X and autosomal genes*

QC and data processing

I have downloaded/received a BAM file - how do I generate a file I can look at in a genome browser?

- tool: `tools/bamCoverage`
- input: your [BAM](#) file with aligned reads

Of course, you could also look at your BAM file in the genome browser. However, generating a bigWig file of read coverages will not drastically reduce the size of the file, it also allows you to normalize the coverage to 1x sequencing depth, which makes a visual comparison of multiple files more feasible.

bamCoverage – input and parameters

output

compare the size to the BAM file's size!

can be downloaded and easily uploaded into IGV browser

How can I assess the reproducibility of my sequencing replicates?

Typically, you're going to be interested in the correlation of the read coverages for different replicates and different samples. What you want to see is that replicates should correlate better than non-replicates. The [ENCODE consortium](#) recommends that *for messenger RNA, (...) biological replicates [should] display 0.9 correlation for transcripts/features*. For more information about correlation calculations, see the background description for `tools/plotCorrelation`.

- `tools: tools/multiBamSummary` followed by `tools/plotCorrelation`
- **input: BAM files**
 - you can compare as many samples as you want, though the more you use the longer the computation will take

multiBamSummary calculates average read coverages for a list of two or more BAM files (Galaxy Version 2.0.1.0)

Bam file

7: Input.bam

6: H3K9Me3.bam

5: H3K4Me3.bam

4: H3K4Me1.bam

3: H3K27Me3.bam

The BAM file must be sorted. (--bamfiles)

Choose computation mode

Bins

In the bins mode, the coverage is computed for equally sized bins. In the BED mode, a list of genomic regions in BED format has to be given. For each region in the BED file the number of overlapping reads is counted in each of the BAM files.

Bin size in bp

10000

Length in bases of the window used to sample the genome. (--binSize)

Distance between bins

0

By default, multiBamSummary considers consecutive bins of the specified 'Bin size'. However, to reduce the computation time, a larger distance between bins can be given. Larger distances result in fewer bins being considered. (--distanceBetweenBins)

Region of the genome to limit the operation to

19

This is useful when testing parameters to reduce the time to finish. The format is chr:start-end, for example chr10:456700:891000. (--chr10:456700:891000)

Show advanced options

no

do explore those! e.g. for ignoring duplicates

Save raw counts (coverages) to file

Yes No

if you would like a tabular version of the output

Execute

select as many BAM files as you like (hold cmd + click)

you could also use a file with regions of interest here

default output

14: multiBamSummary

on data 6, data 5, and others: correlation matrix

57.4 KB

format: deeptools_coverage_matrix, database: hg19

Number of bins found: 5911

Compressed binary file

must be used with either plotCorrelation or plotPCA

optional output

15: multiBamSummary

on data 6, data 5, and others: bin counts

5,911 lines, 1 comments

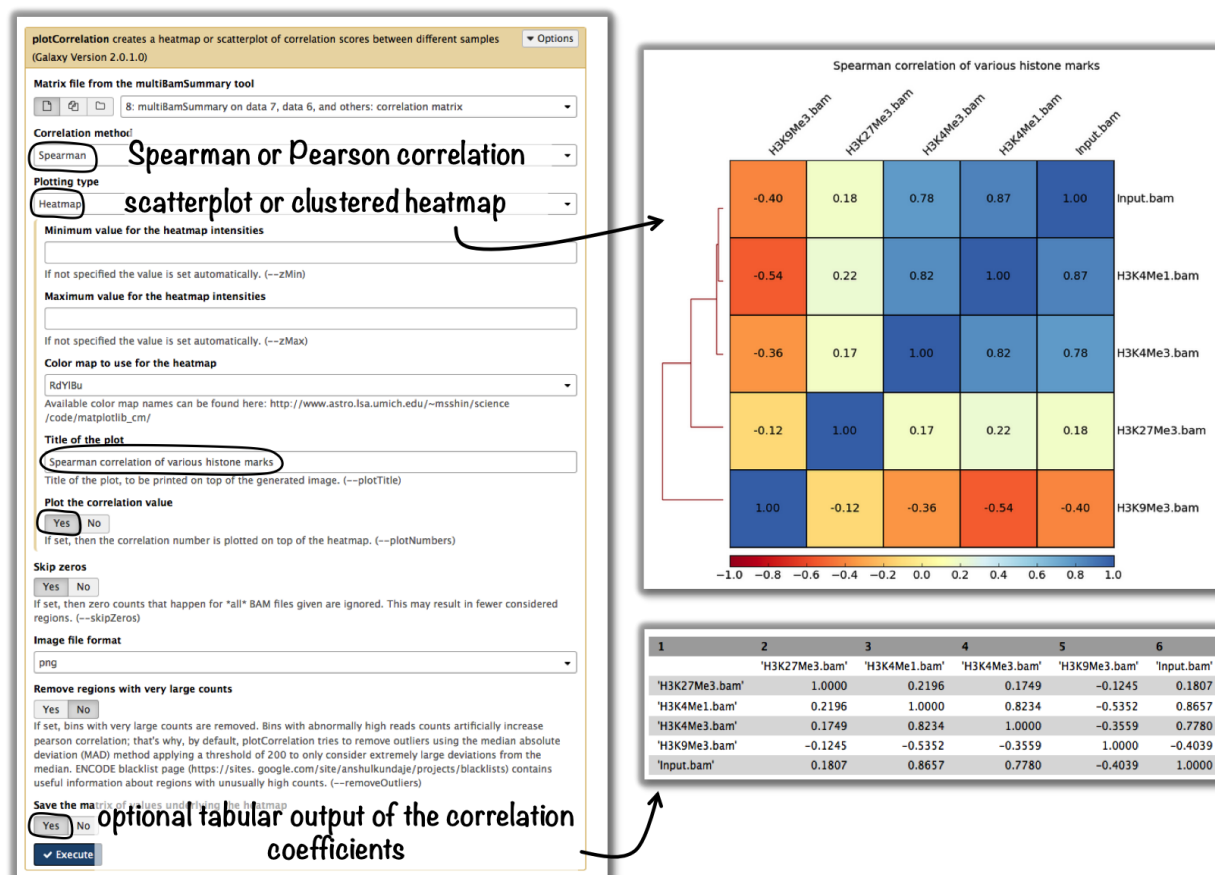
format: tabular, database: hg19

Number of bins found: 5911

1	2	3	4	5
#'chr'	'start'	'end'	'H3K27Me3.bam'	'H'
19	10000	20000	0.0	0.
19	20000	30000	0.0	0.
19	30000	40000	0.0	0.
19	40000	50000	0.0	0.
19	50000	60000	0.0	0.

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Chapter 1. Contents:



Tip: If you would like to do a similar analysis based on bigWig files, use the tool multiBigwigSummary instead.

How do I know whether my sample is GC biased? And if it is, how do I correct for it?

- input: *BAM* file
- use the tool `tools/computeGCBias` on that BAM file (default settings, just **make sure your reference genome and genome size are matching**)

computeGCBias – input and parameters

computeGCBias (version 1.0.2)

BAM file:
 select BAM file
The BAM file must be sorted.

Reference genome:

Using reference genome:
 select correct reference genome
If your genome of interest is not listed, contact the Galaxy team

Effective genome size:

The effective genome size is the portion of the genome that is mappable. Large fractions of NNNN that should be discarded. Also, if repetitive regions were not included in the mapping size needs to be adjusted accordingly. See Table 2 of <http://www.plosone.org/article/info%3Adoi%2F10.1371%2Fjournal.pone.0030377> or <http://www.nature.com/nbt/journal/v> for several effective genome sizes.

Fragment length used for the sequencing:
 indicate fragment length
If paired-end reads are used, the fragment length is computed from the BAM file.

Show advanced options:

GC bias plot:

If given, a diagnostic image summarizing the GC bias found on the sample will be created.

output

S9: computeGCBias GC-bias Plot
20.7 KB
format: pdf, database: hg19
reads_per_bp: 0.00651585140051 genome_size: 3137161264 total_reads: 15976607 filter_out: None 2bit: /data2/galaxy/data/hg19.2bit extra_sampling_file: None min_reads: 0.0 bam: local_baminput.bam max_reads: 27.0 computing frequencies

pdf file/plots

Image in pdf format

S8: computeGCBias on data 44
301 lines
format: tabular, database: hg19
reads_per_bp: 0.00651585140051 genome_size: 3137161264 total_reads: 15976607 filter_out: None 2bit: /data2/galaxy/data/hg19.2bit extra_sampling_file: None min_reads: 0.0 bam: local_baminput.bam max_reads: 27.0 computing frequencies

can be used with correctGCBias

- have a look at the image that is produced and compare it to the examples here
- if your sample shows an almost linear increase in exp/obs coverage (on the log scale of the lower plot), then you should consider correcting the GC bias - *if* you think that the biological interpretation of this data would otherwise be compromised (e.g. by comparing it to another sample that does not have an inherent GC bias)
 - the GC bias can be corrected with the tool `tools/correctGCBias` using the second output of the `computeGCBias` tool that you had to run anyway

correctGCBias – input and parameters

correctGCBias (version 1.0.2)

Output of computeGCBias:

BAM file:

This should be same file that was used for computeGCBias. The BAM file must be sorted.

Reference genome:

Using reference genome:

If your genome of interest is not listed, contact the Galaxy team

Effective genome size:

The effective genome size is the portion of the genome that is mappable. Large fractions of the genome that should be discarded. Also, if repetitive regions were not included in the mapping of reads, then to be adjusted accordingly. See Table 2 of <http://www.plosone.org/article/info%3Adoi%2F10.1371/journal.pone.0151811> for several effective genome sizes.

File format of the output:

Show advanced options:

output

103: correctGCBias on data 44 and data 58

696.0 MB
format: bam, database: hg19
applying correction genome partition size for multiprocessing: 61388754 using region None Sam for chr1:0-61388754 MainProcess, processing 415589 (6445.2 per sec) reads @ chr1:0-61388754 duplicated reads removed 19848 of 415589 (4.78) /dev/shm/tmpB1uXC

display at UCSC [main test](#)
display at Ensembl [Current](#)
display with IGV [web current](#) [local](#)
display in IGB [Local Web](#)

Binary bam alignments file

Warning: correctGCBias will add reads to otherwise depleted regions (typically GC-poor regions), that means that you should **not** remove duplicates in any downstream analyses based on the GC-corrected BAM file. We therefore recommend removing duplicates before doing the correction so that only those duplicate reads are kept that were produced by the GC correction procedure.

How do I get an input-normalized ChIP-seq coverage file?

- input: you need two BAM files, one for the input and one for the ChIP-seq experiment
- tool: tools/bamCompare with ChIP = treatment, input = control sample

bamCompare – input and parameters

bamCompare (version 1.0.2)

Treatment BAM file:
 my "treatment" sample (ChIP sample in this case)

The BAM file must be sorted.

BAM file:
 my control sample

The BAM file must be sorted.

Length of the average fragment size:

Reads will be extended to match this length unless they are paired-end, in which case they will be extended to match the fragment length. If this value is set to the read length or smaller, the read will not be extended. *Warning* the fragment length affects the normalization to 1x (see "normalize coverage to 1x"). The formula to normalize using the sequencing depth is genomeSize/(number of mapped reads * fragment length). *NOTE*: If the BAM files contain mated and unmated paired-end reads, unmated reads will be extended to match the fragment length.

Bin size in bp:
 the smaller the bin, the bigger the output file

The genome will be divided in bins (also called tiles) of the specified length. For each bin the overlapping number of fragments (or reads) will be reported. If only half a fragment overlaps, this fraction will be reported.

Method to use for scaling the largest sample to the smallest:
 to account for differences in sequencing depth between the 2 samples - choose your favorite option

read count

How to compare the two files:

compute log2 of the number of reads ratio

Coverage file format:

bigwig

Show advanced options:
☒ *many different options possible*

yes

Smooth values using the following length (in bp):

The smooth length defines a window, larger than the bin size, to average the number of reads. For example, if the bin size is set to 20 bp and the smooth length is set to 60 bp, then, for each bin size the average of it and its left and right neighbors is considered. Any value smaller than the bin size will be ignored and no smoothing will be applied.

Region of the genome to limit the operation to:
 just for us, to test the tool and save computation time; leave empty if you want the whole genome

chr2

Do not extend paired ends:
☐

If set, reads are not extended to match the fragment length reported in the BAM file, instead they will be extended to match the fragment length. Default is to extend the reads if paired end information is available.

Ignore duplicates:
☒ *define which reads should be included for the read count*

If set, reads that have the same orientation and start position will be considered only once. If reads are paired, the mate position also has to coincide to ignore a read.

Minimum mapping quality (e.g. BOWTIE2 measures):

If set, only reads that have a mapping quality score higher than the given value are considered. *Note* Bowtie's Mapping quality is related to uniqueness: the higher the score, the more unique is a read. A mapping quality defined by Bowtie of 10 or less indicates that there is at least a 1 in 10 chance that the read truly originated elsewhere.

Treat missing data as zero:
☒

This parameter determines if missing data should be treated as zeros. If unchecked, missing data will be ignored and not included in the output file. Missing data is defined as those regions for which both BAM files have 0 reads.

output

should give meaningful name, e.g. "log2ratio_ChIP_input_H3K27ac.bw"

69: bamCompare on data 44 and data 42

18.3 MB

format: bigwig, database: hg19

The scaling factors are: [1. 0.60636248]

[display at UCSC main test](#)

[display in IGB Local Web](#)

Binary UCSC Bigwig file

can be uploaded to UCSC or IGV browsers

How can I compare the ChIP strength for different ChIP experiments?

- tool: tools/plotFingerprint
- input: as many BAM files of ChIP-seq samples as you'd like to compare (it is helpful to include the input control to see what a hopefully non-enriched sample looks like)

1.3. Example usage

1.3. Example usage

1.3. Example usage

1.3. Example usage

- ### 1.3. Example usage

download gene lists

Table Browser

Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and to retrieve DNA sequence covered by a track. For help in using this application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) for general information and sample queries, and the OpenHelix Table Browser [tutorial](#) for a narrated presentation of the software features and usage. For more complex queries, you may want to use [Galaxy](#) or our [public MySQL server](#). To examine the biological function of your set through annotation enrichments, send the data to [GREAT](#). Refer to the [Credits](#) page for the list of contributors and usage restrictions associated with these data. All tables can be downloaded in their entirety from the [Sequence and Annotation Downloads](#) page.

clade: Mammal genome: Human assembly: Feb. 2009 (GRCh37/hg19)

group: Genes and Gene Prediction Tracks track: RefSeq Genes

track hubs

table: refGene describe table schema

region: ☒ genome ☐ ENCODE Pilot regions ☐ position chr21:33031597-33041570 lookup

define regions

identifiers (names/accessions): paste list upload list

filter: create

intersection: create

correlation: create

output format: all fields from selected table Send output to ☒ Galaxy ☐ GREAT

output file: (leave blank to keep output in browser)

file type returned: ☒ plain text ☐ gzip compressed

get output summary/statistics

To reset all user cart settings (including custom tracks), [click here](#).

output

72: UCSC Main on Human: refGene (genome)

48,120 regions

format: bed, database: hg19

display at UCSC [main test](#)

display in IGB [Local Web](#)

display at Ensembl [Current](#)

display at RViewer [main](#)

bed-file of genes

1. Chrom	2. Start	3. End	4. Name	5. 6. Strand	7	8
chr1	66555024	67210768	IM_022251	0 +	67000041	672000
chr1	1611252	1651552	155122	155606	205153	206516
chr1	40550526	50405626	IM_022705	0 -	40550444	504056
chr1	16767166	16706504	IM_010050	0 +	16767256	167056
chr1	22546713	22505555	IM_052550	0 +	22547050	225055
chr1	16767166	16706504	IM_001145270	0 +	16767256	167056

- use tools/computeMatrix with the bigWig file and the BED file
- indicate reference-point (and whatever other option you would like to tune, see screenshot below)

computeMatrix – input and parameters

regions to plot

regions to plot 1

Regions to plot:

72: UCSC Main on Human: refGene (genome) *genes we previously obtained through UCSC*

Label:

Genes

Score file:

105: bamCoverage on data 37 *seq: depth-normalized file from previous step*

computeMatrix has two main output options:

reference-point

beginning of region (e.g. TSS)

The reference point for the plotting:

beginning of region (e.g. TSS)

Discard any values after the region end:

☐

Distance upstream of the start site of the regions defined in the region file:

2000 *totally up to you*

Distance downstream of the end site of the given regions:

4000

Show advanced output settings:

no

Show advanced options:

no

Execute

output

106: computeMatrix on data 72 and data 105: Matrix

647.9 KB

format: bgzip, database: hg19

Warning: 94.16% of regions are not associated to a score in the given BAM file. Check that the chromosome names from the BED file are consistent with the chromosome names in the given BAM file and that both files refer to the same species

warning is raised because the bamCoverage we used contained reads for chr2 only, while the gene file contained all genes

- use the output from `tools/computeMatrix` with `tools/plotHeatmap`
 - if you would like to cluster the signals, choose k-means clustering (last option of “advanced options”) with a reasonable number of clusters (usually between 2 to 7)

heatmapper – input and parameters

heatmapper (version 1.0.2)

Matrix file from the computeMatrix tool:

106: computeMatrix on data 72 and data 105: Matrix

Show advanced output settings:

no

Show advanced options:

yes

Sort regions:

descending order

Whether the heatmap should present the regions sorted. The default is to sort in descending order based on the mean value per region.

Method used for sorting:

mean

For each row the method is computed.

Type of statistic that should be plotted in the summary image above the heatmap:

mean

Missing data color:

black

If 'black' is not set, such cases will be colored in black by default. By using this parameter a different color can be set. A value between 0 and 1 will be used for a gray scale (black is 0). Also color names can be used, see a list here: http://packages.python.org/ete2/reference/reference_sv_colors.html. Alternatively colors can be specified using the #rrggbb notation.

Color map to use for the heatmap:

winter reversed

Available color map names can be found here: http://www.astro.lsa.umich.edu/~msshin/science/code/matplotlib_cm/

Minimum value for the heatmap intensities. Leave empty for automatic values:

Maximum value for the heatmap intensities. Leave empty for automatic values:

Minimum value for the Y-axis of the summary plot. Leave empty for automatic values:

Maximum value for Y-axis of the summary plot. Leave empty for automatic values:

Description for the x-axis label:

distance from TSS (bp)

Description for the y-axis label for the top panel:

genes

Heatmap width in cm:

7.5

The minimum value is 1 and the maximum is 100.

Heatmap height in cm:

15.0

The minimum value is 3 and the maximum is 100.

What to show:

summary plot, heatmap and colorbar

The default is to include a summary or profile plot on top of the heatmap and a heatmap colorbar.

Label for the region start:

TSS

[only for scale-regions mode] Label shown in the plot for the start of the region. Default is TSS (transcription start site), but could be changed to anything, e.g. "peak start".

Label for the region end:

TES

[only for scale-regions mode] Label shown in the plot for the region end. Default is TES (transcription end site).

Reference point label:

TSS

[only for scale-regions mode] Label shown in the plot for the reference-point. Default is the same as the reference point selected (e.g. TSS), but could be anything, e.g. "peak start" etc.

Labels for the regions plotted in the heatmap:

genes

If more than one region is being plotted a list of labels separated by comma and limited by quotes, is required. For example, "label1, label2".

Title of the plot:

My clustered heatmap

Title of the plot, to be printed on top of the generated image. Leave blank for no title.

Do one plot per group:

☐

When the region file contains groups separated by "#", the default is to plot the averages for the distinct plots in one plot. If this option is set, each group will get its own plot, stacked on top of each other.

Did you used multiple regions in ComputeMatrix?:

No, I used only one region.

That option is only relevant if you want to cluster the results. Clustering is only available with one selected region in ComputeMatrix.

Clustering algorithm:

Kmeans clustering

Number of clusters to compute:

3

When this option is set, then the matrix is split into clusters using the kmeans algorithm. Only works for data that is not grouped, otherwise only the first group will be clustered. If more specific clustering methods are required it is advisable to save the underlying matrix and run the clustering using other software. The plotting of the clustering may fail (Error: Segmentation fault) if a cluster has very few members compared to the total number or regions. (default: None).

Buttons:

output

117: heatmapper image

232.4 KB

format: png, database: hg19




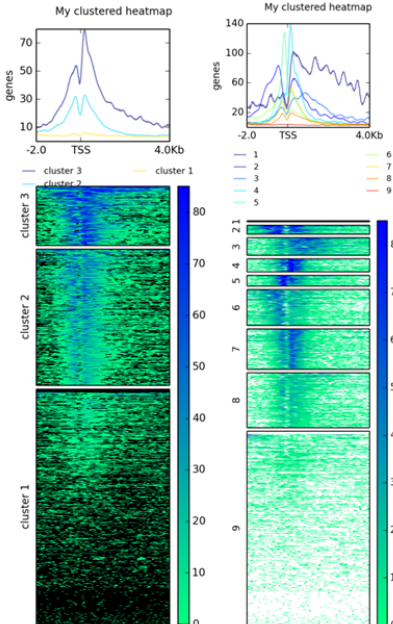
  

Image in png format



these 2 heatmaps were generated using the same computeMatrix output! only two entries differed in heatmapper between these two plots:

- a) color for missing data (black vs. white)
- b) 3 vs. 9 clusters

note how the clustering can group genes with down- and upstream enrichments in addition to strong and weak signals

clustering will overwrite any user-specified groups of regions which is why we recommend to use it only for cases where you supplied just one BED file to computeMatrix

How can I compare the average signal for X-specific and autosomal genes for 2 or more different sequencing experiments?

Make sure you're familiar with `computeMatrix` and `plotProfile` before using this protocol.

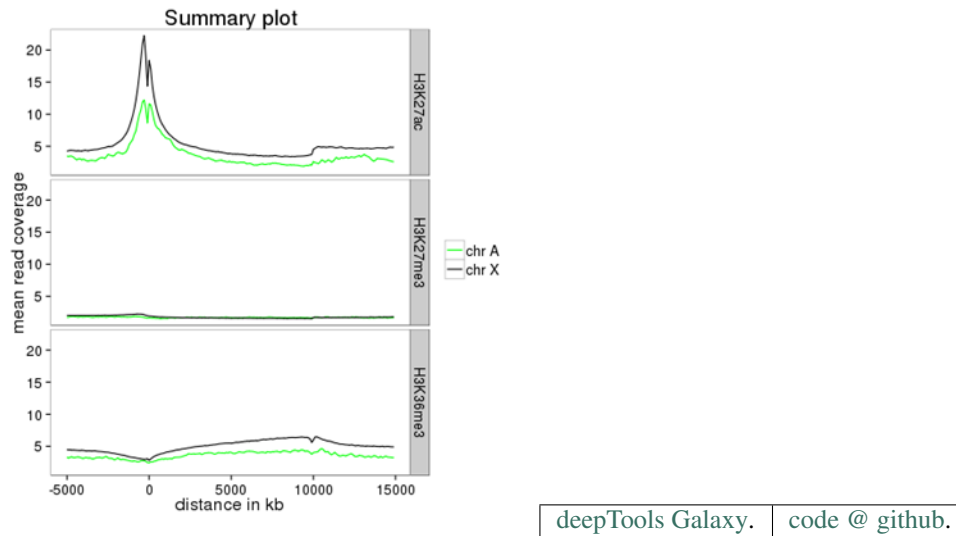
- **tools:**
 - Filter data on any column using simple expressions
 - `computeMatrix`
 - `plotProfile`
 - (plotting the summary plots for multiple samples)
- **inputs:**
 - several bigWig files (one for each sequencing experiment you would like to compare)
 - two BED files, one with X-chromosomal and one with autosomal genes

How to obtain a BED file for X chromosomal and autosomal genes each

1. download a full list of genes via “Get Data” -> “UCSC main table browser” -> group: “Genes and Gene Predictions” -> tracks: (e.g.) “RefSeqGenes” -> send to Galaxy
2. filter the list twice using the tool **“Filter data on any column using simple expressions”**
 - first use the expression: `c1==chrX` to filter the list of all genes -> this will generate a list of X-linked genes
 - then re-run the filtering, now with `c1!=chrX`, which will generate a list of genes that do not belong to chromosome X (!= indicates “not matching”)

Compute the average values for X and autosomal genes

- use `tools/computeMatrix` for all of the signal files (bigWig format) at once
 - supply both filtered BED files (click on “Add new regions to plot” once) and label them
 - indicate the corresponding signal files
- now use `tools/plotProfile` on the resulting file
 - important: display the “advanced output options” and select “save the data underlying the average profile” -> this will generate a table in addition to the summary plot images



1.3.2 Gallery of deepTools plots

Note: If you have a nice deepTools plot that you'd like to share, we'll be happy to add it to our Gallery! Just send us an email: deeptools@googlegroups.com

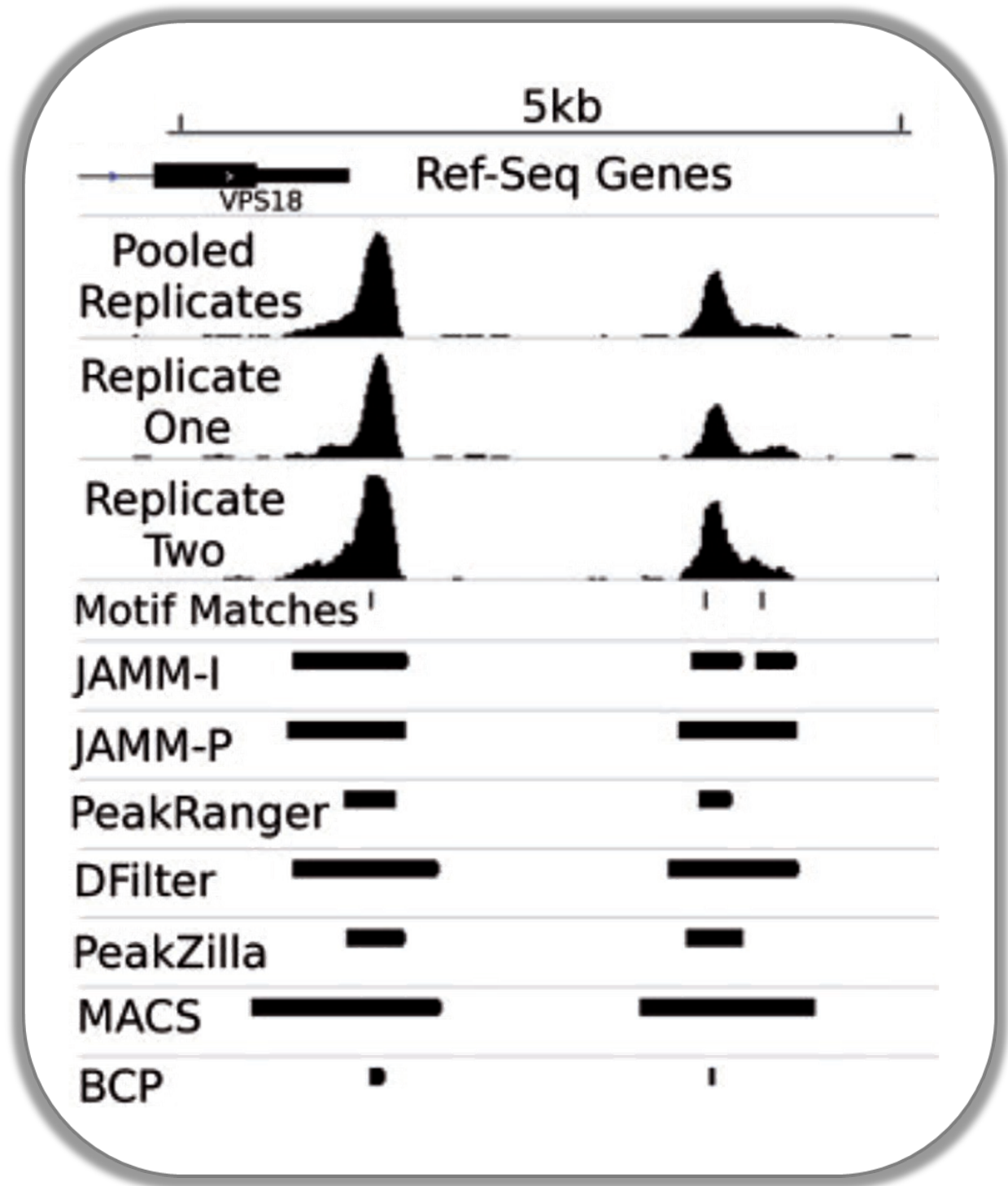
Published example plots

- *Normalized ChIP-seq signals and peak regions*
- *DNase accessibility at enhancers in murine ES cells*
- *TATA box enrichments around the TSS of mouse genes*
- *Visualizing the GC content for mouse and fly genes*
- *CpG methylation around murine transcription start sites in two different cell types*
- *Histone marks for genes of the mosquito Anopheles gambiae*
- *Signals of repressive chromatin marks, their enzymes and repeat element conservation scores*

We're trying to collect a wide variety of plots generated using deepTools. For the plots that we created ourselves, we try to point out the options that were used to create each image, so perhaps these can serve as inspiration for you.

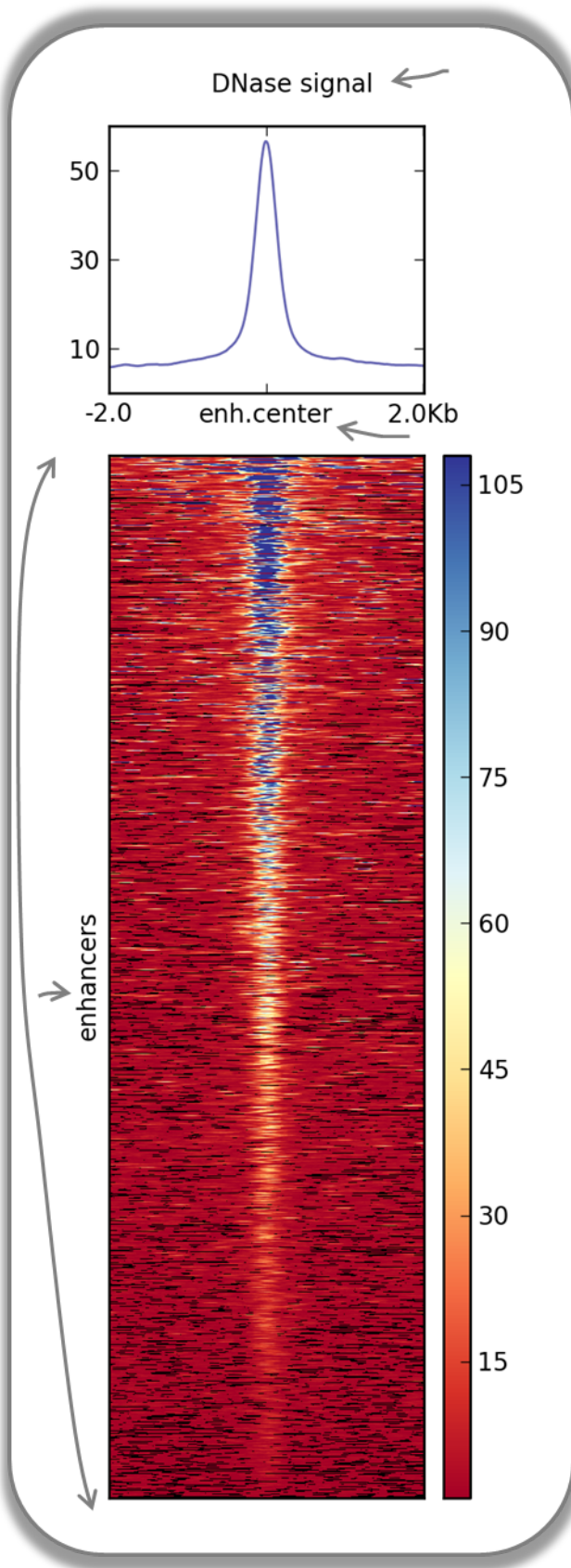
Normalized ChIP-seq signals and peak regions

This image was published by [Ibrahim et al., 2014 \(NAR\)](#). They used deepTools to generate extended reads per kilobase per million reads at 10 base resolution and visualized the resulting coverage files in IGV.



DNase accessibility at enhancers in murine ES cells

The following image demonstrates that enhancer regions are typically small stretches of highly accessible chromatin (more information on enhancers can be found, for example, [here](#)). In the heatmap, yellow and blue tiles indicate a large numbers of reads that were sequenced (indicative of open chromatin) and black spots indicate missing data points. An appropriate labeling of the y-axis was neglected.



Fast Facts:

- *computeMatrix* mode: reference-point
- *regions file*: BED file with typical enhancer regions from [Whyte et al., 2013](#) (download [here](#))
- *signal file*: bigWig file with DNase signal from UCSC
- *heatmap cosmetics*: labels, titles, heatmap height

Command:

```
$ computeMatrix reference-point \
-S DNase_mouse.bigwig \
-R Whyte_TypicalEnhancers_ESC.bed \
--referencePoint center \
-a 2000 -b 2000 \ ## regions before and after the enhancer centers
-out matrix_Enhancers_DNase_ESC.tab.gz

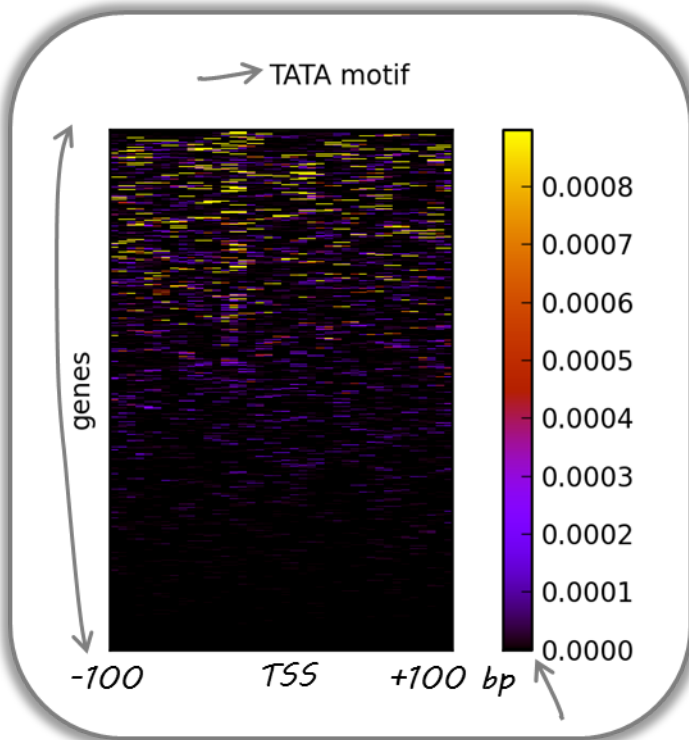
$ plotHeatmap \
-m matrix_Enhancers_DNase_ESC.tab.gz \
-out hm_DNase_ESC.png \
--heatmapHeight 15 \
--refPointLabel enh.center \
--regionsLabel enhancers \
--plotTitle 'DNase signal' \
```

TATA box enrichments around the TSS of mouse genes

Using the [TRAP](#) suite, we produced a bigWig file that contained TRAP scores for the well-known TATA box motif along the mouse genome. The TRAP score is a measure for the strength of a protein-DNA interaction at a given DNA sequence; the higher the score, the closer the motif is to the consensus motif sequence. The following heatmap demonstrates that:

- TATA-like motifs occur quite frequently
- there is an obvious clustering of TATA motifs slightly upstream of the TSS of many mouse genes
- there are many genes that do not contain TATA-like motifs at their promoter

Note that the heatmap shows *all* mouse RefSeq genes, so ca. 15,000 genes!

**Fast Facts:**

- *computeMatrix mode*: reference-point
- *regions file*: BED file with all mouse genes (from UCSC table browser)
- *signal file*: bigWig file of TATA psem scores
- *heatmap cosmetics*: color scheme, labels, titles, heatmap height, only showing heatmap + colorbar

Command:

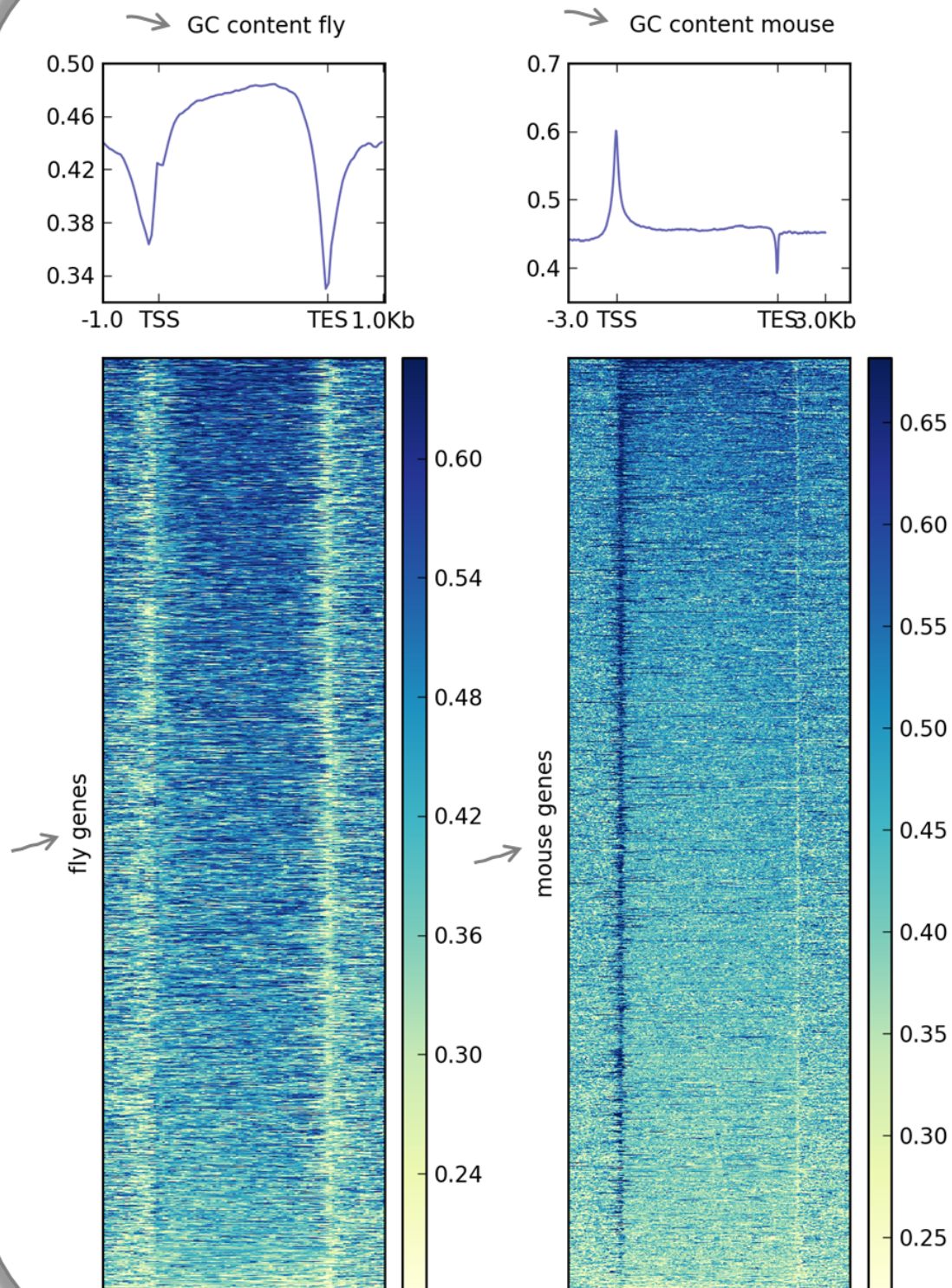
```
$ computeMatrix reference-point \
-S TATA_01_pssm.bw \
-R RefSeq_genes.bed \
--referencePoint TSS \
-a 100 -b 100 \
--binSize 5 \

$ plotHeatmap \
-m matrix_Genes_TATA.tab.gz \
-out hm_allGenes_TATA.png \
--colorMap hot_r \
--missingDataColor .4 \
--heatmapHeight 7 \
--plotTitle 'TATA motif' \
--whatToShow 'heatmap and colorbar' \
--sortRegions ascend
```

Visualizing the GC content for mouse and fly genes

It is well known that different species have different genome GC contents. Here, we used two bigWig files where the GC content was calculated for 50 base windows along the genome of mice and flies and the resulting scores visualized for gene regions.

The images nicely illustrate the completely opposite GC distributions in flies and mice: while the gene starts of mammalian genomes are enriched for Gs and Cs, fly promoters show depletion of GC content.



Fast Facts	
computeMatrix mode	scale-regions
regions files	BED files with mouse and fly genes (from UCSC table browser)
signal file	bigwig files with GC content
heatmap cosmetics	color scheme, labels, titles, color for missing data was set to white, heatmap height

Fly and mouse genes were scaled to different sizes due to the different median sizes of the two species' genes (genes of *D.melanogaster* contain many fewer introns and are considerably shorter than mammalian genes). Thus, computeMatrix had to be run with slightly different parameters while the plotHeatmap commands were virtually identical (except for the labels).

```
$ computeMatrix scale-regions \
-S GCcontent_Mm9_50_5.bw \
-R RefSeq_genes_uniqNM.bed \
-bs 50
-m 10000 -b 3000 -a 3000 \
-out matrix_GCcont_Mm9_scaledGenes.tab.gz \
--skipZeros \
--missingDataAsZero

$ computeMatrix scale-regions \
-S GCcontent_Dm3_50_5.bw \
-R Dm530.genes.bed \
-bs 50
-m 3000 -b 1000 -a 1000 \
-out matrix_GCcont_Dm3_scaledGenes.tab.gz \
--skipZeros --missingDataAsZero

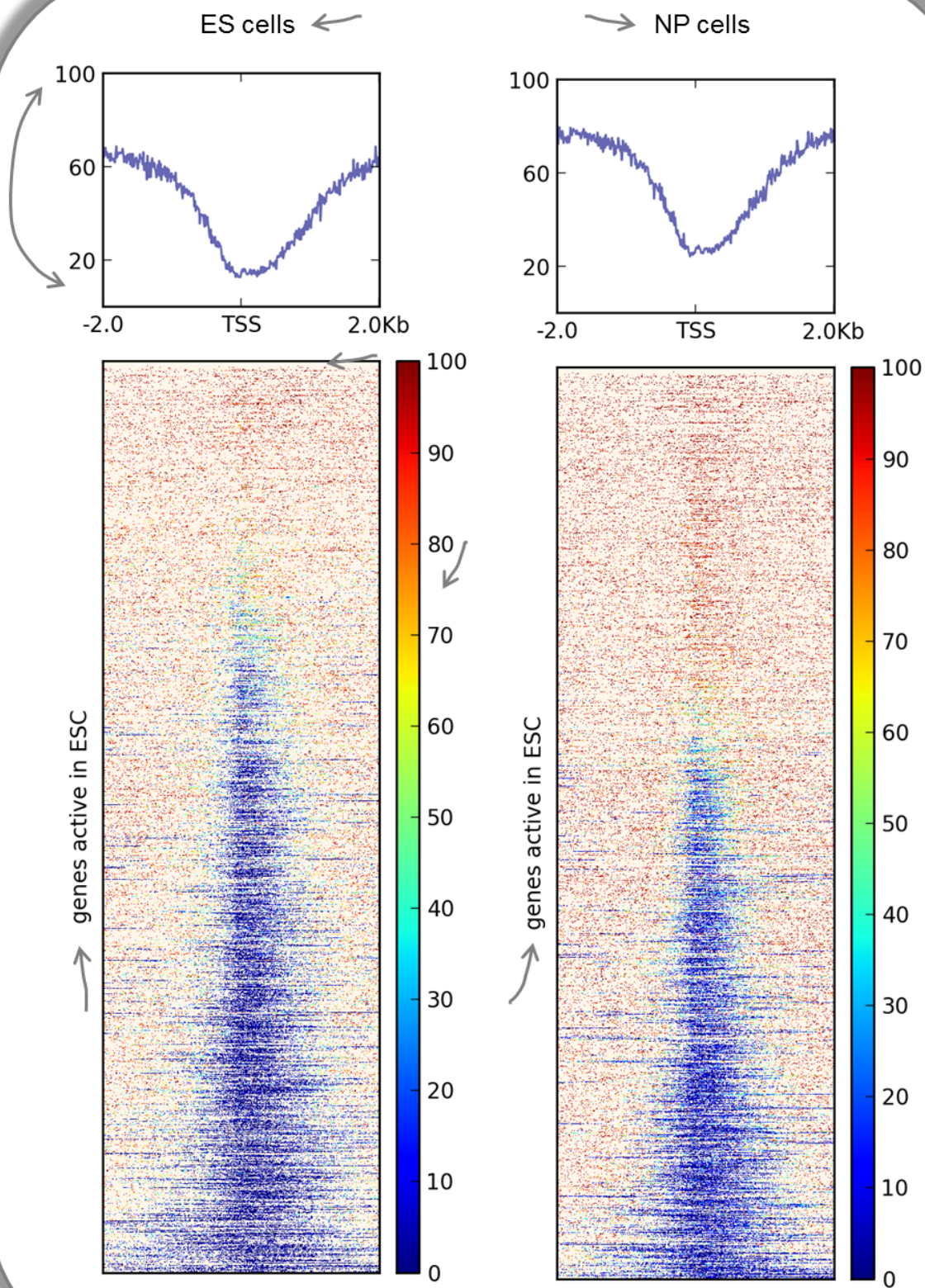
$ plotHeatmap \
-m matrix_GCcont_Dm3_scaledGenes.tab.gz \
-out hm_GCcont_Dm3_scaledGenes.png \
--colorMap YlGnBu \
--regionsLabel 'fly genes' \
--heatmapHeight 15 \
--plotTitle 'GC content fly' &

$ plotHeatmap \
-m matrix_GCcont_Mm9_scaledGenes.tab.gz \
-out hm_GCcont_Mm9_scaledGenes.png \
--colorMap YlGnBu \
--regionsLabel 'mouse genes' \
--heatmapHeight 15 \
--plotTitle 'GC content mouse' &
```

CpG methylation around murine transcription start sites in two different cell types

In addition to the methylation of histone tails, the cytosines can also be methylated (for more information on CpG methylation, read [here](#)). In mammalian genomes, most CpGs are methylated unless they are in gene promoters that need to be kept unmethylated to allow full transcriptional activity. In the following heatmaps, we used genes expressed primarily in ES cells and checked the percentages of methylated cytosines around their transcription start sites. The blue signal indicates that very few methylated cytosines are found. When you compare the CpG methylation signal between ES cells and neuronal progenitor (NP) cells, you can see that the majority of genes remain unmethylated, but the general amount of CpG methylation around the TSS increases, as indicated by the stronger red signal and the slight elevation of the CpG methylation signal in the summary plot. This supports the notion that genes stored in the BED file indeed tend to be more expressed in ES than in NP cells.

This image was taken from [Chelmicki & Dündar et al. \(2014\), eLife](#).



Fast Facts	
computeMatrix mode	reference-point
regions files	<i>BED</i> file mouse genes expressed in ES cells
signal file	<i>bigWig</i> files with fraction of methylated cytosins (from Stadler et al., 2011)
heatmap cosmetics	color scheme, labels, titles, color for missing data was set to customized color, y-axis of profiles were changed, heatmap height

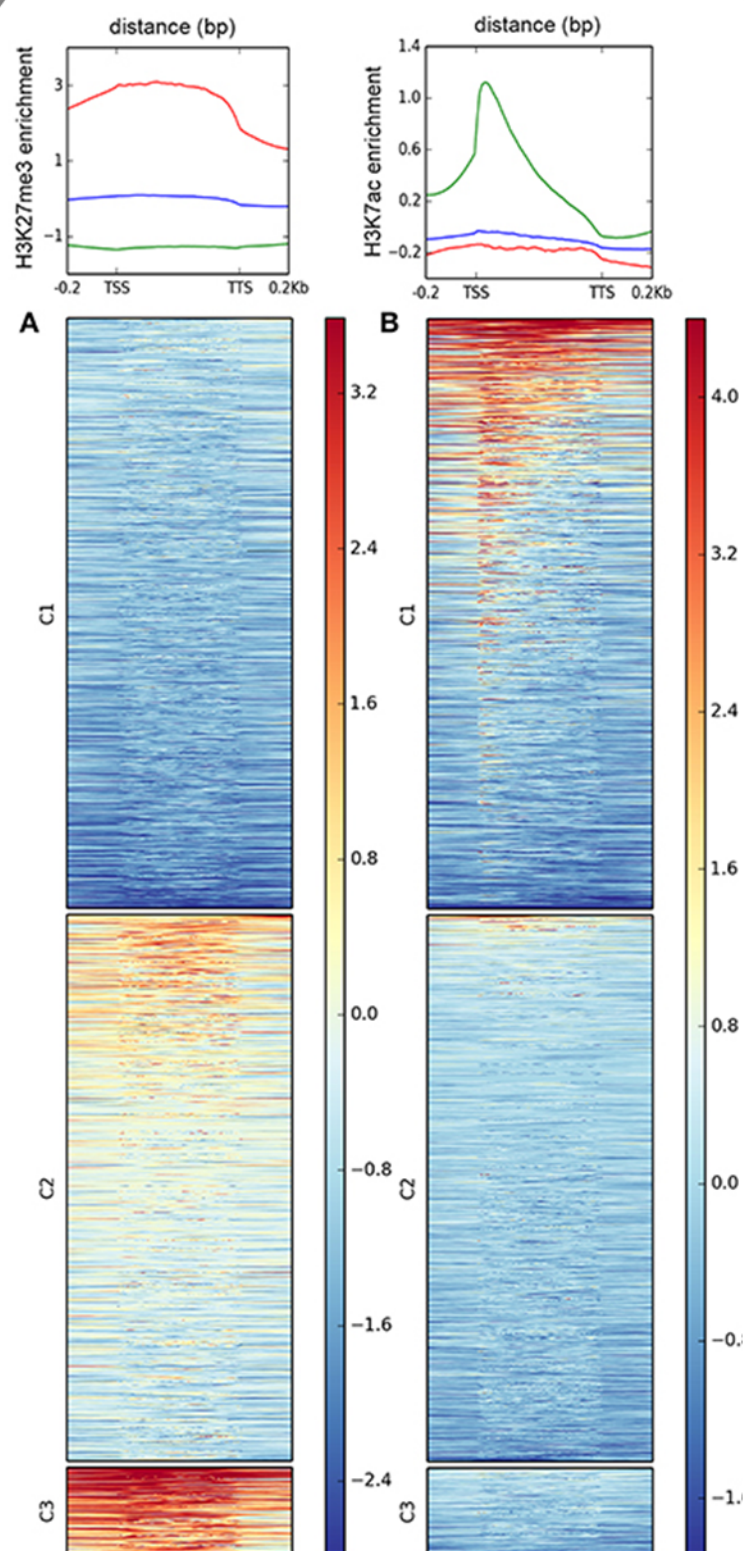
The commands for the bigWig files from the ES and NP cells were the same:

```
$ computeMatrix reference-point \
-S GSE30202_ES_CpGmeth.bw \
-R activeGenes_ESOnly.bed \
--referencePoint TSS \
-a 2000 -b 2000 \
-out matrix_Genes_ES_CpGmeth.tab.gz

$ plotHeatmap \
-m matrix_Genes_ES_CpGmeth.tab.gz \
-out hm_activeESCGenes_CpG_ES_indSort.png \
--colorMap jet \
--missingDataColor "#FFF6EB" \
--heatmapHeight 15 \
--yMin 0 --yMax 100 \
--plotTitle 'ES cells' \
--regionsLabel 'genes active in ESC'
```

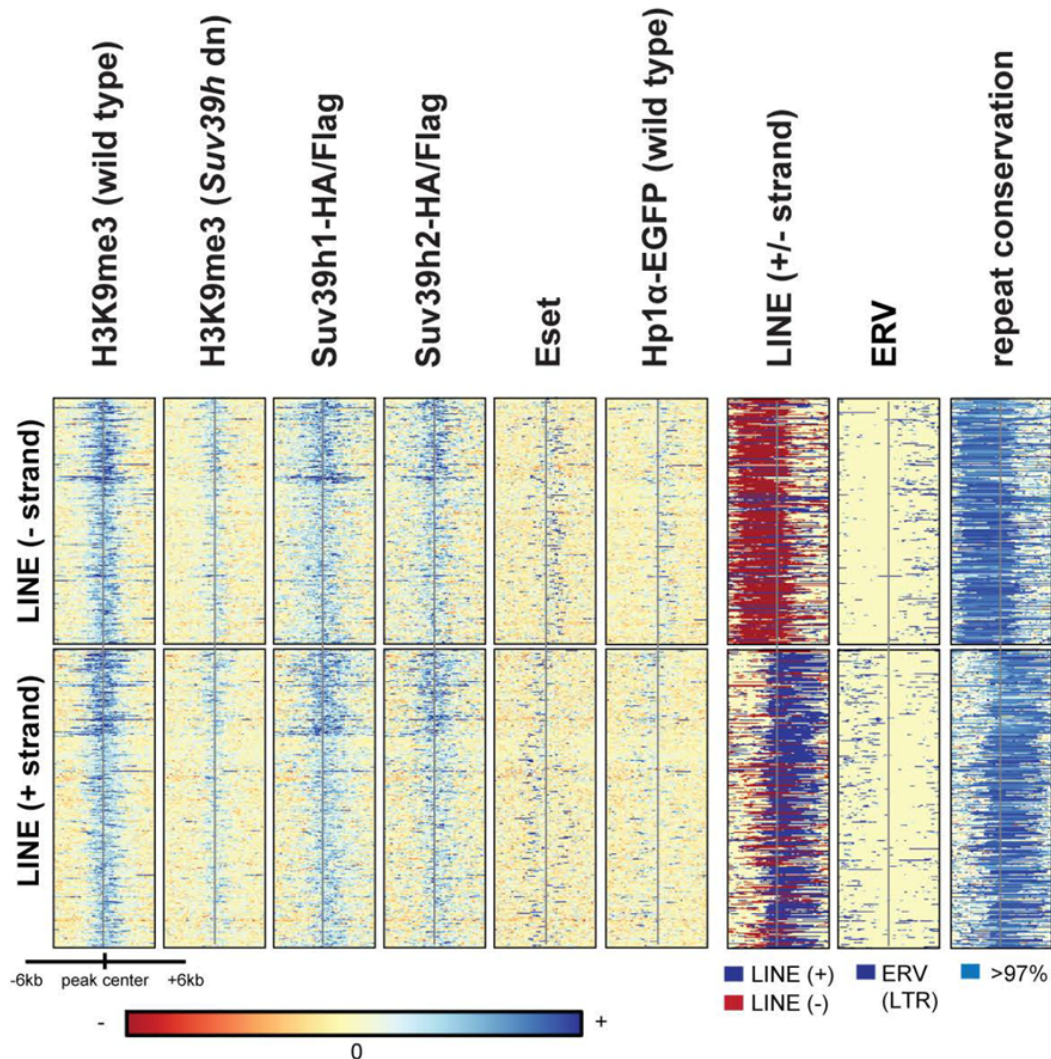
Histone marks for genes of the mosquito *Anopheles gambiae*

This figure was taken from [Gómez-Díaz et al. \(2014\): Insights into the epigenomic landscape of the human malaria vector *Anopheles gambiae*](#). From *Genet* Aug15;5:277. It shows the distribution of H3K27Me3 (left) and H3K27Ac (right) over gene features in *A. gambiae* midguts. The enrichment or depletion is shown relative to chromatin input. The regions in the map comprise gene bodies flanked by a segment of 200 bases at the 5 end of TSSs and TTSs. Average profile across gene regions ± 200 bases for each histone modification are shown on top.



Signals of repressive chromatin marks, their enzymes and repeat element conservation scores

This image is from Bulut-Karsliogu and De La Rosa-Velázquez et al. (2014), Mol Cell. The heatmaps depict various signal types for unscaled peak regions of proteins and histone marks associated with repressed chromatin. The peaks were separated into those containing long interspersed elements (LINEs) on the forward and reverse strand. The signals include normalized ChIP-seq signals for H3K9Me3, Suv39h1, Suv39h2, Eset, and HP1alpha-EGFP, followed by LINE and ERV content and repeat conservation scores.

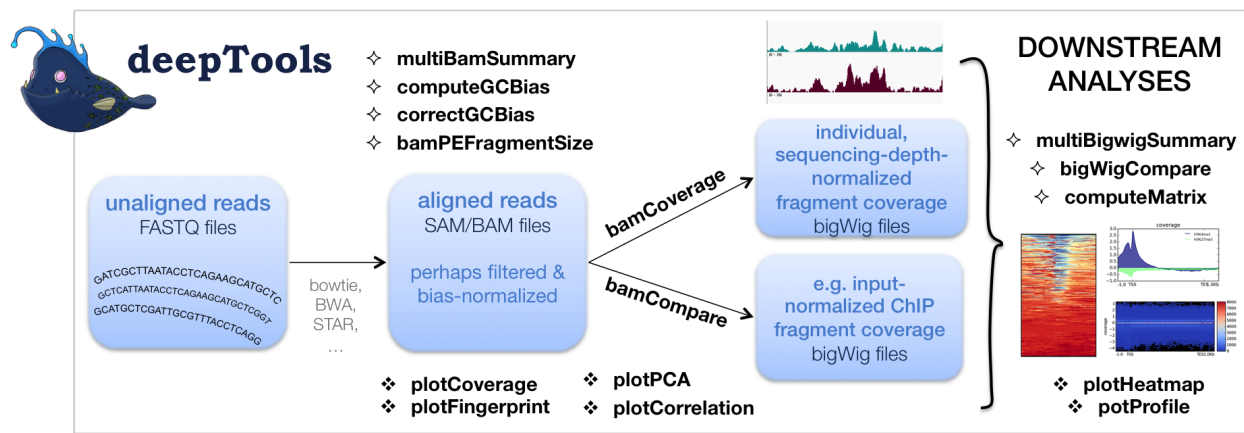


deepTools Galaxy. | code @ github.

1.3.3 How we use deepTools for ChIP-seq analyses

To get a feeling for what deepTools can do, we'd like to give you a brief glimpse into how we typically use deepTools for ChIP-seq analyses. For more detailed examples and descriptions of the tools, simply follow the respective links.

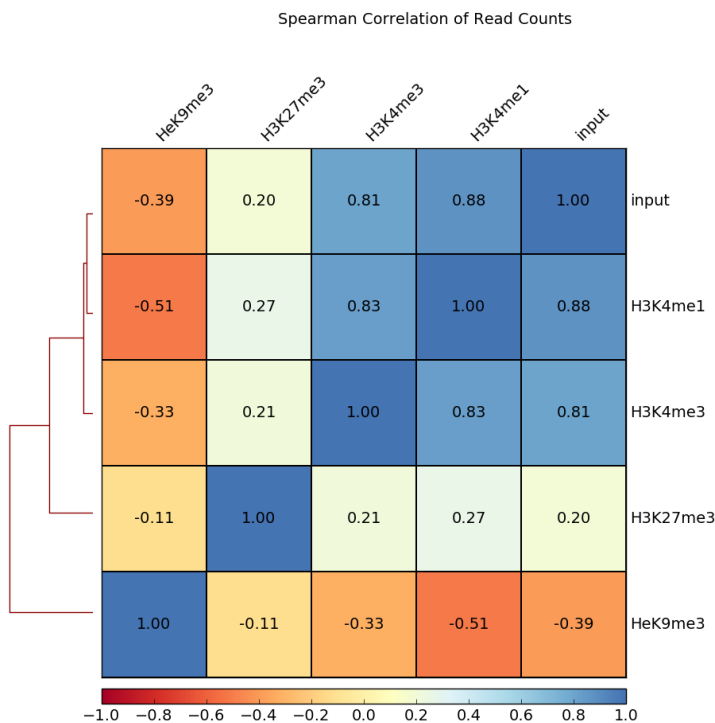
Note: While some tools, such as `tools/plotFingerprint`, specifically address ChIP-seq-issues, the majority of tools is widely applicable to deep-sequencing data, including RNA-seq.



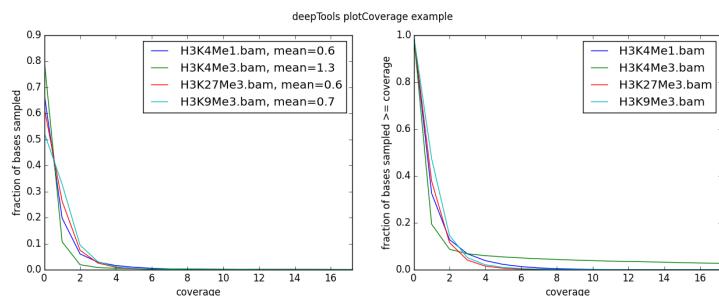
As shown in the flow chart above, our work usually begins with one or more *FASTQ* file(s) of deeply-sequenced samples. After preliminary quality control using *FASTQC*, we align the reads to the reference genome, e.g., using *bowtie2*. The standard output of *bowtie2* (and other mapping tools) is in the form of sorted and indexed *BAM* files that provide the common input and starting point for all subsequent deepTools analyses. We then use deepTools to assess the quality of the aligned reads:

1. **Correlation between BAM files** (`tools/multiBamSummary` and `tools/plotCorrelation`). Together, these two modules perform a very basic test to see whether the sequenced and aligned reads meet your expectations. We use this check to assess reproducibility - either between replicates and/or between different experiments that might have used the same antibody or the same cell type, etc. For instance, replicates should correlate better than differently treated samples.

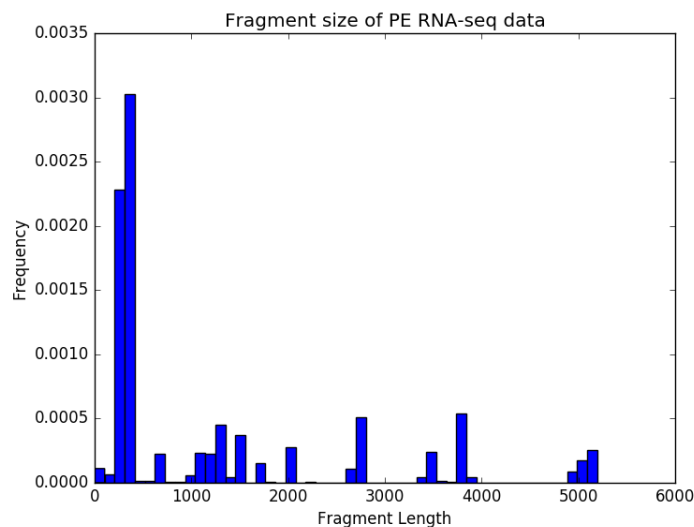
Tip: You can also assess the correlation of *bigWig* files using `tools/multiBigwigSummary`.



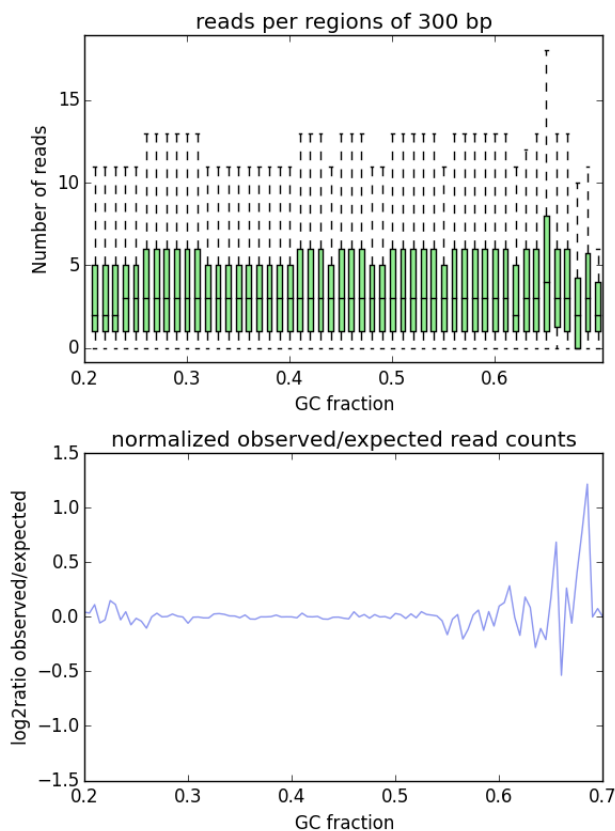
- Coverage check** (`tools/plotCoverage`). To see how many bp in the genome are actually covered by (a good number) of sequencing reads, we use `tools/plotCoverage` which generates two diagnostic plots that help us decide whether we need to sequence deeper or not. The option `--ignoreDuplicates` is particularly useful here!



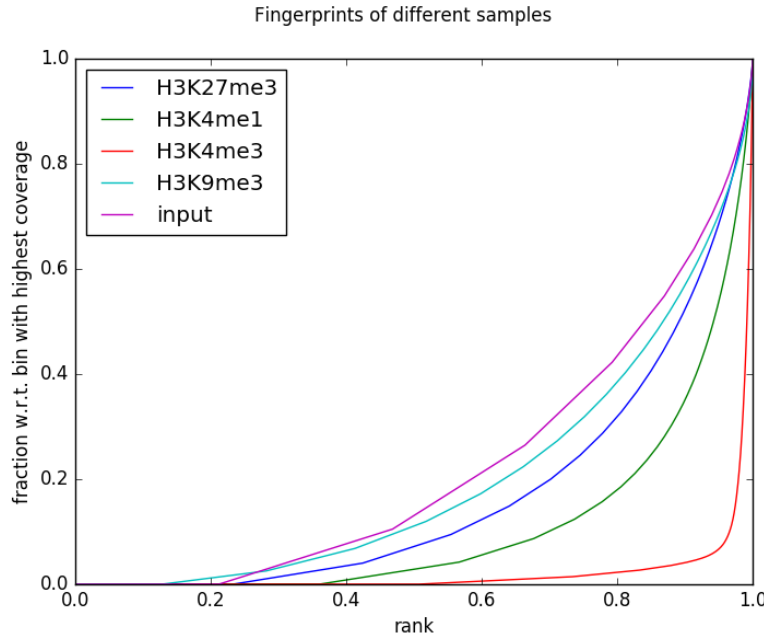
For paired-end samples, we often additionally check whether the fragment sizes are more or less what we would expect based on the library preparation. The module `tools/bamPEfragmentSize` can be used for that.



3. **GC-bias check** (`tools/computeGCBias`). Many sequencing protocols require several rounds of PCR-based DNA amplification, which often introduces notable bias, due to many DNA polymerases preferentially amplifying GC-rich templates. Depending on the sample (preparation), the GC-bias can vary significantly and we routinely check its extent. When we need to compare files with different GC biases, we use the `tools/correctGCBias` module. See the paper by [Benjamini and Speed](#) for many insights into this problem.



4. **Assessing the ChIP strength.** We do this quality control step to get a feeling for the signal-to-noise ratio in samples from ChIP-seq experiments. It is based on the insights published by [Diaz et al.](#)



Once we're satisfied with the basic quality checks, we normally **convert** the large *BAM* files into a leaner data format, typically *bigWig*. *bigWig* files have several advantages over *BAM* files, mainly stemming from their significantly decreased size:

- useful for data sharing and storage
- intuitive visualization in Genome Browsers (e.g. *IGV*)
- more efficient downstream analyses are possible

The deepTools modules `tools/bamCompare` and `tools/bamCoverage` not only allow for simple conversion of *BAM* to *bigWig* (or *bedGraph* for that matter), but also for normalization, such that different samples can be compared despite differences in their sequencing depth.

Finally, once all the converted files have passed our visual inspections (e.g., using the *Integrative Genomics Viewer*), the fun of downstream analysis with `tools/computeMatrix`, `tools/plotHeatmap` and `tools/plotProfile` can begin!

[deepTools Galaxy.](#) | [code @ github.](#)

1.4 Changes in deepTools2.0

- *Major changes*
 - Accommodating additional data types
 - Structural updates
 - Renamed tools
 - Increased efficiency
 - New features and tools
- *Minor changes*
 - Changed parameters names and settings
 - Bug fixes

1.4.1 Major changes

Note: The major changes encompass features for **increased efficiency**, **new sequencing data types**, and **additional plots**, particularly for QC.

Moreover, deepTools modules can now be used by other python programs. The *deepTools API example* is part of the new documentation.

Accommodating additional data types

- correlation and comparisons can now be calculated for **bigWig files** (in addition to BAM files) using `multiBigwigSummary` and `bigwigCompare`
- **RNA-seq:** split-reads are now natively supported
- **MNase-seq:** using the new option `--MNase` in `bamCoverage`, one can now compute read coverage only taking the 2 central base pairs of each mapped fragment into account.

Structural updates

- All modules have comprehensive and automatic tests that evaluate proper functioning after any modification of the code.
- Virtualization for stability: we now provide a `docker` image and enable the easy deployment of deepTools via the Galaxy `toolshed`.
- Our documentation is now version-aware thanks to `readthedocs` and `sphinx`.
- The API is public and documented.

Renamed tools

- **heatmapper** to `tools/plotHeatmap`
- **profiler** to `tools/plotProfile`
- **bamCorrelate** to `tools/multiBamSummary`
- **bigwigCorrelate** to `tools/multiBigwigSummary`
- **bamFingerprint** to `tools/plotFingerprint`

Increased efficiency

- We dramatically improved the **speed** of bigwig related tools (`tools/multiBigwigSummary` and `computeMatrix`) by using the new `pyBigWig` module.
- It is now possible to generate one composite heatmap and/or meta-gene image based on **multiple bigwig files** in one go (see `tools/computeMatrix`, `tools/plotHeatmap`, and `tools/plotProfile` for examples)
- `computeMatrix` now also accepts multiple input BED files. Each is treated as a group within a sample and is plotted independently.

- We added **additional filtering options for handling BAM files**, decreasing the need for prior filtering using tools other than deepTools: The `--samFlagInclude` and `--samFlagExclude` parameters can, for example, be used to only include (or exclude) forward reads in an analysis.
- We separated the generation of read count tables from the calculation of pairwise correlations that was previously handled by `bamCorrelate`. Now, read counts are calculated first using `multiBamSummary` or `multiBigWigCoverage` and the resulting output file can be used for calculating and plotting pairwise correlations using `plotCorrelation` or for doing a principal component analysis using `plotPCA`.

New features and tools

- Correlation analyses are no longer limited to BAM files – bigwig files are possible, too! (see `tools/multiBigwigSummary`)
- Correlation coefficients can now be computed even if the data contains NaNs.
- **Added new quality control tools:**
 - use `tools/plotCoverage` to plot the coverage over base pairs
 - use `tools/plotPCA` for principal component analysis
 - `tools/bamPEFragmentSize` can be used to calculate the average fragment size for paired-end read data
- Added the possibility for **hierarchical clustering**, besides *k*-means to `plotProfile` and `plotHeatmap`
- `plotProfile` has many more options to make compelling summary plots

1.4.2 Minor changes

Changed parameters names and settings

- `computeMatrix` can now read files with DOS newline characters.
- `--missingDataAsZero` was renamed to `--skipNonCoveredRegions` for clarity in `bamCoverage` and `bamCompare`.
- Read extension was made optional and we removed the need to specify a default fragment length for most of the tools: `--fragmentLength` was thus replaced by the new optional parameter `--extendReads`.
- Added option `--skipChromosomes` to `multiBigwigSummary`, which can be used to, for example, skip all ‘random’ chromosomes.
- Added the option for adding titles to QC plots.

Bug fixes

- Resolved an error introduced by numpy version 1.10 in `computeMatrix`.
- Improved plotting features for `plotProfile` when using as plot type: ‘overlapped_lines’ and ‘heatmap’
- Fixed problem with BED intervals in `multiBigwigSummary` and `multiBamSummary` that returned wrongly labeled raw counts.
- `multiBigwigSummary` now also considers chromosomes as identical when the names between samples differ by ‘chr’ prefix, e.g. chr1 vs. 1.

- Fixed problem with wrongly labeled proper read pairs in a BAM file. We now have additional checks to determine if a read pair is a proper pair: the reads must face each other and are not allowed to be farther apart than 4x the mean fragment length.
- For `bamCoverage` and `bamCompare`, the behavior of `scaleFactor` was updated such that now, if given in combination with the normalization options (`--normalizeTo1x` or `--normalizeUsingRPKM`), the given scaling factor will be multiplied with the factor computed by the respective normalization method.

deepTools Galaxy.	code @ github.
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1.5 Using deepTools within Galaxy

Galaxy is a tremendously useful platform developed by the Galaxy Team at Penn State and the Emory University. This platform is meant to offer access to a large variety of bioinformatics tools that can be used without computer programming experiences. That means, that the basic features of Galaxy will apply to every tool, i.e. every tool provided within a Galaxy framework will look very similar and will follow the concepts of Galaxy.

Our publicly available deepTools Galaxy instance can be found here: deeptools.ie-freiburg.mpg.de. This server also contains some additional tools that will enable users to analyse and visualize data from high-throughput sequencing experiments, starting from aligned reads.

Table of content

- *Basic features of Galaxy*
 - *The start site*
 - *Details*
 - *Handling failed files*
 - *Workflows*

1.5.1 Data import into Galaxy

There are three main ways to populate your Galaxy history with data files plus an additional one for sharing data within Galaxy.

- *Upload files from your computer*
- *Import data sets from the Galaxy data library*
- *Download annotation files from public data bases*
- *Copy data sets between histories*

Upload files from your computer

The data upload of files **smaller than 2 GB** that lie on your computer is fairly straight-forward: click on the category “Get data” and choose the tool “Upload file”. Then select the file via the “Browse” button.

Upload File (version 1.1.3)

File Format:

Auto-detect

Which format? See help below

if you're not sure about the data type, leave it up to Galaxy, but it's always good to know before what you're going to upload

File:

Browse...

No file selected.

files < 2GB can directly be uploaded

TIP: Due to browser limitations, uploading files larger than 2GB is guaranteed to fail. To upload large files, use the URL method (below) or FTP (if enabled by the site administrator).

URL/Text:

you can also insert a URL here if that's where your data lies

Here you may specify a list of URLs (one per line) or paste the contents of a file.

Files uploaded via FTP:

File	Size	Date
Your FTP upload directory contains no files.		

This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **deeptools.ie-freiburg.mpg.de** using your Galaxy credentials (email address and password).

Convert spaces to tabs:

☐ Yes

Use this option if you are entering intervals by hand.

Genome:

----- Additional Species Are Below -----

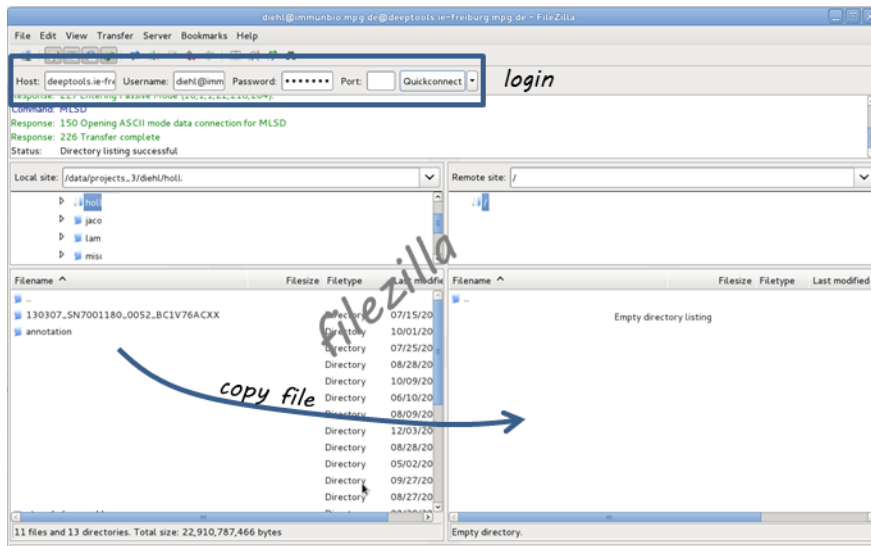
Execute

important! specify the reference genome that was used for aligning the reads!

For files **greater than 2GB**, there's the option to upload via an FTP server. If your data is available via an URL that links to an FTP server, you can simply paste the URL in the empty text box.

If you do not have access to an FTP server, you can directly upload to our Galaxy's FTP.

1. Register with deeptools.ie-freiburg.mpg.de (via "User" "register"; registration requires an email address and is free of charge)
2. You will also need an FTP client, e.g. [filezilla](#).
3. Then login to the **FTP client** using your **deepTools Galaxy user name and password** (host: deeptools.ie-freiburg.mpg.de). Down below you see a screenshot of what that looks like with filezilla.
4. Copy the file you wish to upload to the remote site (in filezilla, you can simply drag the file to the window on the right hand side)
5. Go back to [deepTools Galaxy](#).
6. Click on the tool "Upload file" ("Files uploaded via FTP") - here, the files you just copied over via filezilla should appear. Select the files you want and hit "execute". They will be moved from the FTP server to your history (i.e. they will be deleted from the FTP once the upload was successful).



Import data sets from the Galaxy data library

If you would like to play around with sample data, you can import files that we have saved within the general data storage of the deepTools Galaxy server. Everyone can import them into his or her own history, they will not contribute to the user's disk quota.

You can reach the data library via “Shared Data” in the top menu, then select “Data Libraries”.

Within the Data Library you will find a folder called “Sample Data” that contains data that we downloaded from the [Roadmap project](#) and [UCSC](#). More precisely, we downloaded the [FASTQ][] files of various ChIP-seq samples and the corresponding input and mapped the reads to the human reference genome (version hg19) to obtain the [BAM][] files you see. In addition, you will find bigWig files created using `bamCoverage` and some annotation files in BED format as well as RNA-seq data.

Note: To keep the file size smallish, all files contain data for chromosome 19 and chromosome X only!

Galaxy / deepTools

Tools

search tools

Get Data
Text Manipulation
Filter and Sort
Join, Subtract and Group
Operate on Genomic Intervals
BEDtools
UCSC tools
NGS: Peak Calling
deepTools
Convert Formats

Workflows

All workflows

deepTools

user-friendly tools for the non-bioinformatician

deep-sequencing data

Welcome to the MPI-IE's Galaxy instance dedicated to high-throughput sequencing data!

Did you just receive a large file from your high-throughput sequencing center and now you're wondering how to turn these millions of lines into a heatmap of read coverage? Or how to generate a file you can upload to the UCSC or the IGV browser that you can look at? Thinking more about it, you would actually like to compare your sample that should be the profile of a knock-down cell with the sample from the wild type? And how to make sure that everything went fine with the preparation and sequencing of the samples?

The good news: here, you'll most likely find all the tools and help you need as we have developed several tools to help you make sense out of the data generated by high-throughput sequencing. You will find them under the header **deepTools** in the menu on the left-hand side.

How to get started?

Please make sure that you have read our [Terms of Use](#).

Are you not familiar with the Galaxy framework at all? Please have a look at this manual [here](#). You just need a refresher of how to upload data into Galaxy? [This](#) will help. You do not know the difference between a BAM and a bigWig file? Please make sure you do after looking at [this](#). And finally, [here](#) you can find an overview of the functionalities of deepTools that starts with an overview of typical NGS data analysis workflows and ends with a demonstration to the variability and power of heatmap visualizations for NGS data. For a quick start, the information below might be sufficient though.

Data Libraries

search dataset name, info, message, dbkey

Advanced Search

Data library name

Sample data

the only folder currently available within deepTools Galaxy

click on it

Galaxy / deepTools

Analyze Data Workflow Shared Data Visualization Help User

Using 2%

Data Library "Sample data"

Add datasets Add folder Library Actions

Name	Message	Data type	Date uploaded	File size
<input type="checkbox"/> IMR90_H3K27ac_SRX012496.bam	View information	bam	2013-12-11	842.5 MB
<input type="checkbox"/> IMR90_H3K27ac_SRX012496.bam	Import this dataset into selected histories	bam	2013-12-11	1.1 GB
<input type="checkbox"/> IMR90_H3K27ac_SRX012496.bam	Download this dataset	bam	2013-12-11	565.9 MB
<input type="checkbox"/> IMR90_H3K27me3_SRX012498.bam		bam	2013-12-11	2.0 GB
<input type="checkbox"/> IMR90_H3K27me3_SRX017508.bam		bam	2013-12-11	1.8 GB
<input type="checkbox"/> IMR90_H3K36me3_SRX017509_4.bam		bam	2013-12-11	623.7 MB
<input type="checkbox"/> IMR90_H3K36me3_SRX017511.bam		bam	2013-12-11	1021.0 MB
<input type="checkbox"/> IMR90_Input_SRX017548.bam		bam	2013-12-11	818.3 MB

For selected datasets: [Import to current history](#) [Go](#)

tick here

choose what you'd like to do: import into Galaxy history, download to your computer etc.

click here to return to your history

this will appear if you click on the triangle/arrow

Download annotation files from public data bases

In many cases you will want to query your sequencing data results for known genome annotation, such as genes, exons, transcription start sites etc. These information can be obtained via the two main sources of genome annotation, UCSC and BioMart.

Warning: UCSC and BioMart cater to different ways of genome annotation, i.e. genes defined in UCSC might not correspond to the same regions in a gene file downloaded from BioMart. (For a brief overview over the issues of genome annotation, you can check out [Wikipedia](#), if you always wanted to know much more about those issues, [this](#) might be a good start.)

You can access the data stored at UCSC or BioMart conveniently through our Galaxy instance which will import the resulting files into your history. Just go to “Get data” “UCSC” or “BioMart”.

The majority of annotation files will probably be in [BED][] format, however, you can also find other data sets. UCSC, for example, offers a wide range of data that you can browse via the “group” and “track” menus (for example, you could download the GC content of the genome as a signal file from UCSC via the “group” menu (“Mapping and Sequencing Tracks”).

Warning: The download through this interface is limited to 100,000 lines per file which might not be sufficient for some mammalian data sets.

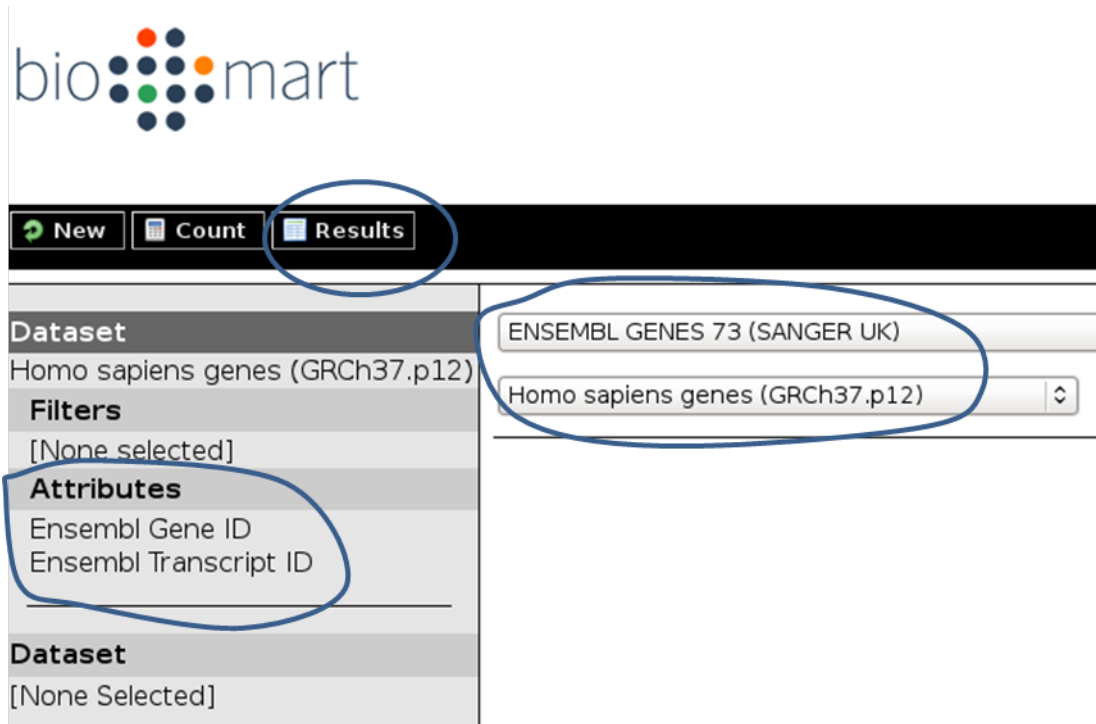
Here’s a screenshot from downloading a BED-file of all RefSeq genes defined for the human genome (version hg19):

The screenshot shows the UCSC Table Browser interface. The top navigation bar includes links for Genomes, Genome Browser, Tools, Mirrors, Downloads, and My Data. The main section is titled "Table Browser". Below this, several configuration options are visible:

- clade:** Mammal (dropdown)
- genome:** Human (dropdown)
- assembly:** Feb. 2009 (GRCh37/hg19) (dropdown)
- group:** Genes and Gene Prediction Tracks (dropdown)
- track:** RefSeq Genes (dropdown)
- table:** refGene (dropdown)
- region:** genome (radio button selected), ENCODE Pilot regions, position (with a text input field containing chr21:33031597-33041570)
- identifiers (names/accessions):** paste list, upload list
- filter:** create
- intersection:** create
- correlation:** create
- output format:** BED - browser extensible data (dropdown)
- Send output to:** ☒ Galaxy (checkbox)
- output file:** (text input field)
- file type returned:** plain text (radio button selected), gzip compressed
- Buttons:** add custom tracks, track hubs, describe table schema, lookup, define regions, paste list, upload list, create, create, create, GREAT, get output, summary/statistics

 Several elements are circled in blue: the "clade", "genome", and "assembly" dropdowns; the "group" and "track" dropdowns; the "output format" dropdown; the "Send output to" checkbox; and the "get output" button.

And here’s how you would do it for the BioMart approach:



Tip: Per default, **BioMart** will not output a **BED** file like UCSC does. It is therefore important that you make sure you get all the information you need (most likely: chromosome, gene start, gene end, ID, strand) via the “Attributes” section. You can click on the “Results” button at any time to check the format of the table that will be sent to Galaxy (Note that the strand information will be decoded as 1 for “forward” or “plus” strand and -1 for “reverse” or “minus” strand).

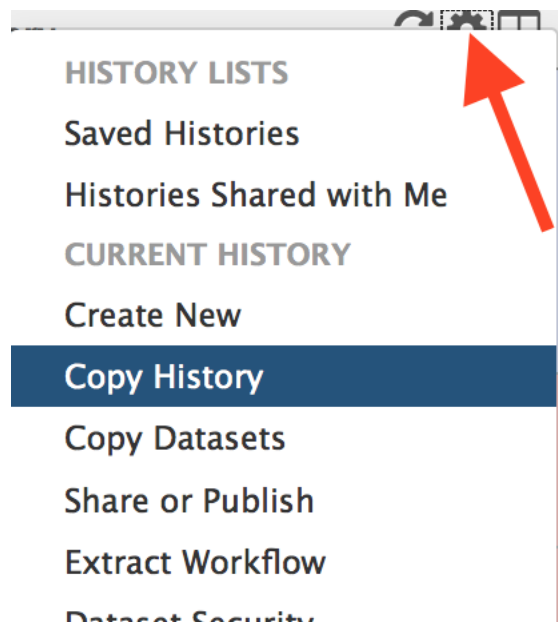
Warning: Be aware, that BED files from UCSC will have chromosomes labelled with “chr” while ENSEMBL usually returns just the number – this might lead to incompatibilities, i.e. when working with annotations from UCSC and ENSEMBL, you need to make sure to use the same naming!

Copy data sets between histories

If you have registered with deepTools Galaxy you can have more than one history.

In order to minimize the disk space you’re occupying we strongly suggest to **copy** data sets between histories when you’re using the same data set in different histories.

Note: Copying data sets is only possible for registered users.



Copying can easily be done via the History panel's option button "Copy dataset". In the main frame, you should now be able to select the history you would like to copy from on the left hand side and the target history on the right hand side.

More help

Hint: If you encounter a failing data set (marked in red), please **send a bug report** via the Galaxy bug report button and we will get in touch if you indicate your email address.

http://wiki.galaxyproject.org/Learn	Help for Galaxy usage in general
deepTools Galaxy FAQs	Frequently encountered issues with our specific Galaxy instance
deeptools@googlegroups.com	For issues not addressed in the FAQs

deepTools Galaxy.	code @ github.
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1.5.2 Which tools can I find in the deepTools Galaxy?

As mentioned before, each Galaxy installation can be tuned to the individual interests. Our goal is to provide a Galaxy that enables you to **quality check, process and normalize and subsequently visualize your data obtained by high-throughput DNA sequencing.**

Tip: If you do not know the difference between a BAM and a BED file, that's fine. You can read up on them in our [Glossary of NGS terms](#).

Tip: For more specific help, check our [Galaxy-related FAQ](#) and the [Step-by-step protocols](#).

We provide the following kinds of tools:

- *deepTools*
 - *Tools for BAM and bigWig file processing*
 - *Tools for QC of NGS data*
 - *Heatmaps and summary plots*
- *Working with text files and tables*
 - *Text manipulation*
 - *Filter and Sort*
 - *Join, Subtract, Group*
- *Basic arithmetics for tables*

deepTools

The most important category is called “**deepTools**” that contains all the main tools we have developed.

Tools for BAM and bigWig file processing

multiBamSummary	get read counts for the binned genome or user-specified regions
multiBigwigSummary	calculate score summaries for the binned genome or user-specified regions
correctGCBias	obtain a BAM file with reads distributed according to the genome’s GC content
bamCoverage	obtain the normalized read coverage of a single BAM file
bamCompare	normalize 2 BAM files to each other (e.g. log2ratio, difference)
bigwigCompare	normalize the scores of two bigWig files to each other (e.g., ratios)
computeMatrix	compute the values needed for heatmaps and summary plots

Tools for QC of NGS data

plotCorrelation	calculate and visualize the pairwise Spearman or Pearson correlation of read counts (or other scores)
plotPCA	perform PCA and visualize the results
plotFingerprint	assess the ChIP enrichment strength
bamPEFragmentSize	obtain the average fragment length for paired-end samples
computeGCBias	assess the GC bias by calculating the expected and observed GC distribution of aligned reads
plotCoverage	obtain the normalized read coverage of a single BAM file

Heatmaps and summary plots

plotHeatmap	visualize read counts or other scores in heatmaps with one row per genomic region
plotProfile	visualize read counts or other scores using average profiles (e.g., meta-gene profiles)

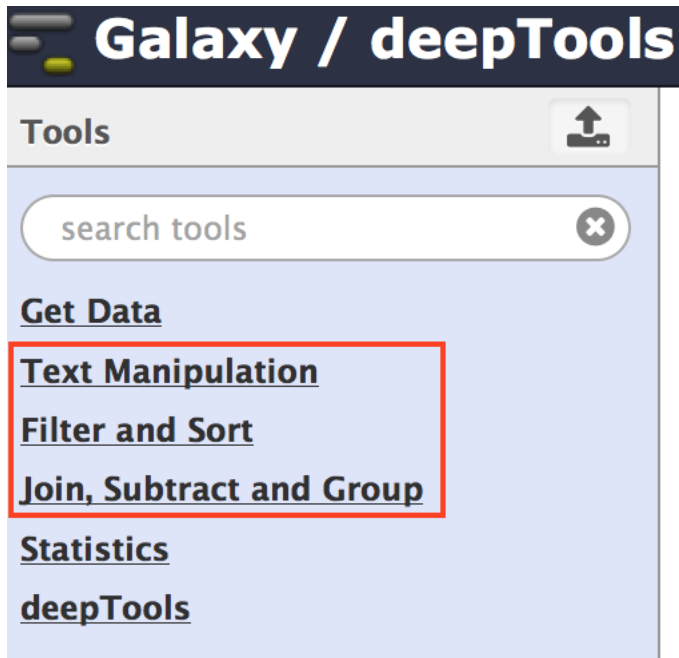
For each tool, you can find example usages and tips within Galaxy once you select the tool.

In addition, you may want to check our pages about [Example usage](#), particularly [Step-by-step protocols](#).

Working with text files and tables

In addition to deepTools that were specifically developed for the handling of NGS data, we have incorporated several standard Galaxy tools that enable you to manipulate tab-separated files such as gene lists, peak lists, data matrices etc.

There are 3 main categories;



Text manipulation

Unlike Excel, where you can easily interact with your text and tables via the mouse, data manipulations within Galaxy are strictly based on commands.

If you feel like you would like to do something to certain *columns* of a data set, go through the tools of this category!

Example actions are: * adding columns * extracting columns * pasting two files side by side * selecting random lines * etc.

A very useful tool of this category is called `Trim`: if you need to **remove some characters from a column**, this tool's for you! (for example, sometimes you need to adjust the chromosome naming between two files from different source - using `Trim`, you can remove the "chr" in front of the chromosome name)

Filter and Sort

In addition to the common sorting and filtering, there's the very useful tool to select lines that match an expression. For example, using the expression `cl=='chrM'` will select all rows from a BED file with regions located on the mitochondrial chromosome.

Filter and Sort

Filter data on any column using simple expressions

Sort data in ascending or descending order

Select lines that match an expression

Join, Subtract, Group

The tools of this category are very useful if you have several data sets that you would like to work with, e.g. by comparing them.

Join, Subtract and Group

Join two Datasets side by side on a specified field

Compare two Datasets to find common or distinct rows

Group data by a column and perform aggregate operation on other columns.

Basic arithmetics for tables

We offer some very basic mathematical operations on values stored with tables. The `Summary Statistics` can be used to calculate the sum, mean, standard deviation and percentiles for a set of numbers, e.g. for values stored in a specific column.

Statistics

Summary Statistics for any numerical column

Count occurrences of each record

More help

Hint: If you encounter a failing data set (marked in red), please **send a bug report** via the Galaxy bug report button

and we will get in touch if you indicate your email address.

http://wiki.galaxyproject.org/Learn	Help for Galaxy usage in general
deepTools Galaxy FAQs	Frequently encountered issues with our specific Galaxy instance
deeptools@googlegroups.com	For issues not addressed in the FAQs
deepTools Galaxy.	code @ github.

1.5.3 Basic features of Galaxy

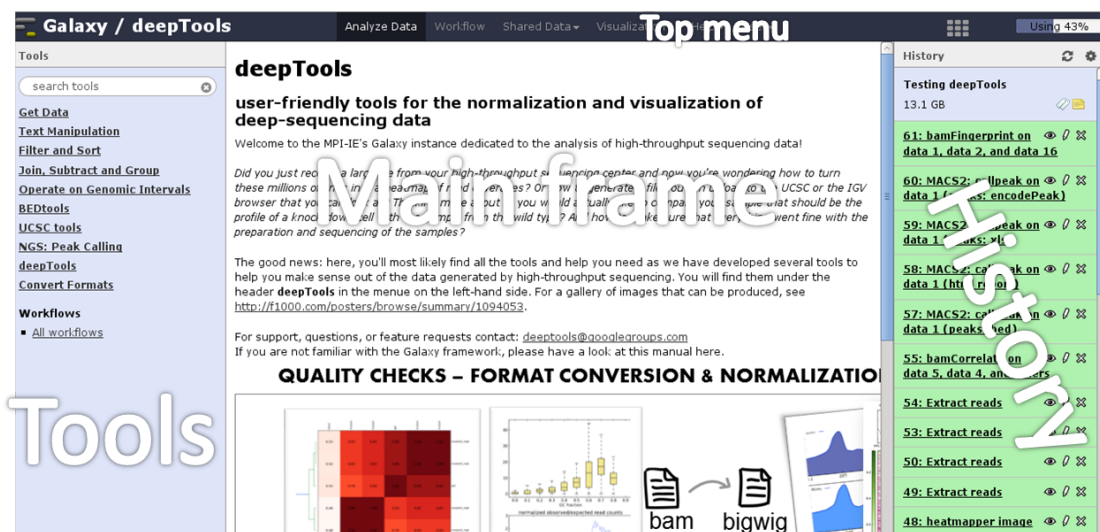
Galaxy is a web-based platform for data intensive, bioinformatics-dependent research and it is being developed by Penn State and John Hopkins University. The original Galaxy can be found [here](#).

Since it is impossible to meet all bioinformatics needs – that can range from evolutionary analysis to data from mass spectrometry to high-throughput DNA sequencing (and way beyond) – with one single web server, many institutes have installed their own versions of the Galaxy platform tuned to their specific needs.

Our **deepTools Galaxy** is such a specialized server dedicated to the analysis of high-throughput DNA sequencing data. The overall makeup of this web server, however, is the same as for any other Galaxy installation, so if you’ve used Galaxy before, you will learn to use deepTools in no time!

The start site

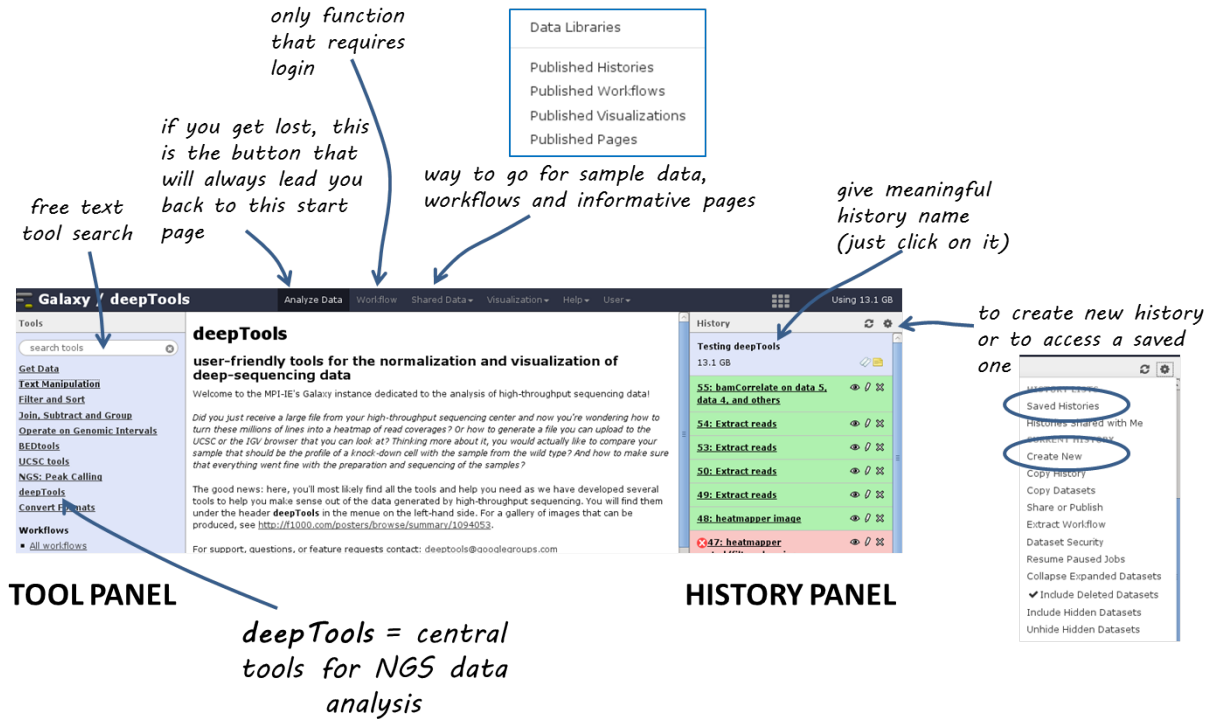
Here is a screenshot of what the start site will approximately look like:



The start site contains 4 main features:

Top menu	Your gateway away from the actual analysis part into other sections of Galaxy, e.g. workflows and shared data .
Tool panel	<i>What can be done?</i> Find all tools installed in this Galaxy instance
Main frame	<i>What am I doing?</i> This is your main working space where input will be required from you once you’ve selected a tool.
History panel	<i>What did I do?</i> The history is like a log book : everything you ever did is recorded here.

For those visual learners, here’s an annotated screenshot:



Details

In the default state of the tool panel you see the **tool categories**, e.g. “Get Data”. If you click on them, you will see the **individual tools** belonging to each category, e.g. “Upload File from your computer”, “UCSC Main table browser” and “Biomart central server” in case you clicked on “Get Data”. To use a tool such as “Upload File from your computer”, just click on it.

The **tool *search* panel** is extremely useful as it allows you to enter a key word (e.g. “bam”) that will lead to all the tools mentioning the key word in the tool name.

Once you’ve uploaded any kind of data, you will find the history on the right hand side filling up with green tiles. Each tile corresponds to one data set that you either uploaded or created. The data sets can be images, raw sequencing files, text files, tables - virtually anything. The content of a data set *cannot* be modified - every time you want to change something *within* a data file (e.g. you would like to sort the values or add a line or cut a column), you will have to use a Galaxy tool that will lead to a *new* data set being produced. This behaviour is often confusing for Galaxy novices (as histories tend to accumulate data sets very quickly), but it is necessary to enforce the strict policy of documenting *every modification* to a given data set. Eventhough your history might be full of data sets with strange names, you will always be able to track back the source and evolution of each file. Also, every data set can be downloaded to your computer individually. Alternatively, you can *download* an entire history or *share* the history with another user.

Have a look at the following screenshot to get a feeling for how many information Galaxy keeps for you (which makes it very feasible to reproduce any analysis):

Attributes

Convert Format

Datatype

Permissions

Edit Attributes

Name:

profiler image

Info:

Annotation / Notes:

Add an annotation or notes to a dataset; annotations are available when a history is viewed.

Database/Build:

Human Feb. 2009 (GRCh37/hg19...

Save

Auto-detect

This will inspect the dataset and attempt to correct the above column values if they are not accurate.

have a look here as well

change the file name

put some info you would like to keep, e.g. what experiment this file is related to, why you generated it etc.

edit attributes

view the file

8: bamCorrelate on data 5, data 4, and others

64.9 KB

format: png, database: hg19

image in png format

delete the file (can be recovered)

re-run an analysis with the exact same parameters !!extremely useful!!

download the file

details about how this file was generated

History

48: heatmap image

47: heatmap sorted/filtered regions

46: heatmap matrix of heatmap values

45: heatmap image

44: computeMatrix on data 32 and data 34 regions

43: computeMatrix on data 32 and data 34

42: computeMatrix on data 32 and data 34 column

41: computeMatrix on data 32 and data 34

40: profiler image

39: heatmap image

38: computeMatrix on data 32 and data 34

37: Extract reads

Include Deleted Datasets

Include Hidden Datasets

Unhide Hidden Datasets

Delete Hidden Datasets

Purge Deleted Datasets

Show Structure

Export to File

Delete

Delete Permanently

Import from File

Tool: bamCorrelate	
Name:	bamCorrelate on data 5, data 4, and others
Created:	Dec 11, 2013
Filesize:	64.9 KB
Dbkey:	hg19
Format:	png
Galaxy Tool Version:	1.0.1
Tool Version:	
Tool Standard Output:	stdout
Tool Standard Error:	stderr
Tool Exit Code:	0
API ID:	a50c14e4ca28bfa0
Input Parameter	
Bam file	6: IMR90_H3K36me3.bam
Label	
Bam file	5: IMR90_H3K27me3_2.bam
Label	
Bam file	4: IMR90_H3K27me3_1.bam
Label	
Bam file	3: IMR90_H3K27ac_3.bam
Label	
Bam file	2: IMR90_H3K27ac_2.bam
Label	
Bam file	1: IMR90_H3K27ac_1.bam
Label	
Length of the average fragment size	200
Correlation method	Pearson
Choose computation mode	bins
Bin size in bp	10000
Number of samples	100000
Show advanced options	no
Show additional output options	no

Each data set can have 4 different states that are intuitively color-coded:

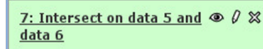
Waiting to be run



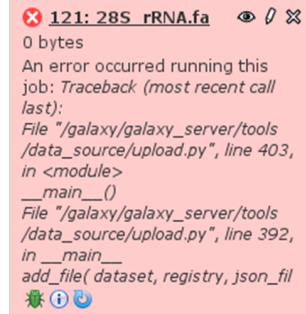
Running



Finished successfully



Failed



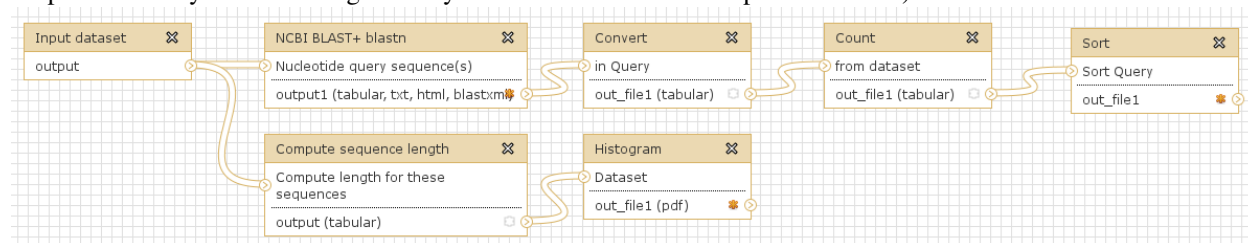
Handling failed files

If you encounter a failed file after you've run a tool, please do the following steps (**in this order**):

1. click on the center button on the lower left corner of the failed data set (i): did you chose the **correct data files**?
2. if you're sure that you chose the correct files, hit the **re-run** button (blue arrow in the lower left corner) - check again whether your files had the **correct file format**. If you suspect that the format might be incorrectly assigned (e.g. a file that should be a BED file is labelled as a tabular file), click the **edit** button (the pencil) of the input data file - there you can change the corresponding attributes
3. if you've checked your input data and the error is persisting, click on the **green bug** (lower left corner of the failed data set) and send the **bug report** to us. You do not need to indicate a valid email-address unless you would like us to get in touch with you once the issue is solved.

Workflows

Workflows are Galaxy's equivalent of protocols. This is a very useful feature as it allows users to *share their protocols and bioinformatic analyses* in a very easy and transparent way. This is the graphical representation of a Galaxy workflow that can easily be modified via drag'n'drop within the workflows manual (you must be registered with deepTools Galaxy to be able to generate your own workflows or edit published ones).



More help

Hint: If you encounter a failing data set (marked in red), please **send a bug report** via the Galaxy bug report button and we will get in touch if you indicate your email address.

http://wiki.galaxyproject.org/Learn	Help for Galaxy usage in general
deepTools Galaxy FAQs	Frequently encountered issues with our specific Galaxy instance
deeptools@googlegroups.com	For issues not addressed in the FAQs

deepTools Galaxy.	code @ github.
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1.6 General FAQ

Feel free to contribute your questions via deeptools@googlegroups.com

Note: We also have a [Galaxy-related FAQ](#) with questions that are more specific to Galaxy rather than deepTools usage.

- *How does deepTools handle data from paired-end sequencing?*
- *How can I test a tool with little computation time?*
- *Can I specify more than one chromosome in the `--regions` option?*
 - General workaround
 - Build-in solutions
- *When should I exclude regions from `computeGCBias`?*
- *When should I use `bamCoverage` or `bamCompare`?*
- *What should I pay attention to when dealing with RNA-seq data?*
- *How does `computeMatrix` handle overlapping genome regions?*
 - Galaxy-based work around
 - Command line-based work arounds
- *Why does the maximum value in the heatmap not equal the maximum value in the matrix?*
- *The heatmap I generated looks very “coarse”, I would like a much more fine-grained image.*
- *How can I change the automatic labels of the clusters in a k-means clustered heatmap?*
- *How can I manually specify several groups of regions (instead of clustering)?*
- *What do I have to pay attention to when working with a draft version of a genome?*
- *How do I calculate the effective genome size for an organism that’s not in your list?*
 - Use GEM
 - Use faCount
 - Use bamCoverage
 - Use genomeCoverageBed
- *Where can I download the 2bit genome files required for `computeGCBias`?*

1.6.1 How does deepTools handle data from paired-end sequencing?

Generally, all the modules working on *BAM* files (`multiBamSummary`, `bamCoverage`, `bamCompare`, `plotFingerprint`, `computeGCBias`) automatically recognize paired-end sequencing data and will use the fragment size based on the distance between read pairs. You can by-pass the typical fragment handling on mate pairs with the option `--doNotExtendPairedEnds` (can be found under “advanced options” in Galaxy).

1.6.2 How can I test a tool with little computation time?

When you’re playing around with the tools to see what kinds of results they will produce, you can limit the operation to one chromosome or a specific region to save time. In Galaxy, you will find this under “advanced output options” → “Region of the genome to limit the operation to”. The command line option is called `--region (CHR:START:END)`.

The following tools currently have this option:

- `tools/multiBamSummary`
- `tools/plotFingerprint`
- `tools/computeGCBias`, `tools/correctGCBias`
- `tools/bamCoverage`, `tools/bamCompare`

It works as follows: first, the *entire* genome represented in the *BAM* file will be regarded and sampled, *then* all the regions or sampled bins that do not overlap the region indicated by the user will be discarded.

Note: You can limit the operation to only **one** chromosome (or **one** specific locus on a chromosome) at a time. If you would like to limit the operation to more than one region, see the answer to the next question.

1.6.3 Can I specify more than one chromosome in the `--regions` option?

The short answer is: no.

Several programs allow specifying a specific regions. For these, the input must be in the format of `chr:start:end`, for example “chr10” or “chr10:456700:891000”.

For these programs, it is **not possible** to indicate more than one region, e.g. chr10, chr11 - **this will not work!** Here are some ideas for workarounds if you none-the-less need to do this:

General workaround

Since all the tools that have the `--region` option work on *BAM* files, you could *filter your reads* prior to running the program, e.g. using `intersectBed` with `--abam` or `samtools view`. Then use the resulting (smaller) BAM file with the deepTools program of your choice.

```
$ samtools view -b -L regionsOfInterest.bed Reads.bam > ReadsOverlappingWithRegionsOfInterest.bam
```

or

```
$ intersectBed -abam Reads.bam -b regionsOfInterest.bed > ReadsOverlappingWithRegionsOfInterest.bam
```

Build-in solutions

`computeGCBias` and `multiBamSummary` offer build-in solutions so that you do not need to resort to tools outside of deepTools.

tools/multiBamSummary has two modes, **bins** and **BED**. If you make use of the BED mode, you can supply a *BED* file of regions that you would like to limit the operation to. This will do the same thing as in the general workaround mentioned above.

tools/computeGCBias has a **--filterOut** option. If you to create a BED file that contains all the regions you are **not** interested in, you can then supply this file to `computeGCBias --filterOut Regions_to_be_ignored.bed` and those regions will subsequently be ignored.

1.6.4 When should I exclude regions from `computeGCBias`?

Note: In general, we recommend to only correct for GC bias (using `tools/computeGCBias` followed by `tools/correctGCBias`) if the majority of the genome (e.g., for mouse and human genomes the region between 30-60%) is GC-biased *and* you want to compare this sample with another sample that is not GC-biased.

Sometimes, a certain GC bias is expected, for example for ChIP samples of H3K4Me3 in mammalian samples where GC-rich promoters are expected to be enriched. To not confound the GC bias caused by the library preparation with the inherent, expected GC-bias, we incorporated the possibility to supply a file of regions to `computeGCBias` that will be excluded from the GC bias calculation. This file should typically contain those regions that one expects to be significantly enriched. This allows the tool to focus on background regions.

1.6.5 When should I use `bamCoverage` or `bamCompare`?

Both tools produce *bigWig* files, i.e. they translate the read-centered information from a *BAM* file into scores for genomic regions of a fixed size. The only difference is the *number of BAM files* that the tools use as input: while `tools/bamCoverage` will only take one *BAM* file and produce a coverage file that is mostly normalized for sequencing depth, `tools/bamCompare` will take *two BAM* files that can be compared with each other using several mathematical operations.

`bamCompare` will always normalize for sequencing depth like `bamCoverage`, but then it will perform additional calculations depending on what the user chose, for example:

- **ChIP vs. *input*** obtain a *bigWig* file of `log2ratios(ChIP/input)`
 - **treatment vs. control** obtain a *bigWig* file of *differences* (treatment - control)
 - **replicate 1 and replicate 2** obtain a *bigWig* file where the values from two *BAM* files are summed up (replicate 1 + replicate 2)
-

1.6.6 What should I pay attention to when dealing with RNA-seq data?

By default, deepTools (**since version 2**) makes use of the information stored in the so-called CIGAR string of the alignment file (*SAM/BAM specification*). The CIGAR tells precisely to which bases of the reference a read maps - and, accordingly, which bases are skipped in the case of reads that span introns. These so-called split reads are natively handled by all modules of deepTools 2.0.

Warning: It is generally **not** recommended to activate the deepTools parameter `--extendReads` for RNA-seq data.

This is because there is no verified information on the fragment alignment outside the actual read sequence. A simple extension of a read over uncovered parts would probably be wrong for a lot of fragments! Activating the read extension also **deactivates** the utilization of the CIGAR.

1.6.7 How does computeMatrix handle overlapping genome regions?

If the *BED* file supplied to `tools/computeMatrix` contains regions that overlap, the tool will report those regions and issue warnings, but they will just be taken as is. If you would like to prevent this, then clean the *BED* file before using `computeMatrix`. There are several methods for modifying your *BED* file.

Let's say your file looks like this:

```
$ cat testBed.bed
chr1      10      20      region1
chr1      7       15      region2
chr1      18      29      region3
chr1      35      40      region4
chr1      10      20      region1Duplicate
```

Galaxy-based work around

To eliminate entries with *identical* genome coordinates, first use the tool “Count” and then filter out all entries that are present more than once.

1	2	3	4
chr1	10	20	region1
chr1	7	15	region2
chr1	18	29	region3
chr1	35	40	region4
chr1	10	20	region1Duplicate

starting point
 INTERVAL file with 1 duplicate entry

output

1	2	3	4
2	chr1	10	20
1	chr1	18	29
1	chr1	35	40
1	chr1	7	15

unique entries will have only 1 count

output

1	2	3
chr1	10	20
chr1	18	29
chr1	35	40
chr1	7	15

feed this into deepTools

Command line-based work arounds

- if you just want to eliminate *identical* entries (here: region1 and region1Duplicate), use `sort` and `uniq` in the shell (note that the label of the identical regions is different - as `uniq` can only ignore fields at the beginning of a file, use `rev` to revert the sorted file, then `uniq` with ignoring the first field (which is now the name column) and then revert back:

```
$ sort -k1,1 -k2,2n testBed.bed | rev | uniq -f1 | rev
chr1      10      20      region1
chr1      7       15      region2
chr1     18      29      region3
chr1     35      40      region4
```

- if you would like to *merge all overlapping regions* into one big one, use the `mergeBed` from the BEDtools suite:
 - again, the BED file must be sorted first
 - `-n` and `-nms` tell `mergeBed` to output the number of overlapping regions and the names of them
 - in the resulting file, regions 1, 2 and 3 are merged

```
$ sort -k1,1 -k2,2n testBed.bed | mergeBed -i stdin -n -nms
chr1      7      29      region2;region1;region1Duplicate;region3      4
chr1     35      40      region4 1
```

- if you would like to *keep only regions that do not overlap* with any other region in the same BED file, use the same `mergeBed` routine but subsequently filter out those regions where several regions were merged.
 - the `awk` command will check the last field of each line (`$NF`) and will print the original line (`$0`) only if the last field contained a number smaller than 2

```
$ sort -k1,1 -k2,2n testBed.bed | mergeBed -i stdin -n -nms | awk '$NF < 2 {print $0}'
chr1     35      40      region4 1
```

1.6.8 Why does the maximum value in the heatmap not equal the maximum value in the matrix?

Additional processing, such as outlier removal, is done on the matrix prior to plotting the heatmap. We've found this beneficial in most cases. You can override this by manually setting `--zMax` and/or `--zMin`, respectively.

1.6.9 The heatmap I generated looks very “coarse”, I would like a much more fine-grained image.

- decrease the *bin size* when generating the matrix using `computeMatrix`

In Galaxy:

- go to “advanced options” → “Length, in base pairs, of the non-overlapping *bin* for averaging the score over the regions length” → define a smaller value, e.g. 50 or 25 bp
 - make sure that you used a sufficiently small *bin* size when calculating the *bigWig* file, though (if generated with deepTools, you can check the option “bin size”)
-

1.6.10 How can I change the automatic labels of the clusters in a k-means clustered heatmap?

Each cluster is treated exactly the same way as different groups of regions. Therefore, you can use the same option to define the labels of the final heatmap:

In Galaxy: plotHeatmap -> “Advanced output options” -> “Labels for the regions plotted in the heatmap”.

If you indicated 2 clusters for k-means clustering, enter here: C1, C2, -> instead of the full default label (“cluster 1”), the heatmap will be labeled with the abbreviations.

Labels for the regions plotted in the heatmap

C1, C2

If more than one region is being plotted then a list of labels separated by comma is required. For example, label1, label2. (`--regionsLabel`)

don't let the default entry “genes” fool you – this is the place to indicate multiple labels if multiple groups of regions are going to be shown in the heatmap

In the command line, use the `--regionsLabel` option to define the customized names for the regions.

1.6.11 How can I manually specify several groups of regions (instead of clustering)?

Simply specify multiple BED files (e.g., genes.bed, exons.bed and introns.bed). This works both in Galaxy and on the command line.

1.6.12 What do I have to pay attention to when working with a draft version of a genome?

If your genome isn't included in our standard dataset then you'll need the following:

1. **Effective genome size** - this is mostly needed for `bamCoverage` and `bamCompare`, see [below](#) for details
 2. **Reference genome sequence in 2bit format** - this is needed for `computeGCBias`, see [2bit](#) for details
-

1.6.13 How do I calculate the effective genome size for an organism that's not in your list?

At the moment we do not provide a tool for this purpose, so you'll have to find a solution outside of deepTools for the time being.

The “real” effective genome size is the part of the genome that is *uniquely mappable*. This means that the value will depend on the genome properties (how many repetitive elements, quality of the assembly etc.) and the length of the sequenced reads as 100 million 36-bp-reads might cover less than 100 million 100-bp-reads.

We currently have these options for you:

1. Use an *GEM*
2. Use *faCount* (only if you let reads be aligned non-uniquely, too!)
3. Use *bamCoverage*
4. Use *genomeCoverageBed*

Use GEM

There is a tool that promises to calculate the mappability for any genome given the read length (k-mer length): *GEM-Mappability Calculator*. According to this [reply here](#), you can calculate the effective genome size after running this program by counting the numbers of “!” which stands for uniquely mappable regions.

Use faCount

If you are using bowtie2, which reports *multimappers* (i.e., *non-uniquely* mapped reads) as a default setting, you can use **faCount from UCSC tools** to report the total number of bases as well as the number of bases that are missing from the genome assembly indicated by ‘N’. The effective genome size would then be the total number of base pairs minus the total number of ‘N’. Here’s an example output of *faCount* on *D. melanogaster* genome version dm3:

\$ UCSCtools/faCount dm3.fa							
#seq	len	A	C	G	T	N	cpg
chr2L	23011544	6699731	4811687	4815192	6684734	200	926264
chr2LHet	368872	90881	58504	57899	90588	71000	10958
chr2R	21146708	6007371	4576037	4574750	5988450	100	917644
chr2RHet	3288761	828553	537840	529242	826306	566820	99227
chr3L	24543557	7113242	5153576	5141498	7135141	100	995078
chr3LHet	2555491	725986	473888	479000	737434	139183	89647
chr3R	27905053	7979156	5995211	5980227	7950459	0	1186894
chr3RHet	2517507	678829	447155	446597	691725	253201	84175
chr4	1351857	430227	238155	242039	441336	100	43274
chrU	10049037	2511952	1672330	1672987	2510979	1680789	335241
chrUextra	29004656	7732998	5109465	5084891	7614402	3462900	986216
chrX	22422827	6409325	4742952	4748415	6432035	90100	959534
chrXHet	204112	61961	40017	41813	60321	0	754
chrYHet	347038	74566	45769	47582	74889	104232	8441
chrM	19517	8152	2003	1479	7883	0	132
total	168736537	47352930	33904589	33863611	47246682	6368725	6650479

In this example: Total no. bp = 168,736,537 Total no. ‘N’ = 6,368,725

Warning: This method only works if multimappers are randomly assigned to their possible locations (in such cases the effective genome size is simply the number of non-N bases).

Use bamCoverage

If you have a sample where you expect the genome to be covered completely, e.g. from genome sequencing, a very trivial solution is to use *tools/bamCoverage* with a bin size of 1 bp and the *--outFileFormat* option set to

‘bedgraph’. You can then count the number of non-Zero bins (bases) which will indicate the mappable genome size for this specific sample.

Use genomeCoverageBed

genomeCoverageBed from the BEDtools suite can be used to calculate the number of bases in the genome for which 0 overlapping reads can be found. As described on the [BEDtools website](#) (go to genomeCov description), you need:

- a file with the chromosome sizes of your sample’s organism
- a position-sorted BAM file

```
$ bedtools genomecov -ibam sortedBAMfile.bam -g genome.size
```

1.6.14 Where can I download the 2bit genome files required for computeGCBias?

The 2bit files of most genomes can be found [here](#). Search for the .2bit ending. Otherwise, **fasta files can be converted to 2bit** using the UCSC program faToTwoBit (available for different platforms from [UCSC here](#)).

[deepTools Galaxy.](#) | [code @ github.](#)

1.7 Galaxy-related FAQ

- *I’ve reached my quota - what can I do to save some space?*
- *Copying from one history to another doesn’t work for me - the data set simply doesn’t show up in the target history!*
- *How can I use a published workflow?*
- *I would like to use one of your workflows - not in the deepTools Galaxy, but in the local Galaxy instance provided by my institute. Is that possible?*
- *plotProfile says that one option will only work if “computeMatrix was run with –missingDataAsZero”. How can I find out whether I ran computeMatrix that way?*
- *How can I have a look at the continuous read coverages from bigWig files? Which genome browser do you recommend?*
 - IGV (recommended)
 - UCSC
- *What’s the best way to integrate the deepTools results with other downstream analyses (outside of Galaxy)?*
- *How can I determine basic parameters of a BAM file, such as the number of reads, read length, duplication rate and average DNA fragment length?*

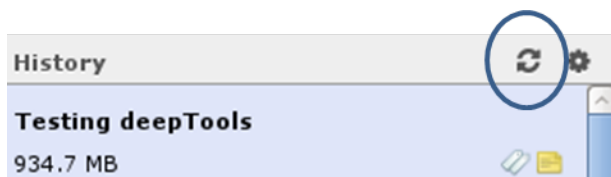
1.7.1 I’ve reached my quota - what can I do to save some space?

1. make sure that all the data sets you deleted are **permanently** eliminated from our disks: go to the history option button and select “Purge deleted data sets”, then hit the “refresh” button on top of your history panel
2. download all data sets for which you’ve completed the analysis, then remove the data sets (click on the “x” and then **make sure they’re purged** (see above)).

1.7.2 Copying from one history to another doesn't work for me - the data set simply doesn't show up in the target history!

Once you've copied a data set from one history to another, check two things:

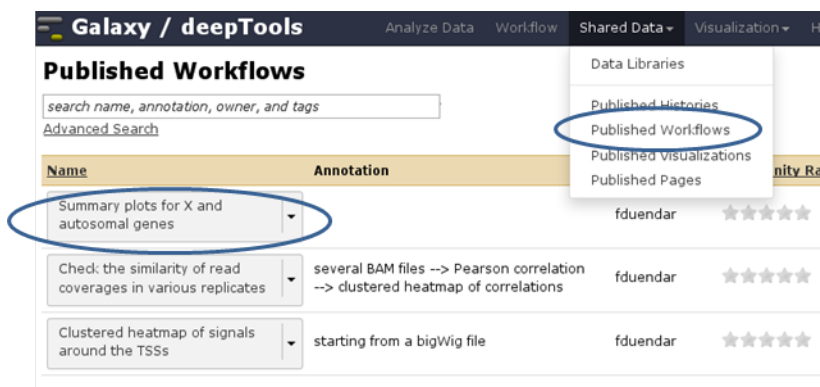
- do you see the destination history in your history panel, i.e. does the title of the current history panel match the name of the destination history you selected in the main frame?
- hit the refresh button



1.7.3 How can I use a published workflow?

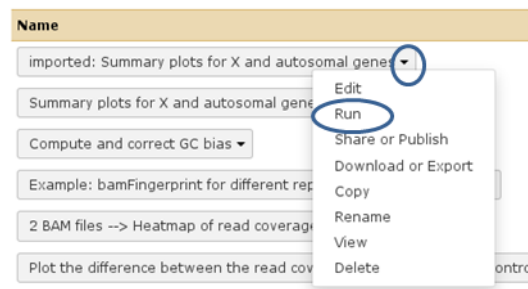
You **must register** if you want to use the workflows within [deepTools Galaxy](#). (“User” → “Register” - all you have to supply is an email address). Make sure to read the Terms of Use, though!

You can find workflows that are public or specifically shared with you by another user via “Shared Data” → “Published Workflows”. Click on the triangle next to the workflow you’re interested in and select “import”.



A green box should appear, there you select “start using this workflow”, which should lead you to your own workflow menu (that you can always access via the top menu “Workflow”). Here, you should now see a workflow labeled “imported: ...”. If you want to use the workflow right away, click on the triangle and select “Run”. The workflow should now be available within the Galaxy main data frame and should be waiting for your input.

Your workflows



1.7.4 I would like to use one of your workflows - not in the deepTools Galaxy, but in the local Galaxy instance provided by my institute. Is that possible?

Yes, it is possible. The only requirement is that your local Galaxy has a recent installation of deepTools.

Go to the workflows, click on the ones you're interested in and go to "Download". This will save the workflows into .ga files on your computer. Now go to your local Galaxy installation and login. Go to the workflow menu and select "import workflow" (top right hand corner of the page). Click on "Browse" and select the saved workflow. If you have the same tool versions installed in your local Galaxy, these workflows should work right away.

1.7.5 plotProfile says that one option will only work if "computeMatrix was run with `--missingDataAsZero`". How can I find out whether I ran computeMatrix that way?

Galaxy keeps track of everything you do. To see which options you chose to generate a specific data set, simply click on the "info" button.

Tool: computeMatrix		
Name:	computeMatrix on data 50, data 48, and data 11: Matrix	
Created:	Sun 31 Jan 2016 12:13:50 AM (UTC)	
Filesize:	5.3 MB	
Dbkey:	hg19	
Format:	deeptools_compute_matrix_archive	
Galaxy Tool ID:	testtoolshed.g2.bx.psu.edu/repos/bgruening/deeptools_compute_matrix/deeptools_compute_matrix/2.0.1.0.0	
Galaxy Tool Version:	2.0.1.0.0	
Tool Version:	computeMatrix 2.0.1	
Tool Standard Output:	stdout	
Tool Standard Error:	stderr	
Tool Exit Code:	0	
History		
Content API ID:	d2b0767063d74bd0	
Job API ID:	5bb18c11f5b70a41	
History API ID:	2a56795cad3c7db3	
UUID:	d8748d40-ecb7-4d56-82b4-168b11836001	
Input Parameter	Value	Note for rerun
Regions to plot	11: genes_chr19.bed	
Score file	48: log2ratio_H3K4me3-input.bw, 50: log2ratio_H3K27me3-input.bw	
computeMatrix has two main output options	scale-regions	
Distance in bases to which all regions are going to be fit	5000	
Set distance up- and downstream of the given regions	yes	
Distance upstream of the region start position	1000	
Distance downstream of the region end position	1000	
Show advanced output settings	yes	
Save the matrix of values underlying the heatmap	False	
Save the regions after skipping zeros or min/max threshold values	False	
Show advanced options	yes	
Length, in bases, of non-overlapping bins used for averaging the score over the regions length	50	
Sort regions	descending order	
Method used for sorting	mean	
Define the type of statistic that should be displayed.	mean	
Convert missing values to 0?	True	
Skip zeros	True	
Minimum threshold	None	
Maximum threshold	None	
Scaling factor	None	

Inheritance Chain

computeMatrix on data 50, data 48, and data 11: Matrix	files on which the current file is based (i.e., you can (and should!) safely rename your files)
--	---

79: computeMatrix on data 50, data 48, and data 11: Matrix
5.3 MB
format: deeptools_compute_matrix_archive, database: hg19

Found:
intervals: 12439
no groups found
/galaxy-central/tool_deps/numpy/1.9/iuc/package_pytho
RuntimeWarning: invalid value encountered in greater_equal
return umath.absolute(a) * self.tole

info needed for plotProfile or plotHeatmap

Compressed binary file

1.7.6 How can I have a look at the continuous read coverages from bigWig files? Which genome browser do you recommend?

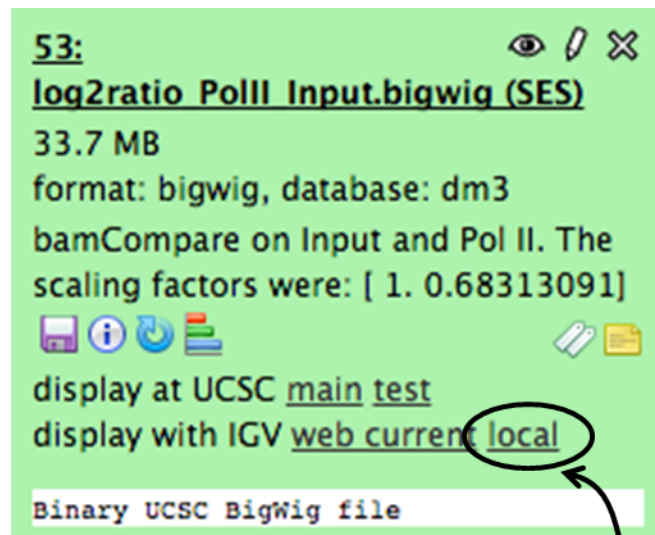
There are 2 popular genome browsers for visualizing continuous data: UCSC and IGV.

IGV (recommended)

We recommend downloading [IGV](#), which is free for academic use. IGV itself needs an up-to-date Java installation and a considerable amount of RAM. It's usage is rather intuitive and the display can be easily customized. In addition, you can download genome-wide annotation data that can be displayed together with your own data.

To display data in IGV, do the following:

1. Go to <http://www.broadinstitute.org/igv/>, register and download IGV
2. Unpack the IGV archive and change to the extracted IGV folder
3. Use the `igv.bat` (Windows), `igv.sh` (Linux) or `igv.command` (OSX) to start IGV (for more information please read the included `readme.txt` file or the IGV documentation).
4. Choose the genome version of the file(s) you would like to visualize (e.g. dm3) **THIS IS THE MOST IMPORTANT STEP!** IGV will not detect the genome version automatically, i.e. if you select mm9 but your file is based on human data, it will still be displayed without an error message (but with the wrong positions, obviously!)
5. Go to your deepTools Galaxy server (<http://deeptools.ie-freiburg.mpg.de/>) and navigate to your data set of choice
6. Click on your data set so that you see its details like in the screenshot below (**Keep in mind that not all datasets can be visualized in IGV or UCSC.** We recommend to use *bigWig* or *BED* files for visualization.)



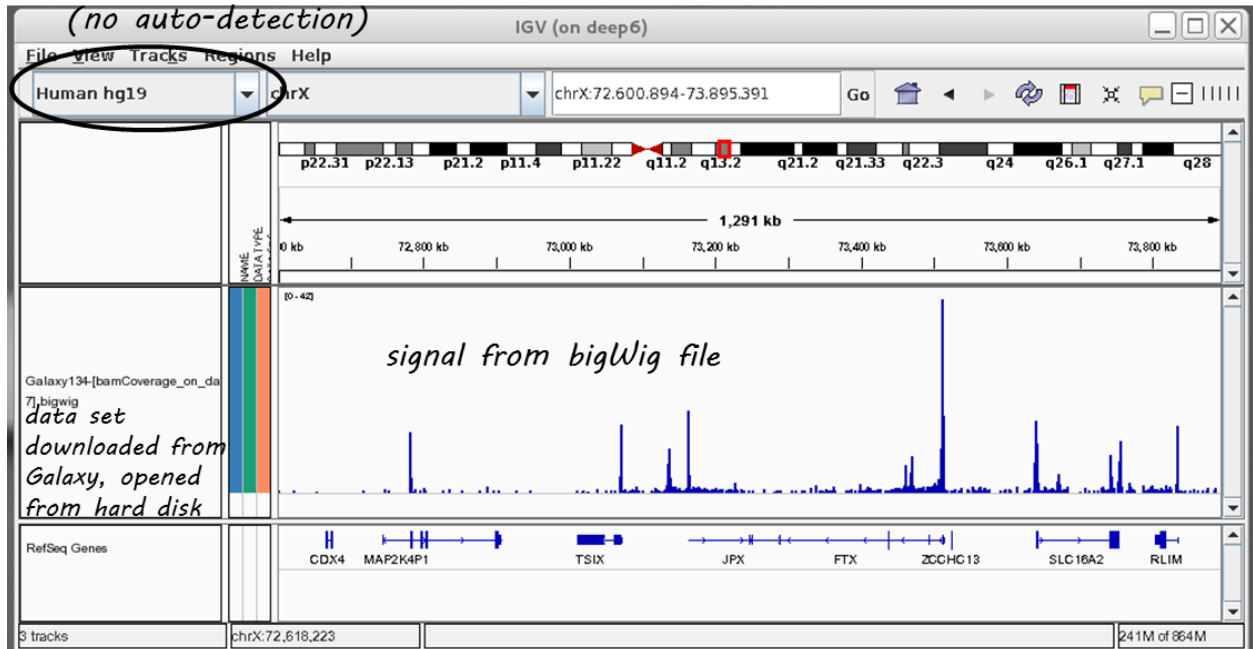
*IGV should already
be running on your
computer BEFORE
clicking here*

Now click on “display with IGV local” to visualize your data set in IGV that should already be running on your computer.

Note: “display with IGV Web current” can be used if you do not have an installed IGV. It will start an IGV web start version. **We do *not* recommend that option.**

Here's a screenshot of a typical bigWig file display:

*Genome MUST be set correctly
(no auto-detection)*



For more information, check out the [IGV documentation](#).

UCSC

There is a direct link from within deepTools Galaxy to stream a data set to UCSC. You can find it in the data set tiles: “display at UCSC”, like here:

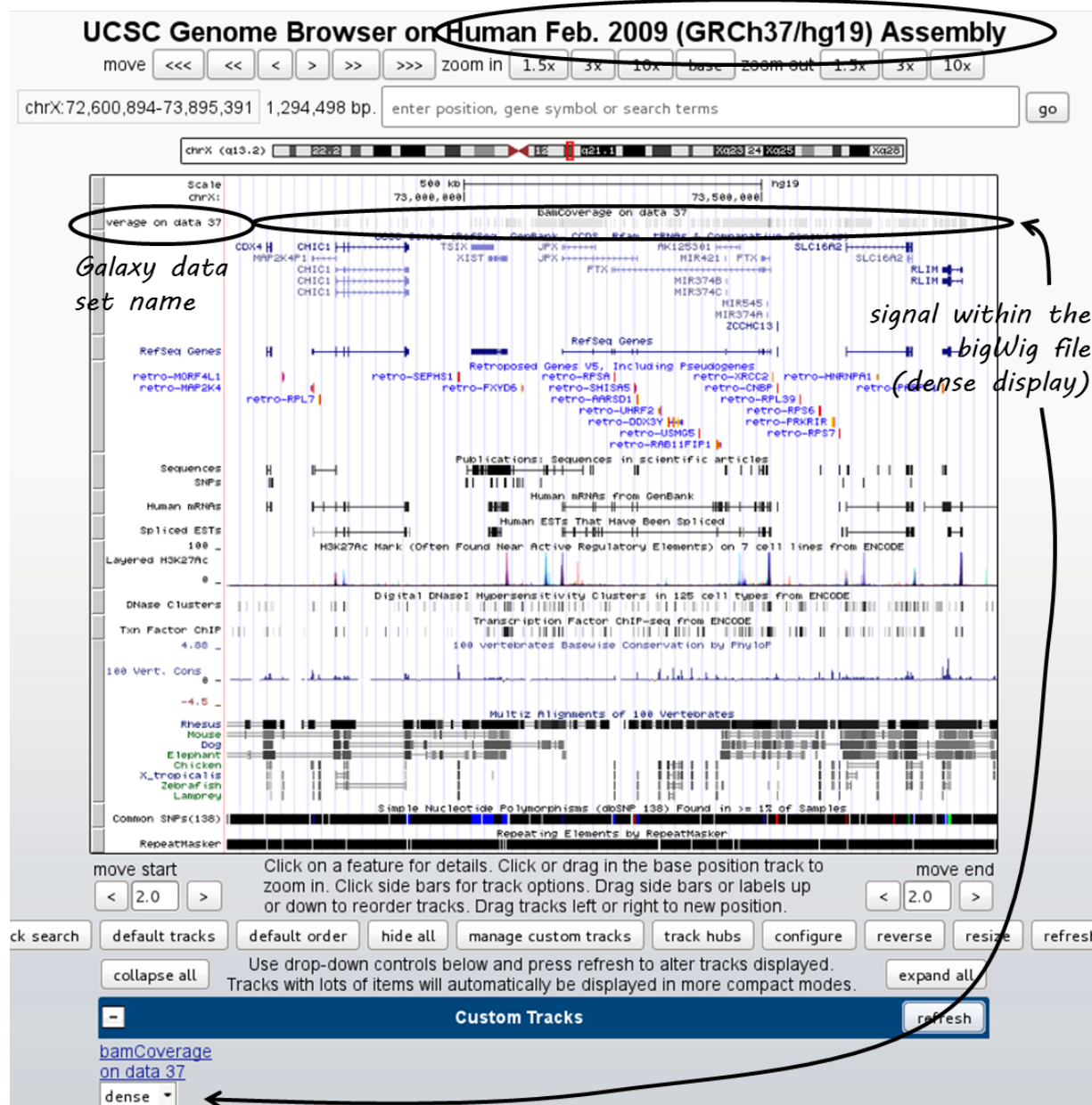
make sure this is correct (can be changed via the pencil icon)

134: bamCoverage on data 37
1.9 MB
format: bigwig, database **hg19**
display at UCSC **main test**
display in IGB [Local Web](#)
Binary UCSC BigWig file

click here to display the bigWig file within the UCSC Genome Browser

Click on “main” and the UCSC browser should open within a new window, displaying the data set that you chose. The default setting for bigWig files is the “dense” display that looks like a heatmap.

genome is determined automatically (based on the information contained in the Galaxy data set's metadata ("database"))



If you would like to display the continuous profile in a “valley-mountain” fashion like the one shown in the IGV screenshot, go to the drop-down menu underneath your custom track and choose “full”.

UCSC has large amounts of public data that you can display which you can find by scrolling down the page, beyond your custom track entry. For more information on how to use the UCSC Genome Browser, go [here](#).

Known issues with UCSC

- **chromosome naming:** UCSC expects chromosome names to be indicated in the format “chr”Number, e.g. chr1. If you mapped your reads to a non-UCSC-standard genome, chances are that chromosomes are labeled just with their number. bigWig files generated from these BAM files will not be recognized by UCSC, i.e. you will see

the data set name, but no signal.

- **no upload of bigWig files from your hard drive:** to minimize the computational strains, UCSC relies on streaming bigWig files (i.e. there's no need to load the entire file at once, the browser will always just load the data for the specific region a user is looking at).

1.7.7 What's the best way to integrate the deepTools results with other downstream analyses (outside of Galaxy)?

You can **save all the data tables** underlying every image produced by deepTools, i.e. if you would like to plot the average profiles in a different way, you could download the corresponding data (after ticking the relevant option under "advanced output options") and import them into R, Excel, GraphPadPrism etc.

The descriptions of the tools within Galaxy will also contain details on how to save the data and what sort of format to expect.

1.7.8 How can I determine basic parameters of a BAM file, such as the number of reads, read length, duplication rate and average DNA fragment length?

If you downloaded the *BAM* file from a public repository, chances are that those characteristics are in fact noted there.

If that's not the case, we recommend to have a look at the tool *FastQC*, which will return all of the above points (except the fragment size). The fragment size distribution can be obtained using the deepTools' tools/bamPEFragmentSize (since deepTools 2.0).

deepTools Galaxy. | code @ github.

1.8 Glossary of NGS terms

Like most specialized fields, next-generation sequencing has inspired many an acronyms. We are trying to keep track of those *Abbreviations* that we heavily use. Do make us aware if something is unclear: deeptools@googlegroups.com

- *Abbreviations*
- *NGS and generic terminology*
 - *bin*
 - *Input*
 - *read*
- *File Formats*
 - *2bit*
 - *BAM*
 - *BED*
 - *bedGraph*
 - *bigWig*
 - *FASTA*
 - *FASTQ*
 - *SAM*
 - * *SAM header section*
 - * *SAM alignment section*

1.8.1 Abbreviations

Reference genomes are usually referred to by their abbreviations, such as:

- hg19 = human genome, version 19
- mm9 = *Mus musculus* genome, version 9
- dm3 = *Drosophila melanogaster*, version 3
- ce10 = *Caenorhabditis elegans*, version 10

For a more comprehensive list of available reference genomes and their abbreviations, see the [UCSC data base](#).

Acronym	full phrase	Synonyms/Explanation
<ANYTHING>seq	sequencing	indicates that an experiment was completed by DNA sequencing using NGS
ChIP-seq	chromatin immunoprecipitation sequencing	NGS technique for detecting transcription factor binding sites and histone modifications (see entry <i>Input</i> for more information)
DNase	deoxyribonuclease I	DNase I digestion is used to determine active (“open”) chromatin regions
HTS	high-throughput sequencing	next-generation sequencing, massive parallel short read sequencing, deep sequencing
MNase	micrococcal nuclease	MNase digestion is used to determine sites with nucleosomes
NGS	next-generation sequencing	high-throughput (DNA) sequencing, massive parallel short read sequencing, deep sequencing
RPGC	reads per genomic content	normalize reads to 1x sequencing depth, sequencing depth is defined as: (mapped reads x fragment length) / effective genome size
RPKM	reads per kilobase per million reads	normalize read numbers: RPKM (per bin) = reads per bin / (mapped reads (in millions) x bin length (kb))

For a review of popular *-seq applications, see [Zentner and Henikoff](#).

1.8.2 NGS and generic terminology

The following are terms that may be new to some:

bin

- synonyms: window, region
- A ‘bin’ is a subset of a larger grouping. Many calculations are performed by first dividing the genome into small regions (bins), on which the calculations are actually performed.

Input

- Control experiment typically done for ChIP-seq experiments
- While ChIP-seq relies on antibodies to enrich for DNA fragments bound to a certain protein, the input sample should be processed exactly the same way, excluding the antibody. This allows one to account for biases introduced by sample handling and the general chromatin structure of the cells

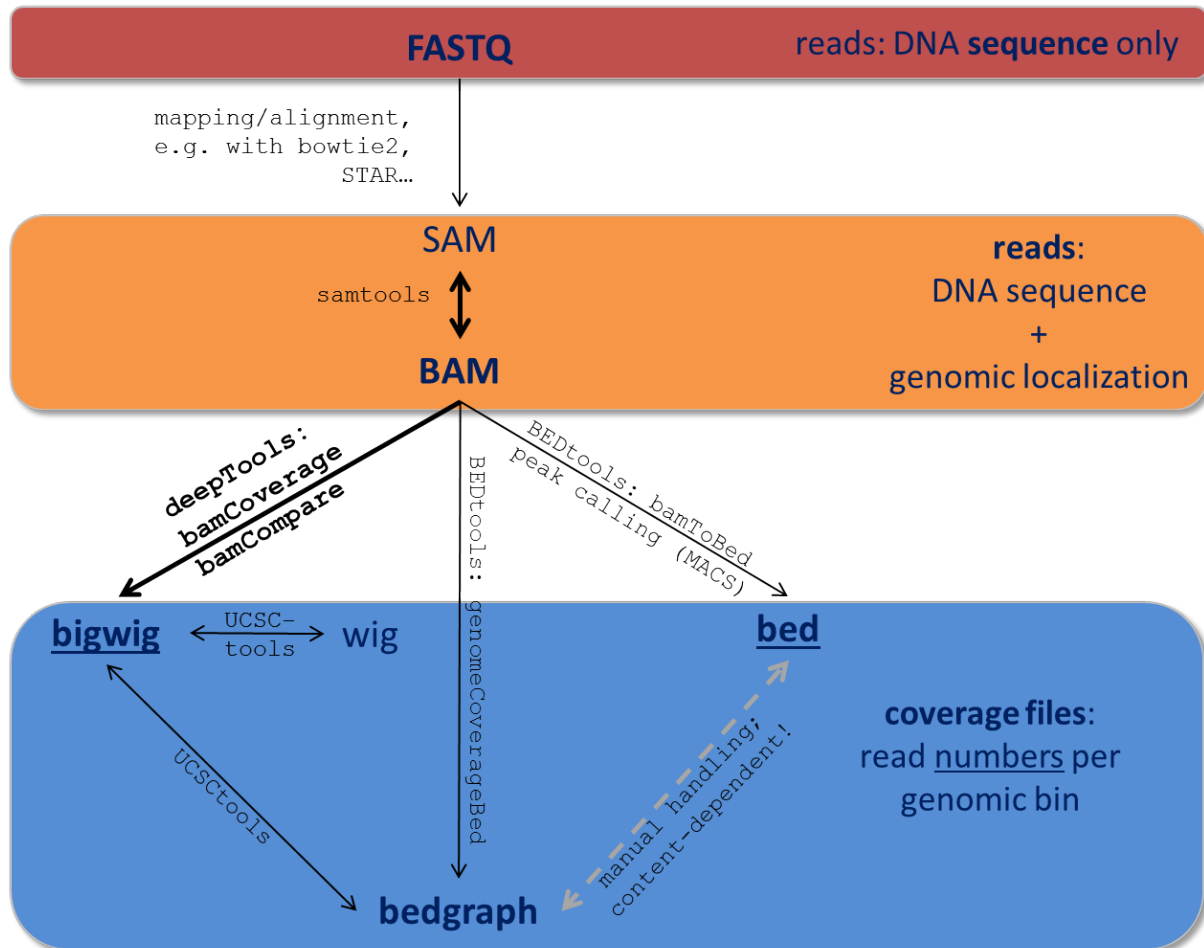
read

- synonym: tag
- This term refers to the piece of DNA that is sequenced (“read”) by the sequencers. We try to differentiate between “read” and “DNA fragment” as the fragments that are put into the sequencer tend to be in the range of 200-1000 bases, of which only the first 50 to 300 bases are typically sequenced. Most of the deepTools will not only take these reads into account, but also extend them to match the original DNA fragment size. (The original size will either be given by you or, if you used paired-end sequencing, be calculated from the distance between the two read mates).

1.8.3 File Formats

Data obtained from next-generation sequencing data must be processed several times. Most of the processing steps are aimed at extracting only that information needed for a specific down-stream analysis, with redundant entries often discarded. Therefore, **specific data formats are often associated with different steps of a data processing pipeline.**

Here, we just want to give very brief key descriptions of the file, for elaborate information we will link to external websites. Be aware, that the file name sorting here is alphabetical, not according to their usage within an analysis pipeline that is depicted here:



Follow the links for more information on the different tool collections mentioned in the figure:

[samtools](#) | [UCSCtools](#) | [BEDtools](#) |

2bit

- compressed, binary version of genome sequences that are often stored in [FASTA](#)
- most genomes in 2bit format can be found at [UCSC](#)
- [FASTA](#) files can be converted to 2bit using the UCSC program *faToTwoBit*, which is available for different platforms at [UCSC](#)
- more information can be found [here](#)

BAM

- typical file extension: `.bam`
- *binary* file format (complement to [SAM](#))
- contains information about sequenced reads (typically) *after alignment* to a reference genome
- **each line = 1 mapped read, with information about:**
 - its mapping quality (how likelihood that the reported alignment is correct)
 - its sequencing quality (the probability that each base is correct)
 - its sequence
 - its location in the genome
 - etc.
- highly recommended format for storing data
- to make a BAM file human-readable, one can, for example, use the program *samtools view*
- for more information, see below for the definition of [SAM](#) files

BED

- typical file extension: `.bed`
- text file
- used for genomic intervals, e.g. genes, peak regions etc.
- the format can be found at [UCSC](#)
- for deepTools, the first 3 columns are important: chromosome, start position of the region, end position of the genome
- do not confuse it with the *bedGraph* format (although they are related)
- example lines from a BED file of mouse genes (note that the start position is 0-based, the end-position 1-based, following UCSC conventions for BED files):

chr1	3204562	3661579	NM_001011874	Xkr4	-
chr1	4481008	4486494	NM_011441	Sox17	-
chr1	4763278	4775807	NM_001177658	Mrpl15	-
chr1	4797973	4836816	NM_008866	Lypla1	+

bedGraph

- typical file extension: `.bg`, `.bedGraph`
- text file
- similar to BED file (not the same!), it can *only* contain 4 columns and the 4th column *must* be a score
- again, read the [UCSC description](#) for more details
- 4 example lines from a bedGraph file (like BED files following the UCSC convention, the start position is 0-based, the end-position 1-based in bedGraph files):

```
chr1 10 20 1.5
chr1 20 30 1.7
chr1 30 40 2.0
chr1 40 50 1.8
```

bigWig

- typical file extension: `.bw`, `.bigwig`
- *binary* version of a *bedGraph* or wig file
- contains coordinates for an interval and an associated score
- the score can be anything, e.g. an average read coverage
- [UCSC description](#) for more details

FASTA

- typical file extension: `.fasta`
- text file, often gzipped (`.fasta.gz`)
- very simple format for **DNA/RNA** or **protein** sequences, this can be anything from small pieces of DNA or proteins to an entire genome (most likely, you will get the genome sequence of your organism of interest in fasta format)
- see the [2bit](#) file format entry for a compressed alternative
- example from [wikipedia](#) showing exactly one sequence:

```
>gi|5524211|gb|AAD44166.1| cytochrome b [Elephas maximus maximus]
LCLYTHIGRNIYGSYLYSETWNTGIMLLITMATAFMGYVLPWGQMSFWGATVITNLFSAIPYIGTNLV
EWIWGGFSVDKATLNRFFAFHFILPFTMVALAGVHLTFLHETGSNNPLGLTSDSDKIPFHPYYTIKDFLG
LLLLILLLLALLSPDMLGDPDNHMPADPLNTPLHIKPEWYFLFAYAILRSVPNKLGGVLALFLSIVIL
GLMPFLHTSKHRSMMLRPLSQALFWTLTMDLLTLTWIGSQPVEYPYTIIGQMASILYFSIILAFLPIAGX
IENY
```

FASTQ

- typical file extension: `.fastq`, `.fq`
- text file, often gzipped (\rightarrow `.fastq.gz`)
- **contains raw read information – 4 lines per read:**
 - read ID

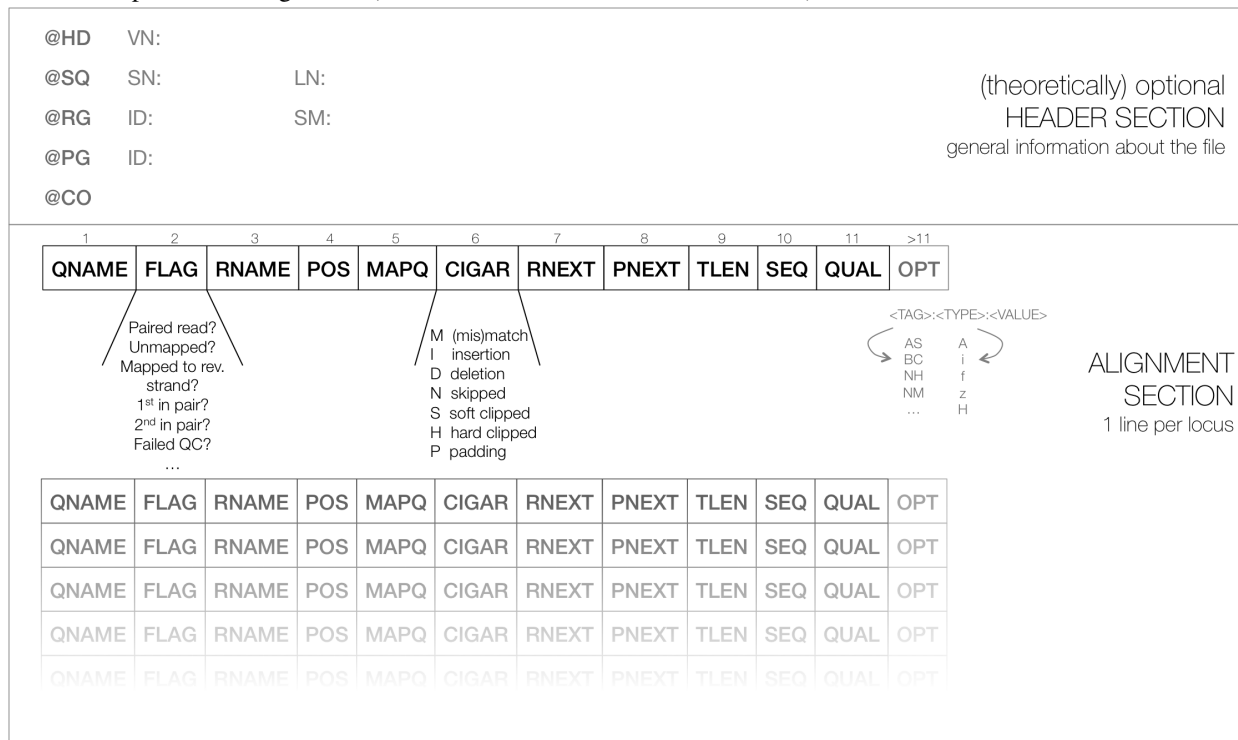
- base calls
- additional information or empty line
- sequencing quality measures - 1 per base call
- note that there is no information about where in the genome the read originated from
- example from the [wikipedia page](#), which contains further information:

```
@read001
GATTTCGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTGTCAACTCACAGTTT      # read sequence
+
!' '*((( (***+)) %%%++) (%%%) .1***-+* ' ')) **55CCF>>>>>CCCCCCC65      # ASCII-encoded quality scores
```

- if you need to find out what type of ASCII-encoding your .fastq file contains, you can simply run [FastQC](#) – its summary file will tell you

SAM

- typical file extension: .sam
- usually the result of an alignment of sequenced reads to a reference genome
- contains a short header section (entries are marked by @ signs) and an alignment section where each line corresponds to a single read (thus, there can be millions of these lines)



SAM header section

- tab-delimited lines, beginning with @, followed by tag:value pairs
- tag = two-letter string that defines the content and the format of value

SAM alignment section

- each line contains information about its mapping quality, its sequence, its location in the genome etc.

```
r001 163 chr1 7 30 8M2I4M1D3M = 37 39 TTAGATAAAGGATACTG *
r002 0 chr1 9 30 3S6M1P1I4M * 0 0 AAAAGATAAGGATA *
```

- the **flag in the second field** contains the answer to several yes/no assessments that are encoded in a single number
- for more details on the flag, see [this thorough explanation](#) or [this more technical explanation](#)
- the **CIGAR string in the 6th field** represents the types of operations that were needed in order to align the read to the specific genome location:
 - insertion
 - deletion (small deletions denoted with *D*, bigger deletions, e.g., for spliced reads, denoted with *N*)
 - clipping (deletion at the ends of a read)

Warning: Although the SAM/BAM format is rather meticulously defined and documented, whether an alignment program will produce a SAM/BAM file that adheres to these principles is completely up to the programmer. The mapping score, CIGAR string, and particularly, **all optional flags** (fields >11) are often **very differently defined depending on the program**. If you plan on filtering your data based on any of these criteria, make sure you know exactly how these entries were calculated and set!

[deepTools Galaxy.](#) | [code @ github.](#)

1.9 deepTools API

deepTools consists of several command line and Galaxy wrappers for summarizing the information of Next Generation Sequencing data that can be mapped to a reference genome. Through the API, the engine powering the deepTools commands can be used for other purposes as well.

Our [deepTools API example](#) explains step-by-step how to make use of some deepTools modules to achieve analyses outside the scope of the deepTools suite such as counting reads for certain genome regions and computing the FRiP score.

1.9.1 deepTools API example

The following is a short overview of the most useful methods and classes from deepTools. Complete information can be found in the following links: [genindex](#) and [modindex](#)

Finding read coverage over a region

With deepTools, the read coverage over multiple genomic regions and multiple files can be computed quite quickly using multiple processors. First, we start with a simple example that is later expanded upon to demonstrate the use of multiple processors. In this example we compute the coverage of reads over a small region for bins of 50bp. For this we need the `deeptools.countReadsPerBin` class.

```
import deeptools.countReadsPerBin
```

We also need a BAM file containing the aligned reads. The BAM file must be indexed to allow quick access to reads falling into the regions of interest.

```
bam_file = "file.bam"
```

Now, the `countReadsPerBin` object can be initialized. The first argument to the constructor is a list of BAM files, which in this case is just one file. We are going to use a `binLength` of 50 bases, with subsequent bins adjacent (i.e., the `stepSize` between bins is also 50 bases). Overlapping bin coverages can be used by setting a `stepSize` smaller than `binLength`.

```
cr = countReadsPerBin.CountReadsPerBin([bam_file], binLength=50, stepSize=50)
```

Now, we can compute the coverage over a region in chromosome 2 from position 0 to 1000.

```
cr.count_reads_in_region('chr2L', 0, 1000)
```

```
array([[ 2.],
       [ 3.],
       [ 1.],
       [ 2.],
       [ 3.],
       [ 2.],
       [ 4.],
       [ 3.],
       [ 2.],
       [ 3.],
       [ 4.],
       [ 6.],
       [ 4.],
       [ 2.],
       [ 2.],
       [ 1.]])
```

The result is a numpy array with one row per bin and one column per bam file. Since only one BAM file was used, there is only one column.

Filtering reads

If reads should be filtered, the relevant options simply need to be passed to the constructor. In the following code, the reads are filtered such that only those with a mapping quality of at least 20 and not aligned to the reverse strand are kept (`samFlag_exclude=16`, where 16 is the value for reverse reads, see the [SAM Flag Calculator](<http://broadinstitute.github.io/picard/explain-flags.html>) for more info). Furthermore, duplicated reads are ignored.

```
cr = countReadsPerBin.CountReadsPerBin([bam_file], binLength=50, stepSize=50,
                                       minMappingQuality=20,
                                       samFlag_exclude=16,
                                       ignoreDuplicates=True)
cr.count_reads_in_region('chr2L', 1000000, 1001000)
```

```
array([[ 1.],
       [ 1.],
       [ 0.],
       [ 0.],
       [ 0.],
       [ 0.],
       [ 2.]])
```

```
[ 3.],
[ 1.],
[ 0.],
[ 1.],
[ 2.],
[ 0.],
[ 0.],
[ 1.],
[ 2.],
[ 1.],
[ 0.],
[ 0.],
[ 0.]])
```

Sampling the genome

Instead of adjacent bins, as in the previous cases, a genome can simply be sampled. This is useful to estimate some values, like depth of sequencing, without having to look at the complete genome. In the following example, 10,000 positions of size 1 base are going to be queried from three bam files to compute the average depth of sequencing. For this, we set the *numberOfSamples* parameter in the object constructor. The *skipZeros* parameter is added to exclude regions lacking reads in all BAM files. The *run()* method is used instead of *count_reads_in_region*.

```
cr = countReadsPerBin.CountReadsPerBin([bam_file1, bam_file2, bam_file3],
                                       binLength=1, numberOfSamples=10000,
                                       numberOfProcessors=10,
                                       skipZeros=True)

sequencing_depth = cr.run()
print sequencing_depth.mean(axis=0)
```

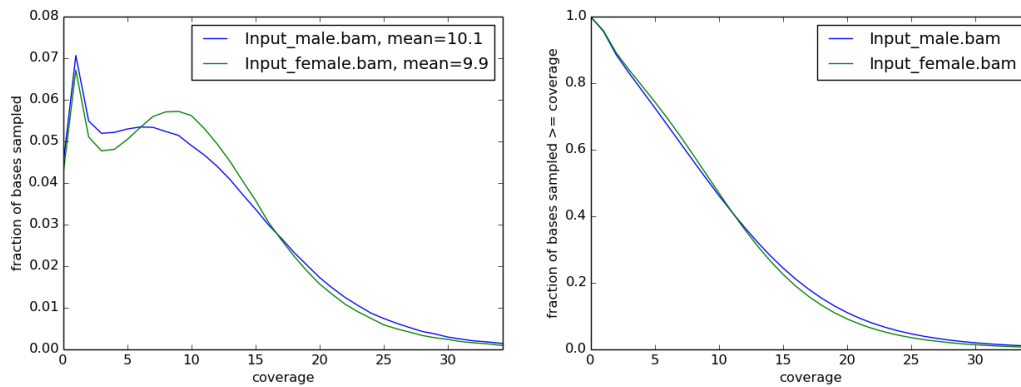
```
[ 1.98923924  2.43743744 22.90102603]
```

The *run()* method splits the computation over 10 processors and collates the results. When the parameter *numberOfSamples* is used, the regions selected for the computation of the coverage are not random. Instead, the genome is split into ‘number-of-samples’ equal parts and the start of each part is queried for its coverage. You can also compute coverage over selected regions by inputting a BED file.

Now it is possible to make some diagnostic plots from the results:

```
fig, axs = plt.subplots(1, 2, figsize=(15,5))
# plot coverage
for col in res.T:
    axs[0].plot(np.bincount(col.astype(int)).astype(float)/total_sites)
    csum = np.bincount(col.astype(int))[:, :-1].cumsum()
    axs[1].plot(csum.astype(float)[:, :-1] / csum.max())
axs[0].set_xlabel('coverage')
axs[0].set_ylabel('fraction of bases sampled')
# plot cumulative coverage

axs[1].set_xlabel('coverage')
axs[1].set_ylabel('fraction of bases sampled >= coverage')
```



Computing the FRiP score

The FRiP score is defined as the fraction of reads that fall into a peak and is often used as a measure of ChIP-seq quality. For this example, we need a BED file containing the peak regions. Such files are usually computed using a peak caller. Also, two bam files are going to be used, corresponding to two biological replicates.

```
bed_file = open("peaks.bed", 'r')
cr = countReadsPerBin.CountReadsPerBin([bam_file1, bam_file2],
                                       bedFile=bed_file,
                                       numberOfProcessors=10)
reads_at_peaks = cr.run()
print reads_at_peaks
```

```
array([[ 322.,  248.],
       [ 231.,  182.],
       [ 112.,  422.],
       ...,
       [ 120.,   76.],
       [ 235.,  341.],
       [ 246.,  265.]])
```

The result is a numpy array with a row for each peak region and a column for each BAM file.

```
reads_at_peaks.shape
```

```
(6295, 2)
```

Now, the total number of reads per peaks per bam file is computed:

```
total = reads_at_peaks.sum(axis=0)
```

Next, we need to find the total number of mapped reads in each of the bam files. For this we use the pysam module.

```
import pysam
bam1 = pysam.AlignmentFile(bam_file1)
bam2 = pysam.AlignmentFile(bam_file2)
```

Now, *bam1.mapped* and *bam2.mapped* contain the total number of mapped reads in each of the bam files, respectively.

Finally, we can compute the FRiP score:

```
frip1 = float(total[0]) / bam1.mapped
frip2 = float(total[1]) / bam2.mapped
print frip1, frip2
```

```
0.170030741997, 0.216740390353
```

Using mapReduce to sample paired-end fragment lengths

deepTools internally uses a map-reduce strategy, in which a computation is split into smaller parts that are sent to different processors. The output from the different processors is subsequently collated. The following example is based on the code available for *bamPEFragmentSize.py*

Here, we retrieve the reads from a BAM file and collect the fragment length. Reads are retrieved using pysam, and the *read* object returned contains the *template_length* attribute, which is the number of bases from the leftmost to the rightmost mapped base in the read pair.

First, we will create a function that can collect fragment lengths over a genomic position from a BAM file. As we will later call this function using mapReduce, the function accepts only one argument, namely a tuple with the parameters: chromosome name, start position, end position, and BAM file name.

```
import pysam
import numpy as np
def get_fragment_length(args):
    chrom, start, end, bam_file_name = args
    bam = pysam.AlignmentFile(bam_file_name)
    f_lens_list = []
    for fetch_start in range(start, end, 1e6):
        # simply get the reads over a region of 10000 bases
        fetch_end = min(end, start + 10000)

        f_lens_list.append(np.array([abs(read.template_length)
                                     for read in bam.fetch(chrom, fetch_start, fetch_end)
                                     if read.is_proper_pair and read.is_read1]))

    # concatenate all results
    return np.concatenate(fragment_lengths)
```

Now, we can use *mapReduce* to call this function and compute fragment lengths over the whole genome. mapReduce needs to know the chromosome sizes, which can be easily retrieved from the BAM file. Furthermore, it needs to know the size of the region(s) sent to each processor. For this example, a region of 10 million bases is sent to each processor using the *genomeChunkLength* parameter. In other words, each processor executes the same *get_fragment_length* function to collect data over different 10 million base regions. The arguments to mapReduce are the list of arguments sent to the function, besides the first obligatory three (chrom start, end). In this case only one extra argument is passed to the function, the BAM file name. The next two positional arguments are the name of the function to call (*get_fragment_length*) and the chromosome sizes.

```
import deeptools.mapReduce
bam = pysam.AlignmentFile(bamFile)
chroms_sizes = zip(bam.references, bam.lengths)

result = mapReduce.mapReduce((bam_file_name, ),
                             get_fragment_length
                             chrom_sizes,
                             genomeChunkLength=10000000,
                             numberOfProcessors=20,
                             verbose=True)

fragment_lengths = np.concatenate(result)

print "mean fragment length {}".format(fragment_lengths.mean())
print "median fragment length {}".format(np.median(fragment_lengths))
```

0.170030741997, 0.216740390353

Indices and tables

- [genindex](#)
- [modindex](#)
- [search](#)

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1.9.2 deeptools package modules

deeptools.SES_scaleFactor module

class `deeptools.SES_scaleFactor.Tester`
 Bases: `object`

`deeptools.SES_scaleFactor.estimateScaleFactor` (*bamFilesList*, *binLength*, *numberOfSamples*, *normalizationLength*, *avg_method='median'*, *numberOfProcessors=1*, *verbose=False*, *chrsToSkip=[]*)

Subdivides the genome into chunks to be analyzed in parallel using several processors. The code handles the creation of workers that compute fragment counts (coverage) for different regions and then collect and integrates the results.

Parameters `bamFilesList` : list

list of bam files to normalize

binLength : int

the window size in bp, where reads are going to be counted.

numberOfSamples : int

number of sites to sample from the genome. For more info see the documentation of the `CountReadsPerBin` class

normalizationLength : int

length, in bp, to normalize the data. For a value of 1, on average 1 read per base pair is found

avg_method : str

defines how the different values are to be summarized. The options are 'mean' and 'median'

chrsToSkip : list

name of the chromosomes to be excluded from the scale estimation. Usually the chrX is included.

Returns dict

Dictionary with the following keys:: 'size_factors' 'size_factors_based_on_mapped_reads' 'size_factors_SES' 'size_factors_based_on_mean' 'size_factors_based_on_median' 'mean' 'meanSES' 'median' 'reads_per_bin' 'std' 'sites_sampled'

Examples

```
>>> test = Tester()
>>> bin_length = 50
>>> num_samples = 4
>>> _dict = estimateScaleFactor([test.bamFile1, test.bamFile2], bin_length, num_samples, 1)
>>> _dict['size_factors']
array([ 1. ,  0.5])
>>> _dict['size_factors_based_on_mean']
array([ 1. ,  0.5])
```

deeptools.bamHandler module

`deeptools.bamHandler.openBam(bamFile)`

deeptools.correctReadCounts module

`deeptools.correctReadCounts.computeCorrectedReadcounts(tileCoverage, args)`

This function is called by the writeBedGraph workers for every tile in the genome that is considered

It computes a pvalue based on an expected lambda coming from the correction of treatment when the input is considered.

`deeptools.correctReadCounts.computeLambda(tileCoverage, args)`

This function is called by the writeBedGraph workers for every tile in the genome that is considered

`deeptools.correctReadCounts.computePvalue(tileCoverage, args)`

This function is called by the writeBedGraph workers for every tile in the genome that is considered

It computes a pvalue based on an expected lambda coming from the correction of treatment when the input is considered.

`deeptools.correctReadCounts.controlLambda(tileCoverage, args)`

`deeptools.correctReadCounts.correctReadCounts(bamFilesList, binLength, numberOfSamples, defaultFragmentLength, outFileName, outFileFormat, outFile-NameCorr=None, region=None, extend-PairedEnds=True, numberOfProcessors=1, Nsigmas=2, maxSignalRatio=10, verbose=False)`

deeptools.correlation module

`class deeptools.correlation.Correlation(matrix_file, corr_method=None, labels=None, remove_outliers=False, skip_zeros=False, log1p=False)`

class to work with matrices having sample data to compute correlations, plot them and make scatter plots

`compute_correlation()`

computes spearman or pearson correlation for the samples in the matrix

The matrix should contain the values of each sample per column that's why the transpose is used.

```
>>> matrix = np.array([[1, 2, 3, np.nan],
...                    [1, 2, 3, 4],
...                    [6, 4, 3, 1]]).T
>>> np.savez_compressed("/tmp/test_matrix.npz", matrix=matrix, labels=['a', 'b', 'c'])
```

```
>>> c = Correlation("/tmp/test_matrix.npz", corr_method='pearson')
```

the results should be as in R

```
>>> c.compute_correlation().filled(np.nan)
array([[ 1.          ,  1.          , -0.98198051],
       [ 1.          ,  1.          , -0.98198051],
       [-0.98198051, -0.98198051,  1.          ]])
>>> c.corr_method = 'spearman'
>>> c.corr_matrix = None
>>> c.compute_correlation()
array([[ 1.,  1., -1.],
       [ 1.,  1., -1.],
       [-1., -1.,  1.]])
```

static get_outlier_indices (*data*, *max_deviation=200*)

The method is based on the median absolute deviation. See Boris Iglewicz and David Hoaglin (1993), “Volume 16: How to Detect and Handle Outliers”, The ASQC Basic References in Quality Control: Statistical Techniques, Edward F. Mykytka, Ph.D., Editor.

returns the list, without the outliers

The *max_deviation=200* is like selecting a z-score larger than 200, just that it is based on the median and the median absolute deviation instead of the mean and the standard deviation.

load_matrix (*matrix_file*)

loads a matrix file saved using the numpy savez method. Two keys are expected: ‘matrix’ and ‘labels’. The matrix should contain one sample per row

plot_correlation (*plot_filename*, *plot_title=''*, *vmax=None*, *vmin=None*, *colormap='jet'*, *image_format=None*, *plot_numbers=False*)

plots a correlation using a symmetric heatmap

plot_pca (*plot_filename*, *plot_title=''*, *image_format=None*, *logIp=False*)

Plot the PCA of a matrix

plot_scatter (*plot_filename*, *plot_title=''*, *image_format=None*, *logIp=False*)

Plot the scatter plots of a matrix in which each row is a sample

remove_outliers (*verbose=True*)

get the outliers *per column* using the median absolute deviation method

Returns the filtered matrix

remove_rows_of_zeros ()

save_corr_matrix (*file_handle*)

saves the correlation matrix

deeptools.correlation_heatmap module

```
deeptools.correlation_heatmap.plot_correlation(corr_matrix, labels, plotFileName,
                                                vmax=None, vmin=None, colormap='jet',
                                                image_format=None,
                                                plot_numbers=False, plot_title='')
```

deeptools.countReadsPerBin module

```
class deeptools.countReadsPerBin.CountReadsPerBin (bamFilesList, binLength=50, num-
                                                    berOfSamples=None, numberOf-
                                                    Processors=1, verbose=False,
                                                    region=None, bedFile=None,
                                                    extendReads=False, minMap-
                                                    pingQuality=None, ignoreDupli-
                                                    cates=False, chrsToSkip=[], step-
                                                    Size=None, center_read=False,
                                                    samFlag_include=None, sam-
                                                    Flag_exclude=None, zerosTo-
                                                    Nans=False, smoothLength=0,
                                                    out_file_for_raw_data=None)
```

Bases: object

Collects coverage over multiple bam files using multiprocessing

This function collects read counts (coverage) from several bam files and returns an numpy array with the results. This class uses multiprocessing to compute the coverage.

Parameters **bamFilesList** : list

List containing the names of indexed bam files. E.g. ['file1.bam', 'file2.bam']

binLength : int

Length of the window/bin. This value is overruled by `bedFile` if present.

numberOfSamples : int

Total number of samples. The genome is divided into `numberOfSamples`, each with a window/bin length equal to `binLength`. This value is overruled by `stepSize` in case such value is present and by `bedFile` in which case the number of samples and bins are defined in the bed file

numberOfProcessors : int

Number of processors to use. Default is 4

verbose : bool

Output messages. Default: False

region : str

Region to limit the computation in the form chrom:start:end.

bedFile : file_handle

File handle of a bed file containing the regions for which to compute the coverage. This option overrules `binLength`, `numberOfSamples` and `stepSize`.

extendReads : bool, int

Whether coverage should be computed for the extended read length (i.e. the region covered by the two mates or the regions expected to be covered by single-reads). If the value is 'int', then this is interpreted as the fragment length to extend reads that are not paired. For Illumina reads, usual values are around 300. This value can be determined using the peak caller MACS2 or can be approximated by the fragment lengths computed when preparing the library for sequencing. If the value is of the variable is true and not value is given, the fragment size is sampled from the library but only if the library is paired-end. Default: False

minMappingQuality : int

Reads of a mapping quality less than the give value are not considered. Default: None

ignoreDuplicates : bool

Whether read duplicates (same start, end position. If paired-end, same start-end for mates) are to be excluded. Default: false

chrToSkip: list

List with names of chromosomes that do not want to be included in the coverage computation. This is useful to remove unwanted chromosomes (e.g. 'random' or 'Het').

stepSize : int

the positions for which the coverage is computed are defined as follows: `range(start, end, stepSize)`. Thus, a `stepSize` of 1, will compute the coverage at each base pair. If the `stepSize` is equal to the `binLength` then the coverage is computed for consecutive bins. If `stepSize` is smaller than the `binLength`, then the bins will overlap.

center_read : bool

Determines if reads should be centered with respect to the fragment length.

samFlag_include : int

Extracts only those reads having the SAM flag. For example, to get only reads that are the first mates a `samFlag` of 64 could be used. Similarly, the `samFlag_include` can be used to select only reads mapping on the reverse strand or to get only properly paired reads.

samFlag_exclude : int

Removes reads that match the SAM flag. For example to get all reads that map to the forward strand a `samFlag_exclude` 16 should be used. Which translates into exclude all reads that map to the reverse strand.

zerosToNans : bool

If true, zero values encountered are transformed to Nans. Default false.

out_file_for_raw_data : str

File name to save the raw counts computed

Returns numpy array

Each row correspond to each bin/bed region and each column correspond to each of the bamFiles.

Examples

The test data contains reads for 200 bp.

```
>>> test = Tester()
```

The transpose function is used to get a nicer looking output. The first line corresponds to the number of reads per bin in bam file 1

```
>>> c = CountReadsPerBin([test.bamFile1, test.bamFile2], 50, 4)
>>> np.transpose(c.run())
array([[ 0.,  0.,  1.,  1.],
       [ 0.,  1.,  1.,  2.]])
```

count_reads_in_region (*chrom, start, end, bed_regions_list=None*)

Counts the reads in each bam file at each 'stepSize' position within the interval (start, end) for a window or bin of size binLength.

The stepSize controls the distance between bins. For example, a step size of 20 and a bin size of 20 will create bins next to each other. If the step size is smaller than the bin size the bins will overlap.

If a list of bedRegions is given, then the number of reads that overlaps with each region is counted.

Parameters **chrom** : str

Chrom name

start : int

start coordinate

end : int

end coordinate

bed_regions_list: list

List of tuples of the form (chrom, start, end) corresponding to bed regions to be processed. If not bed file was passed to the object constructor then this list is empty.

Returns numpy array

The result is a numpy array that as rows each bin and as columns each bam file.

Examples

Initialize some useful values

```
>>> test = Tester()
>>> c = CountReadsPerBin([test.bamFile1, test.bamFile2], 25, 0, stepSize=50)
```

The transpose is used to get better looking numbers. The first line corresponds to the number of reads per bin in the first bamfile.

```
>>> _array, __ = c.count_reads_in_region(test.chrom, 0, 200)
>>> _array
array([[ 0.,  0.],
       [ 0.,  1.],
       [ 1.,  1.],
       [ 1.,  2.]])
```

getReadLength (*read*)

getSmoothRange (*tileIndex, tileSize, smoothRange, maxPosition*)

Given a tile index position and a tile size (length), return the a new indices over a larger range, called the smoothRange. This region is centered in the tileIndex an spans on both sizes to cover the smoothRange. The smoothRange is trimmed in case it is less than zero or greater than maxPosition

```
-----|=====|-----
          tileStart
          |-----|
```

```

| <-- smoothRange --> |
|
tileStart - (smoothRange-tileSize)/2

```

Test for a smooth range that spans 3 tiles.

Examples

```

>>> c = CountReadsPerBin([], 1, 1, 1, 0)
>>> c.getSmoothRange(5, 1, 3, 10)
(4, 7)

```

Test smooth range truncated on start.

```

>>> c.getSmoothRange(0, 10, 30, 200)
(0, 2)

```

Test smooth range truncated on start.

```

>>> c.getSmoothRange(1, 10, 30, 4)
(0, 3)

```

Test smooth range truncated on end.

```

>>> c.getSmoothRange(5, 1, 3, 5)
(4, 5)

```

Test smooth range not multiple of tileSize.

```

>>> c.getSmoothRange(5, 10, 24, 10)
(4, 6)

```

get_coverage_of_region(bamHandle, chrom, start, end, tileSize, fragmentFrom-
Read_func=None)

Returns a numpy array that corresponds to the number of reads that overlap with each tile.

```

>>> test = Tester()
>>> import pysam
>>> c = CountReadsPerBin([], stepSize=1, extendReads=300)

```

For this case the reads are length 36. The number of overlapping read fragments is 4 and 5 for the positions tested.

```

>>> c.get_coverage_of_region(pysam.AlignmentFile(test.bamFile_PE), 'chr2',
... 5000833, 5000835, 1)
array([ 4.,  5.])

```

In the following example a paired read is extended to the fragment length which is 100. The first mate starts at 5000000 and the second at 5000064. Each mate is extended to the fragment length *independently*. At position 500090-500100 one fragment of length 100 overlap, and after position 5000101 there should be zero reads.

```

>>> c.zerosToNans = True
>>> c.get_coverage_of_region(pysam.AlignmentFile(test.bamFile_PE), 'chr2', 5000090, 5000110,
array([ 1., nan])

```

In the following case the reads length is 50. Reads are not extended.

```
>>> c.extendReads=False
>>> c.get_coverage_of_region(pysam.AlignmentFile(test.bamFile2), '3R', 148, 154, 2)
array([ 1.,  2.,  2.])
```

get_fragment_from_read(read)

Get read start and end position of a read. If given, the reads are extended as follows: If reads are paired end, each read mate is extended to match the fragment length, otherwise, a default fragment length is used. If reads are split (give by the CIGAR string) then the multiple positions of the read are returned. When reads are extended the cigar information is skipped.

Parameters *read* : pysam read object

Returns list of tuples

[(fragment start, fragment end)]

```
>>> test = Tester()
```

```
>>> c = CountReadsPerBin([], 1, 1, 200, extendReads=True)
```

```
>>> c.defaultFragmentLength=100
```

```
>>> c.get_fragment_from_read(test.getRead("paired-forward"))
```

```
[(5000000, 5000100)]
```

```
>>> c.get_fragment_from_read(test.getRead("paired-reverse"))
```

```
[(5000000, 5000100)]
```

```
>>> c.defaultFragmentLength = 200
```

```
>>> c.get_fragment_from_read(test.getRead("single-forward"))
```

```
[(5001491, 5001691)]
```

```
>>> c.get_fragment_from_read(test.getRead("single-reverse"))
```

```
[(5001536, 5001736)]
```

```
>>> c.defaultFragmentLength = 'read length'
```

```
>>> c.get_fragment_from_read(test.getRead("single-forward"))
```

```
[(5001491, 5001527)]
```

```
>>> c.defaultFragmentLength = 'read length'
```

```
>>> c.extendReads = False
```

```
>>> c.get_fragment_from_read(test.getRead("paired-forward"))
```

[(5000000, 5000036)]

Tests for read centering.

```
>>> c = CountReadsPerBin([], 1, 1, 200, extendReads=True, center_read=True)
```

```
>>> c.defaultFragmentLength = 100
```

```
>>> c.get_fragment_from_read(test.getRead("paired-forward"))
```

[(5000032, 5000068)]

```
>>> c.defaultFragmentLength = 200
```

```
>>> c.get_fragment_from_read(test.getRead("single-reverse"))
```

[(5001618, 5001654)]

run()

class `deeptools.countReadsPerBin.Tester`

Bases: `object`

getRead (*readType*)

prepare arguments for test

`deeptools.countReadsPerBin.countReadsInRegions_wrapper` (*args*)

Passes the arguments to `countReadsInRegions_worker`. This is a step required given the constraints from the multiprocessing module. The *args* var, contains as first element the 'self' value from the `countReadsPerBin` object

`deeptools.countReadsPerBin.remove_row_of_zeros` (*matrix*)

deeptools.getFragmentAndReadSize module

`deeptools.getFragmentAndReadSize.getFragmentLength_worker` (*chrom, start, end, bam-File, distanceBetween-Bins*)

Queries the reads at the given region for the distance between reads and the read length

Parameters **chrom** : str

chromosome name

start : int

region start

end : int

region end

bamFile : str

BAM file name

distanceBetweenBins : int

the number of bases at the end of each bin to ignore

Returns `np.array`

an np.array, where first column is fragment length, the second is for read length

`deeptools.getFragmentAndReadSize.getFragmentLength_wrapper(args)`

`deeptools.getFragmentAndReadSize.get_read_and_fragment_length(bamFile, return_lengths=False, numberOfProcessors=None, verbose=False, binSize=50000, distanceBetweenBins=1000000)`

Estimates the fragment length and read length through sampling

Parameters `bamFile` : str

BAM file name

return_lengths : bool

numberOfProcessors : int

verbose : bool

binSize : int

distanceBetweenBins : int

Returns `d` : dict

tuple of two dictionaries, one for the fragment length and the other for the read length.
The dictionaries summarise the mean, median etc. values

deeptools.getRatio module

`deeptools.getRatio.compute_ratio(value1, value2, args)`

`deeptools.getRatio.getRatio(tileCoverage, args)`

The mapreduce method calls this function for each tile. The parameters (args) are fixed in the main method.

```
>>> funcArgs= {'valueType': 'ratio', 'scaleFactors': (1,1), 'pseudocount': 1}
>>> getRatio([9, 19], funcArgs)
0.5
>>> getRatio([0, 0], funcArgs)
1.0
>>> getRatio([np.nan, np.nan], funcArgs)
nan
>>> getRatio([np.nan, 1.0], funcArgs)
nan
>>> funcArgs['valueType'] = 'subtract'
>>> getRatio([20, 10], funcArgs)
10
>>> funcArgs['scaleFactors'] = (1, 0.5)
>>> getRatio([10, 20], funcArgs)
0.0
```

The reciprocal ratio is of a and b is: is a/b if a/b > 1 else -1* b/a >>> funcArgs['valueType'] = 'reciprocal_ratio'
>>> funcArgs['scaleFactors'] = (1, 1) >>> funcArgs['pseudocount'] = 0 >>> getRatio([2, 1], funcArgs) 2.0 >>>
getRatio([1, 2], funcArgs) -2.0 >>> getRatio([1, 1], funcArgs) 1.0

deeptools.getScorePerBigWigBin module

class deeptools.getScorePerBigWigBin.**Tester**

Bases: object

deeptools.getScorePerBigWigBin.**countFragmentsInRegions_worker** (*chrom, start, end, bigWigFiles, stepSize, binLength, save_data, bedRegions=None*)

returns the average score in each bigwig file at each ‘stepSize’ position within the interval start, end for a ‘binLength’ window. Because the idea is to get counts for window positions at different positions for sampling the bins are equally spaced and *not adjacent*.

If a list of bedRegions is given, then the number of reads that overlaps with each region is counted.

Test dataset with two samples covering 200 bp. >>> test = Tester()

Fragment coverage. >>> np.transpose(countFragmentsInRegions_worker(test.chrom, 0, 200, [test.bwFile1, test.bwFile2], 50, 25, False)[0]) array([[1., 1., 2., 2.],

[1., 1., 1., 3.]])

```
>>> np.transpose(countFragmentsInRegions_worker(test.chrom, 0, 200, [test.bwFile1, test.bwFile2],
array([[ 1.5],
[ 1.5]]))
```

BED regions: >>> bedRegions = [(test.chrom, 45, 55), (test.chrom, 95, 105), (test.chrom, 145, 155)]
>>> np.transpose(countFragmentsInRegions_worker(test.chrom, 0, 200,[test.bwFile1, test.bwFile2], 200, 200, False, ... bedRegions=bedRegions)[0]) array([[1. , 1.5, 2.],

[1. , 1. , 2.]])

deeptools.getScorePerBigWigBin.**countReadsInRegions_wrapper** (*args*)

deeptools.getScorePerBigWigBin.**getChromSizes** (*bigwigFilesList*)

Get chromosome sizes from bigWig file with pyBigWig

Test dataset with two samples covering 200 bp. >>> test = Tester()

Chromosome name(s) and size(s). >>> getChromSizes([test.bwFile1, test.bwFile2]) [(‘3R’, 200L)], set([]))

deeptools.getScorePerBigWigBin.**getScorePerBin** (*bigWigFiles, binLength, numberOfProcessors=1, verbose=False, region=None, bedFile=None, stepSize=None, chrsToSkip=[], out_file_for_raw_data=None*)

This function returns a matrix containing scores (median) for the coverage of fragments within a region. Each row corresponds to a sampled region. Likewise, each column corresponds to a bigwig file.

Test dataset with two samples covering 200 bp. >>> test = Tester() >>> np.transpose(getScorePerBin([test.bwFile1, test.bwFile2], 50, 3)) array([[1., 1., 2., 2.],

[1., 1., 1., 3.]])

deeptools.heatmapper module

deeptools.heatmapper.**compute_sub_matrix_wrapper** (*args*)

class `deeptools.heatmapper.heatmapper`

Bases: `object`

Class to handle the reading and plotting of matrices.

static `change_chrom_names(chrom)`

Changes UCSC chromosome names to ensembl chromosome names and vice versa.

computeMatrix (*score_file_list, regions_file, parameters, verbose=False*)

Splits into multiple cores the computation of the scores per bin for each region (defined by a hash '#' in the regions (BED/GFF) file.

static `compute_sub_matrix_worker(score_file_list, regions, parameters)`

Parameters `score_file_list` : list

List of strings. Contains the list of bam or bigwig files to be used.

regions : list of dictionaries

Each item in the list is a dictionary containing containing the fields: 'chrom', 'start', 'end', 'name' and 'strand.

parameters : dict

Contains values that specify the length of bins, the number of bp after a reference point etc.

Returns numpy matrix

A numpy matrix that contains per each row the values found per each of the regions given

static `coverage_from_array(valuesArray, zones, binSize, avgType)`

static `coverage_from_bam(bamfile, chrom, zones, binSize, avgType, verbose=True)`

currently this method is deactivated because is too slow. It is preferred to create a coverage bigwig file from the bam file and then run heatmapper.

static `coverage_from_big_wig(bigwig, chrom, zones, binSize, avgType, nansAsZeros=False, verbose=True)`

uses bigwig file reader from bx-python to query a region define by chrom and zones. The output is an array that contains the bigwig value per base pair. The summary over bins is done in a later step when `coverage_from_array` is called. This method is more reliable than quering the bins directly from the bigwig, which should be more efficient.

By default, any region, even if no chromosome match is found on the bigwig file, produces a result. In other words no regions are skipped.

zones: array as follows **zone0:** region before the region start,

zone1: the body of the region (not always present) zone2: the region from the end of the region downstream

each zone is a tuple containing start, end, and number of bins

This is useful if several matrices wants to be merged or if the sorted BED output of one `computeMatrix` operation needs to be used for other cases

get_individual_matrices (*matrix*)

In case multiple matrices are saved one after the other this method splits them appart. Returns a list containing the matrices

get_num_individual_matrix_cols()

returns the number of columns that each matrix should have. This is done because the final matrix that is plotted can be composed of smaller matrices that are merged one after the other.

static get_regions_and_groups (*regions_file*, *onlyMultiplesOf=1*, *default_group_name='genes'*,
verbose=None)

Reads a bed file. In case is hash sign '#' is found in the file, this is considered as a delimiter to split the heatmap into groups

Returns a list of regions with a label index appended to each and a list of labels

static matrix_avg (*matrix*, *avgType='mean'*)

matrix_from_dict (*matrixDict*, *regionsDict*, *parameters*)

static my_average (*valuesArray*, *avgType='mean'*)

computes the mean, median, etc but only for those values that are not Nan

read_matrix_file (*matrix_file*)

save_BED (*file_handle*)

save_matrix (*file_name*)

saves the data required to reconstruct the matrix the format is: A header containing the parameters used to create the matrix encoded as: @key:value key2:value2 etc... The rest of the file has the same first 5 columns of a BED file: chromosome name, start, end, name, score and strand, all separated by tabs. After the fifth column the matrix values are appended separated by tabs. Groups are separated by adding a line starting with a hash (#) and followed by the group name.

The file is gzipped.

save_matrix_values (*file_name*)

save_tabulated_values (*file_handle*, *reference_point_label='TSS'*, *start_label='TSS'*,
end_label='TES', *averagetype='mean'*)

Saves the values averaged by col using the avg_type given

Args: file_handle: file name to save the file reference_point_label: Name of the reference point label
start_label: Name of the star label end_label: Name of the end label averagetype: average type (e.g. mean, median, std)

deeptools.heatmapper_utilities module

deeptools.heatmapper_utilities.**getProfileTicks** (*hm*, *referencePointLabel*, *startLabel*,
endLabel)

returns the position and labelling of the xticks that correspond to the heatmap

deeptools.heatmapper_utilities.**plot_single** (*ax*, *ma*, *average_type*, *color*, *label*,
plot_type='simple')

Adds a line to the plot in the given ax using the specified method

Parameters *ax* : matplotlib axis

matplotlib axis

ma : numpy array

numpy array The data on this matrix is summarized according to the *average_type* argument.

average_type : str

string values are sum mean median min max std

color : str

a valid color: either a html color name, hex (e.g #002233), RGB + alpha tuple or list or RGB tuple or list

label : str

label

plot_type: str

type of plot. Either 'se' for standard error, 'std' for standard deviation, 'overlapped_lines' to plot each line of the matrix, fill to plot the area between the x axis and the value or None, just to plot the average line.

Returns ax

matplotlib axis

Examples

```
>>> import matplotlib.pyplot as plt
>>> fig = plt.figure()
>>> ax = fig.add_subplot(111)
>>> matrix = np.array([[1,2,3],
...                   [4,5,6],
...                   [7,8,9]])
>>> ax = plot_single(ax, matrix -2, 'mean', color=[0.6, 0.8, 0.9], label='fill light blue', plot_type='fill')
>>> ax = plot_single(ax, matrix, 'mean', color='blue', label='red')
>>> ax = plot_single(ax, matrix + 5, 'mean', color='red', label='red', plot_type='std')
>>> ax = plot_single(ax, matrix + 10, 'mean', color='#cccccc', label='gray se', plot_type='se')
>>> ax = plot_single(ax, matrix + 20, 'mean', color=(0.9, 0.5, 0.9), label='violet', plot_type='std')
>>> ax = plot_single(ax, matrix + 30, 'mean', color=(0.9, 0.5, 0.9, 0.5), label='violet with alp')
>>> leg = ax.legend()
>>> plt.savefig("/tmp/test.pdf")
>>> fig = plt.figure()
```

deeptools.mapReduce module

deeptools.mapReduce.**BED_to_interval_tree**(BED_file)

Creates an index of intervals, using an interval tree, for each BED entry

Parameters BED_file – file handler of a BED file

:return interval tree

deeptools.mapReduce.**getUserRegion**(chrom_sizes, region_string, max_chunk_size=1000000.0)

Verifies if a given region argument, given by the user is valid. The format of the region_string is chrom:start:end:tileSize where start, end and tileSize are optional.

Parameters

- **chrom_sizes** – dictionary of chromosome/scaffold size. Key=chromosome name
- **region_string** – a string of the form chr:start:end
- **max_chunk_size** – upper limit for the chunk size

Returns tuple chrom_size for the region start, region end, chunk size

```
#>>> data = getUserRegion({'chr2': 1000}, "chr1:10:10") #Traceback (most recent call last): # ... #NameError:
Unknown chromosome: chr1 #Known chromosomes are: ['chr2']
```

If the region end is bigger than the chromosome size, this value is used instead >>> getUserRegion({'chr2': 1000}, "chr2:10:1001") ([('chr2', 1000)], 10, 1000, 990)

Test chunk and regions size reduction to match tile size >>> getUserRegion({'chr2': 200000}, "chr2:10:123344:3") ([('chr2', 123344)], 9, 123345, 123336)

```
deeptools.mapReduce.mapReduce (staticArgs, func, chromSize, genomeChunkLength=None, re-
                                gion=None, bedFile=None, numberOfProcessors=4, ver-
                               bose=False, self_=None)
```

Split the genome into parts that are sent to workers using a defined number of procesors. Results are collected and returned.

For each genomic region the given 'func' is called using the following parameters:

chrom, start, end, staticArgs

The *arg* are static, *pickable* variables that need to be sent to workers.

The genome chunk length corresponds to a fraction of the genome, in bp, that is send to each of the workers for processing.

Depending on the type of process a larger or shorter regions may be preferred

Parameters

- **chromSize** – A list of duples containing the chromosome name and its length
- **region** – The format is chr:start:end:tileSize (see function getUserRegion)
- **staticArgs** – tuple of arguments that are sent to the given 'func'
- **func** – function to call. The function is called using the following parameters (chrom, start, end, staticArgs)
- **bedFile** – Is a bed file is given, the args to the func to be called are extended to include a list of bed defined regions.
- **self** – In case mapreduce should make a call to an object the self variable has to be passed.

deeptools.readBed module

```
class deeptools.readBed.BedInterval (field_data_dict, line)
```

Bases: object

simple place holder for the line data of a bed file

```
class deeptools.readBed.ReadBed (file_handle)
```

Bases: object

Reads a bed file. Based on the number of fields it tries to guess the type of bed file used. Current options are bed3, bed6 and bed12

Examples

```
bed = readBed(open("file.bed", 'r')) for interval in bed: ... print bed['start']
```

```
get_no_comment_line ()
```

Skips comment lines starting with '#' "track" or "browser" in the bed files

```
get_values_from_line (bed_line)
    Processes each bed line from a bed file and casts the values

guess_file_type (line_values)
    try to guess type of bed file by counting the fields

next ()

    Returns bedInterval object
```

deeptools.utilities module

```
deeptools.utilities.copyFileInMemory (filePath, suffix='')
    copies a file into the special /dev/shm device which moves the file into memory. This process speeds up the
    multiprocessor access to such files

deeptools.utilities.getCommonChrNames (bamFileHandlers, verbose=True)
    Compares the names and lengths of a list of bam file handlers. The input is list of pysam file handlers.

    The function returns a tuple containing the common chromosome names and the common chromosome lengths.

    Hopefully, only _random and chrM are not common.

deeptools.utilities.getGC_content (dnaString, as_fraction=True)

deeptools.utilities.getTempFileName (suffix='')
    returns a temporary file name. If the special /dev/shm device is available, the temporary file would be located in
    that folder. /dev/shm is a folder that resides in memory and which has much faster accession.

deeptools.utilities.tbitToBamChrName (tbitNames, bamNames)
    checks if the chromosome names from the two-bit and bam file coincide. In case they do not coincide, a fix is
    tried. If successful, then a mapping table is returned. tbitNames and bamNames should be lists

deeptools.utilities.which (program)
    method to identify if a program is on the user PATH variable. From:
    http://stackoverflow.com/questions/377017/test-if-executable-exists-in-python
```

deeptools.writeBedGraph module

```
class deeptools.writeBedGraph.WriteBedGraph (bamFilesList, binLength=50, numberOfSam-
    ples=None, numberOfProcessors=1, ver-
    bose=False, region=None, bedFile=None, ex-
    tendReads=False, minMappingQuality=None,
    ignoreDuplicates=False, chrsToSkip=[],
    stepSize=None, center_read=False, sam-
    Flag_include=None, samFlag_exclude=None,
    zerosToNans=False, smoothLength=0,
    out_file_for_raw_data=None)

    Bases: deeptools.countReadsPerBin.CountReadsPerBin

    Reads bam files coverages and writes a bedgraph or bigwig file

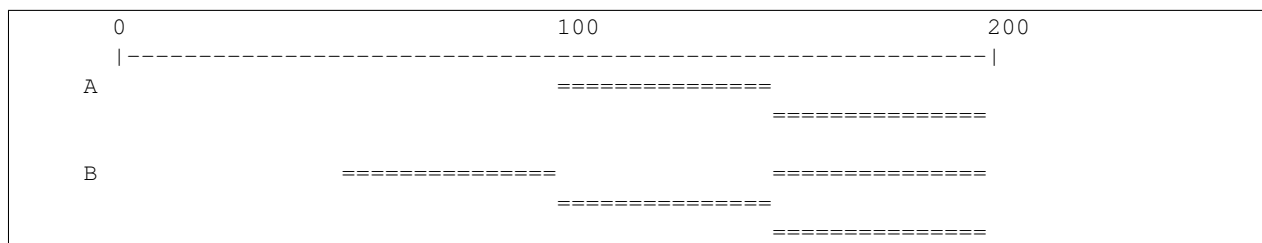
    Extends the CountReadsPerBin object such that the coverage of bam files is written to multiple bedgraph files at
    once.

    The bedgraph files are later merge into one and converted into a bigwig file if necessary.

    The constructor arguments are the same as for CountReadsPerBin. However, when calling the run method, the
    following parameters have to be passed
```

Examples

Given the following distribution of reads that cover 200 on a chromosome named '3R':



```
>>> import tempfile
>>> test_path = os.path.dirname(os.path.abspath(__file__)) + "/test/test_data/"
```

```
>>> outFile = tempfile.NamedTemporaryFile()
>>> bam_file = test_path + "testA.bam"
```

For the example a simple scaling function is going to be used. This function takes the coverage found at each region and multiplies it to the scaling factor. In this case the scaling factor is 1.5

```
>>> function_to_call = scaleCoverage
>>> funcArgs = {'scaleFactor': 1.5}
```

Restrict process to a region between positions 0 and 200 of chromosome 3R

```
>>> region = '3R:0:200'
```

Set up such that coverage is computed for consecutive bins of length 25 bp >>> bin_length = 25 >>> step_size = 25

```
>>> num_sample_sites = 0 #overruled by step_size
>>> c = WriteBedGraph([bam_file], binLength=bin_length, region=region, stepSize=step_size)
>>> c.run(function_to_call, funcArgs, outFile.name)
>>> open(outFile.name, 'r').readlines()
['3R\t0\t100\t0.00\n', '3R\t100\t200\t1.5\n']
>>> outFile.close()
```

run (*func_to_call*, *func_args*, *out_file_name*, *format*='bedgraph', *smoothLength*=0)

Given a list of bamfiles, a function and a function arguments, this method writes a bedgraph file (or bigwig) file for a partition of the genome into tiles of given size and a value for each tile that corresponds to the given function and that is related to the coverage underlying the tile.

Parameters **func_to_call** : str

function name to be called to convert the list of coverages computed for each bam file at each position into a single value. An example is a function that takes the ratio between the coverage of two bam files.

func_args : dict

dict of arguments to pass to *func*. E.g. {'scaleFactor':1.0}

out_file_name : str

name of the file to save the resulting data.

smoothLength : int

Distance in bp for smoothing the coverage per tile.

writeBedGraph_worker (*chrom, start, end, func_to_call, func_args, bed_regions_list=None*)

Writes a bedgraph based on the read coverage found on bamFiles

The given func is called to compute the desired bedgraph value using the funcArgs

Parameters **chrom** : str

Chrom name

start : int

start coordinate

end : int

end coordinate

func_to_call : str

function name to be called to convert the list of coverages computed for each bam file at each position into a single value. An example is a function that takes the ratio between the coverage of two bam files.

func_args : dict

dict of arguments to pass to *func*.

smoothLength : int

Distance in bp for smoothing the coverage per tile.

bed_regions_list: list

List of tuples of the form (chrom, start, end) corresponding to bed regions to be processed. If not bed file was passed to the object constructor then this list is empty.

Returns temporary file with the bedgraph results for the region queried.

Examples

```
>>> test_path = os.path.dirname(os.path.abspath(__file__)) + "/test/test_data/"
>>> bamFile1 = test_path + "testA.bam"
>>> bin_length = 50
>>> number_of_samples = 0 # overruled by step_size
>>> func_to_call = scaleCoverage
>>> funcArgs = {'scaleFactor': 1.0}
```

```
>>> c = WriteBedGraph([bamFile1], bin_length, number_of_samples, stepSize=50)
>>> tempFile = c.writeBedGraph_worker('3R', 0, 200, func_to_call, funcArgs)
>>> open(tempFile, 'r').readlines()
['3R\t0\t100\t0.00\n', '3R\t100\t200\t1.0\n']
>>> os.remove(tempFile)
```

deeptools.writeBedGraph.**bedGraphToBigWig** (*chromSizes, bedGraphPath, bigWigPath, sort=True*)

takes a bedgraph file, orders it and converts it to a bigwig file using pyBigWig.

deeptools.writeBedGraph.**getGenomeChunkLength** (*bamHandlers, tile_size*)

Tries to estimate the length of the genome sent to the workers based on the density of reads per bam file and the number of bam files.

The chunk length should be a multiple of the tileSize

`deeptools.writeBedGraph.ratio(tile_coverage, args)`

tileCoverage should be an list of two elements

`deeptools.writeBedGraph.scaleCoverage(tile_coverage, args)`

tileCoverage should be an list with only one element

`deeptools.writeBedGraph.writeBedGraph_wrapper(args)`

Passes the arguments to writeBedGraph_worker. This is a step required given the constraints from the multiprocessing module. The args var, contains as first element the 'self' value from the WriteBedGraph object

deeptools.writeBedGraph_bam_and_bw module

`deeptools.writeBedGraph_bam_and_bw.getCoverageFromBigwig(bigwigHandle, chrom, start, end, tileSize, missingDataAsZero=False)`

`deeptools.writeBedGraph_bam_and_bw.writeBedGraph(bamOrBwFileList, outputFile-Name, fragmentLength, func, funcArgs, tileSize=25, region=None, numberOfProcessors=None, format='bedgraph', extendPairedEnds=True, missingDataAsZero=False, smoothLength=0, fixed_step=False)`

Given a list of bamfiles, a function and a function arguments, this method writes a bedgraph file (or bigwig) file for a partition of the genome into tiles of given size and a value for each tile that corresponds to the given function and that is related to the coverage underlying the tile.

`deeptools.writeBedGraph_bam_and_bw.writeBedGraph_worker(chrom, start, end, tileSize, defaultFragmentLength, bamOrBwFileList, func, funcArgs, extendPairedEnds=True, smoothLength=0, missingDataAsZero=False, fixed_step=False)`

Writes a bedgraph having as base a number of bam files.

The given func is called to compute the desired bedgraph value using the funcArgs

tileSize

`deeptools.writeBedGraph_bam_and_bw.writeBedGraph_wrapper(args)`

Module contents

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Complete information can be found in the following links: [genindex](#) and [modindex](#)

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1.10 About

Please cite deepTools as follows: Fidel Ramírez, Friederike Dündar, Sarah Diehl, Björn A. Grünig, and Thomas Manke. [deepTools: a flexible platform for exploring deep-sequencing data](#). Nucl. Acids Res., 2014

doi:10.1093/nar/gku365



This tool suite is developed by the Bioinformatics Facility at the Max Planck Institute for Immunobiology and Epigenetics, Freiburg.

[deepTools Galaxy.](#) | [code @ github.](#)

While developing deepTools, we continuously strive to create software that fulfills the following criteria:

- **efficiently extract reads from BAM files** and perform various computations on them
- **turn BAM files of aligned reads into bigWig files** using different normalization strategies
- make use of **multiple processors** (speed!)
- generation of **highly customizable images** (change colours, size, labels, file format, etc.)
- enable **customized down-stream analyses**, meaning that every data set created can be stored by the user
- **modular approach** - compatibility, flexibility, scalability (i.e. we can add more and more modules and make use of established methods)

Tip: For support, questions, or feature requests contact: deeptools@googlegroups.com

Please cite deepTools as follows: Fidel Ramírez, Friederike Dündar, Sarah Diehl, Björn A. Grüning, and Thomas Manke. deepTools: a flexible platform for exploring deep-sequencing data. Nucl. Acids Res., 2014 doi:10.1093/nar/gku365



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